

Examination of Single Nucleotide Polymorphisms (SNPs) in Transient Receptor Potential (TRP) Ion Channels in Chronic Fatigue Syndrome Patients

Sonya M. Marshall-Gradisnik^{1,2}, Peter Smith², Ekuwa W. Brenu^{1,2}, Bernd Nilius³, Sandra B. Ramos^{1,2} and Donald R. Staines²

¹School of Medical Science, ²The National Centre for Neuroimmunology and Emerging Diseases, Menzies Health Institute Queensland, Griffith University, Gold Coast, Australia. ³Department of Cellular and Molecular Medicine, Laboratory of Ion Channel Research, KU Leuven, Belgium.

BACKGROUND: The transient receptor potential (TRP) superfamily in humans comprises 27 cation channels with permeability to monovalent and divalent cations. These channels are widely expressed within humans on cells and tissues and have significant sensory and regulatory roles on most physiological functions. Chronic fatigue syndrome (CFS) is an unexplained disorder with multiple physiological impairments.

OBJECTIVES: The purpose of this study was to determine the role of TRPs in CFS.

METHODS: The study comprised 115 CFS patients (age = 48.68 ± 1.06 years) and 90 nonfatigued controls (age = 46.48 ± 1.22 years). CFS patients were defined according to the 1994 Center for Disease Prevention and Control criteria for CFS. A total of 240 single nucleotide polymorphisms (SNPs) for 21 mammalian TRP ion channel genes (*TRPA1*, *TRPC1*, *TRPC2*, *TRPC3*, *TRPC4*, *TRPC6*, *TRPC7*, *TRPM1*, *TRPM2*, *TRPM3*, *TRPM4*, *TRPM5*, *TRPM6*, *TRPM7*, *TRPM8*, *TRPV1*, *TRPV2*, *TRPV3*, *TRPV4*, *TRPV5*, and *TRPV6*) were examined via the Agena Biosciences iPLEX Gold assay. Statistical analysis was performed using the PLINK analysis software.

RESULTS: Thirteen SNPs were significantly associated with CFS patients compared with the controls. Nine of these SNPs were associated with *TRPM3* (rs12682832; $P \leq 0.003$, rs11142508; $P < 0.004$, rs1160742; $P < 0.08$, rs4454352; $P \leq 0.013$, rs1328153; $P \leq 0.013$, rs3763619; $P \leq 0.014$, rs7865858; $P \leq 0.021$, rs1504401; $P \leq 0.041$, rs10115622; $P \leq 0.050$), while the remainder were associated with *TRPA1* (rs2383844; $P \leq 0.040$, rs4738202; $P \leq 0.018$) and *TRPC4* (rs6650469; $P \leq 0.016$, rs655207; $P \leq 0.018$).

CONCLUSION: The data from this pilot study suggest an association between TRP ion channels, predominantly TRPM3 and CFS. This and other TRPs identified may contribute to the etiology and pathomechanism of CFS.

KEYWORDS: chronic fatigue syndrome, transient receptor potential, single nucleotide polymorphisms

CITATION: Marshall-Gradisnik et al. Examination of Single Nucleotide Polymorphisms (SNPs) in Transient Receptor Potential (TRP) Ion Channels in Chronic Fatigue Syndrome Patients. *Immunology and Immunogenetics Insights* 2015:7 1–6 doi:10.4137/III.S25147.

RECEIVED: February 19, 2015. **RESUBMITTED:** April 15, 2015. **ACCEPTED FOR PUBLICATION:** April 15, 2015.

ACADEMIC EDITOR: Souvenir Tachado, editorial board member

TYPE: Original Research

FUNDING: This study was supported by funding from the Alison Hunter Memorial Foundation and Mason Foundation. The authors confirm that the funder had no influence over the study design, content of the article, or selection of this journal.

COMPETING INTERESTS: SMM-G, DRS and PS have a provisional patent pending on diagnostic methods. Other authors disclose no competing interests.

COPYRIGHT: © the authors, publisher and licensee Libertas Academica Limited. This is an open-access article distributed under the terms of the Creative Commons CC-BY-NC 3.0 License.

CORRESPONDENCE: s.marshall-gradisnik@griffith.edu.au

Paper subject to independent expert blind peer review by minimum of two reviewers. All editorial decisions made by independent academic editor. Upon submission manuscript was subject to anti-plagiarism scanning. Prior to publication all authors have given signed confirmation of agreement to article publication and compliance with all applicable ethical and legal requirements, including the accuracy of author and contributor information, disclosure of competing interests and funding sources, compliance with ethical requirements relating to human and animal study participants, and compliance with any copyright requirements of third parties. This journal is a member of the Committee on Publication Ethics (COPE).

Published by Libertas Academica. Learn more about this journal.

Introduction

Transient receptor potential (TRP) ion channels are cation channels with putative roles in many physiological signaling pathways. Mammalian TRPs are comprised of six main groups: TRPA (ankyrin), TRPC (canonical), TRPM (melastatin), TRPML (mucolipin), TRPP (polycystin), and TRPV (vanilloid).^{1,2} Generally, the TRPC channels are nonselective cation channels; only two are highly permeable Ca²⁺ channels and two are impermeable for Ca²⁺. Importantly, several TRPs are permeable for Mg²⁺ and Zn²⁺.³ TRPs are extensively expressed on almost all cells and therefore are likely to have significant effects on physiological functions.³ Dysregulation in TRPs has been associated with pathological conditions and diseases including chronic pain, overactive bladder, diabetes, chronic obstructive pulmonary disease, cardiac hypertrophy, familial Alzheimer's disease, skin diseases, skeletal dysplasias, motor neuropathies, neurosensory neuropathies

including Charcot-Marie-Tooth disease (type 2C), and cancer.^{4–8} TRP ion channels are activated following fluctuations or deviations in the cellular environment. Factors that may influence these changes are stressors including pathogens, temperature, pressure, chemicals, oxidation/reduction, toxins, osmolarity, and pH.^{9,10}

Chronic fatigue syndrome (CFS) is an unexplained disorder with multiple physiological impairments. Research to date suggests significant immune impairment; however, the mechanism of this disorder remains to be determined. CFS patients may have reactions to a number of environmental and biological factors.^{11–13} Moreover, there is evidence to suggest that CFS may have an allergic component.^{14–16} Atypical TRP expression has been reported in CFS, particularly upregulation in the expression of *TRPV1*.¹⁷ As TRPs regulate a plethora of physiological signaling pathways, they may have a role in CFS. A number of channelopathies



have been associated with TRP genes and these have consequences for cellular function.^{4,18,19} Additionally, TRP channels may be targeted during inflammatory reactions, as they are easily activated in the presence of irritants, inflammatory products, and xenobiotic toxins. Incidentally, CFS patients report significant sensitivity to environmental toxins and irritants, but the causes of these sensitivities remain to be fully investigated. The purpose of this pilot study was to determine whether polymorphisms in SNPs associated with TRP ion channel genes are a contributory factor in the pathogenesis of CFS.

Methodology

Participants. One-hundred and fifteen CFS patients and 90 nonfatigued controls were recruited for this study. Of the 115 CFS patients (age = 48.68 ± 1.06 years), 84 (73.04%) were women and 31 (26.96%) were men. The 90 nonfatigued controls (age = 46.48 ± 1.22 years) comprised 59 (65.56%) women and 31 (34.44%) men. CFS patients were defined in accordance with the 1994 Center for Disease Prevention and Control (CDC) criteria for CFS.²⁰ All participants in the patient and nonfatigued control groups were of European descent, and all were residents of Australia at the time of blood collection. Approval for the study was granted by the Institutional Ethics Review Board at Griffith University (Ref No: MSC/18/13/HREC) and the research complied with the principles of the Declaration of Helsinki.

Ten milliliters of whole blood samples were collected from all participants in to ethylenediamine tetraacetic acid tubes. Written consent was obtained from all participants prior to sample collection.

DNA extraction. Genomic DNA was extracted from all whole blood samples using the Qiagen DNA blood mini-kit as per manufacturer's instructions (Qiagen). The Nanodrop (Nanodrop) was used to assess the quality and quantity of the DNA extracted. Approximately 2 μ g of genomic DNA was used in the SNP assay.

SNP genotyping studies. SNP analysis was performed by Geneworks using the MassARRAY iPLEX Gold Assay (Sequenom Inc.) as previously defined. Customized assays were developed for 240 SNPs across the 21 TRP genes (*TRPA1*, *TRPC1*, *TRPC2*, *TRPC3*, *TRPC4*, *TRPC6*, *TRPC7*, *TRPM1*, *TRPM2*, *TRPM3*, *TRPM4*, *TRPM5*, *TRPM6*, *TRPM7*, *TRPM8*, *TRPV1*, *TRPV2*, *TRPV3*, *TRPV4*, *TRPV5*, and *TRPV6*). Primers and extension primers were created for each of the SNPs using the Assay Designer (Sequenom Inc.) according to the manufacturer's instructions. Briefly, DNA was amplified via polymerase chain reaction (PCR) under the following conditions: 94°C for 2 minutes, 94°C for 30 seconds, 56°C for 30 seconds, and 72°C for 1 minute. Amplification products were then treated with shrimp alkaline phosphatase at 37°C for 40 minutes, 85°C for 5 minutes reaction, and a final incubation at 4°C. Extension primers were optimized to control the signal-to-noise ratio where unextended primers

(UEPs) were examined on the spectroCHIP and evaluated in Typer 4.0 to enable the division into low-mass UEP, medium-mass UEP, and high-mass UEP. To perform the iPLEX extension reaction, a mixture containing iPLEX Gold reaction was prepared using iPLEX Gold Buffer Plus, iPLEX termination mix, iPLEX enzyme, and primer mix. The iPLEX reaction was cycled at an initial denaturation of 94°C for 30 seconds, annealing at 52°C for 5 minutes, extension at 80°C for 5 minutes (five cycles of annealing and extension were performed, but the whole reaction was performed in 40 cycles) and extension again at 72°C for 3 minutes. Resin beads were used to rinse all iPLEX Gold reaction products. Following the iPLEX Gold reaction, MassARRAY was performed using the MassARRAY mass spectrometer, and the data generated were analyzed using the TyperAnalyzer software.

Statistical analysis. The PLINK v1.07²⁰ whole genome analysis tool set was used to determine associations between the CFS patients and the nonfatigued control group. A two-column χ^2 test was used to determine significance, where a *P*-value of <0.05 was determined to be significant. Data analysis was performed by the Australian Genome Research Facility.

Results

SNP association studies. Of the 240 SNPs that were examined in the present study, 233 were successfully identified in both participants groups. Thirteen were observed to be significantly associated with CFS (Table 1). Nine of these SNPs were associated with *TRPM3* (rs12682832; *P* ≤ 0.003, rs11142508; *P* < 0.004, rs1160742; *P* < 0.08, rs4454352; *P* ≤ 0.013, rs1328153; *P* ≤ 0.013, rs3763619; *P* ≤ 0.014, rs7865858; *P* ≤ 0.021, rs1504401; *P* ≤ 0.041, rs10115622; *P* ≤ 0.050), while the remaining SNPs were associated with *TRPA1* (rs2383844; *P* ≤ 0.040, rs4738202; *P* ≤ 0.018), and *TRPC4* (rs6650469; *P* ≤ 0.016, rs655207; *P* ≤ 0.018). A summary of the 233 SNPs that were reported in both participant groups can be found in Supplementary Table 1.

Discussion

The purpose of this pilot study was to determine the presence of possible SNP variations in CFS patients with a specific focus on SNPs within the coding sequences of 21 TRP ion channel genes. Out of the 240 SNPs examined, 13 alleles were found to be significantly associated with CFS patients compared with the nonfatigued controls. As CFS is a heterogeneous condition, it is likely that further stratification of patients may elucidate further allelic associations. These alleles were located in the gene sequence of one of the canonical TRPs ion channels (*TRPC4*), one ankyrin (*TRPA1*), and one melastatin TRP ion channel (*TRPM3*).

SNPs located within a coding sequence may or may not necessarily change the amino acid sequence of the protein that is produced. As such, an SNP that does not alter the polypeptide sequence is termed synonymous (sometimes called a silent

**Table 1.** Analysis of the frequency distribution and significance of TRP single nucleotide polymorphisms (SNPs) in CFS patients and nonfatigued controls in rank order of significance.

GENE	CHROMOSOME	RefSNPID	A1	A2	FREQUENCY_A	FREQUENCY_U	χ^2	P-VALUE
TRPM3	9	rs12682832	A	G	0.444	0.293	8.808	0.003*
TRPM3	9	rs11142508	C	T	0.445	0.298	8.438	0.004*
TRPM3	9	rs1160742	A	G	0.470	0.333	7.063	0.008*
TRPM3	9	rs4454352	C	T	0.240	0.137	6.232	0.013*
TRPM3	9	rs1328153	C	T	0.240	0.137	6.232	0.013*
TRPM3	9	rs3763619	A	C	0.440	0.316	5.990	0.014*
TRPC4	13	rs6650469	T	C	0.505	0.380	5.775	0.016*
TRPC4	13	rs655207	G	T	0.505	0.381	5.639	0.018*
TRPA1	8	rs4738202	A	G	0.369	0.253	5.591	0.018*
TRPM3	9	rs7865858	A	G	0.450	0.331	5.340	0.021*
TRPA1	8	rs2383844	G	A	0.505	0.398	4.218	0.040*
TRPM3	9	rs1504401	T	C	0.100	0.173	4.172	0.041*
TRPM3	9	rs10115622	A	C	0.335	0.435	3.837	0.050*
TRPM4	19	rs10403114	G	A	0.293	0.390	3.802	0.051
TRPV3	17	rs9909424	G	A	0.115	0.060	3.442	0.064
TRPC4	13	rs612308	A	G	0.439	0.537	3.393	0.065
TRPM3	9	rs7860377	A	C	0.350	0.262	3.314	0.069
TRPC7	5	rs2673930	C	A	0.200	0.280	3.218	0.073
TRPC4	13	rs603955	C	T	0.445	0.536	3.008	0.083
TRPM3	9	rs11142798	C	G	0.135	0.202	2.998	0.083
TRPM3	9	rs4744611	G	A	0.360	0.446	2.843	0.092
TRPM2	21	rs1785452	T	C	0.215	0.289	2.67	0.102
TRPM3	9	rs1566838	G	T	0.460	0.375	2.669	0.102
TRPA1	8	rs1384002	T	C	0.495	0.410	2.664	0.103
TRPM6	9	rs2274924	G	A	0.115	0.175	2.652	0.103
TRPM3	9	rs1394309	G	A	0.030	0.065	2.608	0.106
TRPC4	13	rs2985167	G	A	0.340	0.422	2.577	0.108
TRPM5	11	rs2301698	G	T	0.530	0.446	2.551	0.110
TRPM6	9	rs944857	C	T	0.185	0.125	2.476	0.116
TRPM2	21	rs762426	G	A	0.160	0.223	2.325	0.127

Notes: SNP with 115 CFS/ME patients and 84 controls. Data presented are included for $P < 0.130$. Data presented for gene (*TRPM2*, 3, 4, 5, 6, *TRPA1*, *TRPC4*, and *TRPV3*), chromosome location (CHR), reference SNP identification (RefSNPID), base pair (BP) location of SNP, alleles (A1 and A2), allelic frequency A (Frequency_A) of this allele in CFS cases, frequency U (Frequency_U) of this allele in controls, chi-square (χ^2) for basic allelic test (1 df), and (*) P-value for this test set at a significance of <0.05 .

variant), while an SNP that results in a different polypeptide sequences is referred to as nonsynonymous, potentially resulting in altered gene transcripts and disease phenotypes. In this investigation, we report TRP SNP anomalies, suggesting they may mediate the potential onset or clinical presentation of CFS; however, this needs to be confirmed with larger cohorts. Nonetheless, this current study suggests the potential role for aberrant TRPs in CFS. Further rationale is provided below, whereby TRPs are demonstrated to play significant physiological roles. In particular, CFS patients report symptoms that may be associated with aberrant TRP function.

TRPC4 is activated via receptor-dependent activation of the G_{q11} /PLC (phospholipase C)/ γ pathway but also via $G_{\alpha i}$

proteins, $PI(4,5)P_2$ proteins, and also intracellular Ca^{2+} .²¹ It is mainly involved in vasomotor function, aggregation of platelets, and smooth muscle function. Incidentally, Ca^{2+} is known to be required for the regulation of immune cells, as Ca^{2+} acts as a second messenger for most cells, particularly T cells and B cells. Intracellular Ca^{2+} increases when lymphocyte receptors are exposed to antigens.²² In CFS patients, there are numerous reports on compromises to immune function, although there is limited information on the role of Ca^{2+} in these patients. However, dysregulation in TRPCs may affect intracellular calcium concentration and incidentally lymphocyte function. Lymphocytes such as natural killer (NK) cells and T cells have been shown to be compromised in CFS. In NK cells, Ca^{2+} enhances



cytotoxic activity, and its depletion or excessive influx may have severe consequences on NK cells function. In CFS, reduced cytotoxic activity has been consistently reported,^{23–28} and this may be related to the dysregulation in Ca^{2+} .

Dysregulation of TRPCs may affect neuronal responses, in particular those associated with the stimulation of muscarinic receptors. Following activation of TRPCs by PLCs, an influx of Ca^{2+} occurs, causing an induction in muscarinic receptors, and maintains incessant neuronal firing.^{29,30} Hence, secretion of Ca^{2+} and the availability of TRPCs in the neuronal environment are paramount to optimal muscarinic receptor function and overall function of the brain. Importantly, this process is essential for memory, attention, sensory acuity, emotion, pain, and motor control,^{31,32} and occurs in the amygdala, entorhinal cortex, hippocampus, and prefrontal cortex.³³ Neuronal deficits involving memory and attention have been identified in CFS.^{34–36} Deletion or compromises to *TRPC4* may also affect intestinal function. *TRPC4* and *TRPC6* pair with muscarinic receptors in the intestine, activating smooth muscle depolarization, inflow of Ca^{2+} , and smooth muscle contraction.³⁷ Intestinal dysfunction is a component of CFS;³⁸ however, the extent of damage to the intestinal wall or the exact role of ion channels in the intestine remains to be determined. *TRPC4* may be simultaneously regulated by the G protein coupled receptors (GPCRs) $G\alpha i$ and $G\alpha q$.³⁹

TRPA1 is a multiple chemical receptor that has been identified on nociceptive sensory neurons (C fibers) and has a role in the regulation of the release of neuropeptides, pain sensation, and inflammation.⁴⁰ It may be activated by both exogenous and endogenous inflammatory agents, resulting in inflammation and pain.⁴¹ GPCRs also activate *TRPA1* via PLC signaling, sensitizing the ion channel to various stimuli.⁴² *TRPA1* may be activated and subsequently inactivated in the presence of intracellular and extracellular calcium concentrations.^{43,44} *TRPA1* gene has been proposed to affect sensitivity to nociceptive stimuli;⁴⁵ hence CFS patients expressing SNPs in the *TRPA1* gene may increase their sensitivity to nociceptive stimuli. In the CNS, astrocytes express TRPA1 channels, and these channels are necessary for calcium uptake and neuronal regulation in the astrocytes. Changes in the level of calcium may therefore affect the function of astrocytes and interneuron communication.^{44,46} Activation of TRPA1 has been shown to induce acute headache, and this may occur through the calcitonin gene related peptide (CGRP), causing vasodilation in the meningeal artery.^{44,47} Importantly, headache is a prominent symptom of CFS. TRPA1 is also a key player in migraine and neuropathic joint and muscle pain, which is most often experienced by patients with fibromyalgia.^{48,49} TRPA1 forms functional heterotetramers with TRPV1; hence variations in the *TRPA1* gene may suggest functional deficits to *TRPV1* that may not be related to polymorphism in nucleotides.⁵⁰ Importantly, Carreno et al⁵¹ reported that TRPV1 contributes to the genetic susceptibility to migraine in a large study population of 500 Spanish

participants, suggesting TRPA1 and TRPV1 may play a role in symptom presentation of CFS patients. Interestingly, analgesics and antinociceptive drugs target TRPV1 and TRPA1, respectively, to alleviate pain sensation^{52–54} and these drugs are routinely prescribed to CFS patients. Perhaps in CFS, these drugs may not be effective due to impairments or variations in these ion channels.

TRPM channels are mostly permeable to magnesium and calcium. Only TRPM4 and TRPM5 are impermeable for divalent cations. TRPM3 is permeable for cations including Ca^{2+} and Zn^{2+} . However, the permeation profile highly depends on the expressed spliced variant.⁵⁵ No hereditary TRPM3 channelopathy has been described to date. TRPM3 has been implicated in inflammatory pain syndromes, rheumatoid arthritis, and secretion of proinflammatory cytokines. As pancreatic β cells also have a high proportion of TRPM3 channels,^{44,56–58} there is the likelihood of perturbations in insulin/glucose regulation in CFS patients. Metabolic disturbance has also long been identified as a cardinal feature of CFS. The most characterized TRPM3 in humans is in the central nervous system (CNS) and eye⁵⁵ where missense mutation of the TRPM3 gene has also been found to underlie the development of cataract and glaucoma.⁵⁹ TRPM3 is involved in the detection of heat and in pain transmission. TRPM3-deficient mice exhibit clear deficits in their avoidance responses to noxious heat and in the development of inflammatory heat hyperalgesia.⁵⁵ Dysregulation in thermoregulatory responses has been reported in CFS patients.⁶⁰ Generalized pain is a characteristic of CFS and occurs in the absence of tissue damage, and this is suggestive of potential CNS impairments.⁶¹ As TRPM3 has a role in nociception and thermoregulation, it may have a role in the pathomechanism of CFS. Additionally, TRPM3 is activated by pregnenolone sulfate, suggesting that it has neuroendocrine effects^{62,63} and might also be involved in the regulation of glutamatergic signaling in the brain.⁶⁴

These preliminary findings implicate TRP ion channels in the etiology and pathomechanism of CFS. Dysregulation of TRPs, including the TRPM3 family, is likely pertinent in predisposing CFS patients to calcium metabolism perturbations and aligns with symptom presentation. Potentially, dysregulated influx of calcium ions into cells will impact a number of vital components of cell regulatory machinery. These components include calcium-sensitive adenylate cyclases (ACs) and hence cAMP expression and function. For example, isolated cell types that have been shown previously to have calcium-sensitive cell regulatory mechanisms in CFS patients may enable further elucidation of TRP ion channels and the likely consequences in CFS. Furthermore, population analysis of TRP SNPs for CFS susceptibility, as well as the proposed various subtypes, needs to be considered. This undertaking will likely be of considerable importance to public health and public health practitioners, as well as to researchers to assess the role of TRP ion channels in CFS symptomatology, severity, and predisposition.



Acknowledgments

The authors would like to thank Dr Lavinia Gordon, Australian Genome Research Facility, Melbourne, Australia, for completing the bioinformatics SNP analysis.

Author Contributions

Designed and developed all experiments as well as analysis, revisions and final preparation of this article: SMG, DRS, PS, BN. Involved in the sample preparation and drafting of the manuscript: SBR, EWB. All authors reviewed and approved of the final manuscript.

Supplementary Data

Supplementary table 1. Analysis of the frequency distribution and significance of TRP single nucleotide polymorphisms (SNPs) in CFS patients and nonfatigued controls in rank order of significance.

REFERENCES

- Clapham DE. TRP channels as cellular sensors. *Nature*. 2003;426(6966):517–524.
- Nilius B, Owsianik G, Voets T, Peters JA. Transient receptor potential cation channels in disease. *Physiol Rev*. 2007;87(1):165–217.
- Nilius B, Owsianik G. The transient receptor potential family of ion channels. *Genome Biol*. 2011;12(3):218.
- Nilius B, Szallasi A. Transient receptor potential channels as drug targets: from the science of basic research to the art of medicine. *Pharmacol Rev*. 2014;66(3):676–814.
- Nilius B, Biro T, Owsianik G. TRPV3: time to decipher a poorly understood family member! *J Physiol*. 2014;592(pt 2):295–304.
- Nilius B, Biro T. TRPV3: a 'more than skinny' channel. *Exp Dermatol*. 2013;22(7):447–452.
- Nilius B, Voets T. The puzzle of TRPV4 channelopathies. *EMBO Rep*. 2013;14(2):152–163.
- Vennekens R, Menigoz A, Nilius B. TRPs in the brain. *Rev Physiol Biochem Pharmacol*. 2012;163:27–64.
- Moran MM, McAlexander MA, Biro T, Szallasi A. Transient receptor potential channels as therapeutic targets. *Nat Rev Drug Discov*. 2011;10(8):601–620.
- Nieto-Posadas A, Jara-Oseguera A, Rosenbaum T. TRP channel gating physiology. *Curr Top Med Chem*. 2011;11(17):2131–2150.
- Fernandez-Sola J, Lluís Padierna M, Nogue Xarau S, Munne Mas P. [Chronic fatigue syndrome and multiple chemical hypersensitivity after insecticide exposure]. *Med Clin*. 2005;124(12):451–453.
- Lavergne MR, Cole DC, Kerr K, Marshall LM. Functional impairment in chronic fatigue syndrome, fibromyalgia, and multiple chemical sensitivity. *Can Fam Physician*. 2010;56(2):e57–e65.
- Brown MM, Jason LA. Functioning in individuals with chronic fatigue syndrome: increased impairment with co-occurring multiple chemical sensitivity and fibromyalgia. *Dyn Med*. 2007;6:6.
- Lind R, Berstad A, Hatlebakk J, Valeur J. Chronic fatigue in patients with unexplained self-reported food hypersensitivity and irritable bowel syndrome: validation of a Norwegian translation of the Fatigue Impact Scale. *Clin Exp Gastroenterol*. 2013;6:101–107.
- Aboudiab T, Leke L, Skonieczny M, Chouraki JP. [Are IgE-independent food hypersensitivity and chronic fatigue syndrome related?]. *Arch Pediatr*. 2004;11(8):975–977.
- Brunet JL, Fatoohi F, Liaudet AP, Cozon GJ. [Role of pathological delayed-type hypersensitivity in chronic fatigue syndrome: importance of the evaluation of lymphocyte activation by flow cytometry and the measurement of urinary neopterin]. *Allerg Immunol*. 2002;34(2):38–44.
- Light AR, Bateman L, Jo D, et al. Gene expression alterations at baseline and following moderate exercise in patients with chronic fatigue syndrome and fibromyalgia syndrome. *J Intern Med*. 2012;271(1):64–81.
- Gees M, Owsianik G, Nilius B, Voets T. TRP channels. *Compr Physiol*. 2012;2(1):563–608.
- Nilius B, Owsianik G. Transient receptor potential channelopathies. *Pflugers Arch*. 2010;460(2):437–450.
- Harvard. *PLINK Whole Genome Association Analysis Toolset*; 2014. Available at: <http://pngu.mgh.harvard.edu/purcell/plink/>
- Freichel M, Tsvilovskyy V, Camacho-Londono JE. TRPC4- and TRPC4-containing channels. *Handb Exp Pharmacol*. 2014;222:85–128.
- Lewis RS. Calcium signaling mechanisms in T lymphocytes. *Annu Rev Immunol*. 2001;19:497–521.
- Brenu EW, Huth TK, Hardcastle SL, et al. Role of adaptive and innate immune cells in chronic fatigue syndrome/myalgic encephalomyelitis. *Int Immunol*. 2014;26(4):233–242.
- Brenu EW, Ashton KJ, van Driel M, et al. Cytotoxic lymphocyte microRNAs as prospective biomarkers for chronic fatigue syndrome/myalgic encephalomyelitis. *J Affect Disord*. 2012;141(2–3):261–269.
- Brenu EW, van Driel ML, Staines DR, et al. Longitudinal investigation of natural killer cells and cytokines in chronic fatigue syndrome/myalgic encephalomyelitis. *J Transl Med*. 2012;10:88.
- Brenu EW, van Driel ML, Staines DR, et al. Immunological abnormalities as potential biomarkers in chronic fatigue syndrome/myalgic encephalomyelitis. *J Transl Med*. 2011;9:81.
- Brenu EW, Staines DR, Baskurt OK, et al. Immune and hemorheological changes in chronic fatigue syndrome. *J Transl Med*. 2010;8:1.
- Barker E, Fujimura SF, Fadem MB, Landay AL, Levy JA. Immunologic abnormalities associated with chronic fatigue syndrome. *Clin Infect Dis*. 1994;18(suppl 1):S136–S141.
- Zhang Z, Seguela P. Metabotropic induction of persistent activity in layers II/III of anterior cingulate cortex. *Cereb Cortex*. 2010;20(12):2948–2957.
- Zhang Z, Reboreda A, Alonso A, Barker PA, Seguela P. TRPC channels underlie cholinergic plateau potentials and persistent activity in entorhinal cortex. *Hippocampus*. 2011;21(4):386–397.
- Rainville P. Brain mechanisms of pain affect and pain modulation. *Curr Opin Neurobiol*. 2002;12(2):195–204.
- Sewards TV, Sewards MA. The medial pain system: neural representations of the motivational aspect of pain. *Brain Res Bull*. 2002;59(3):163–180.
- Yan HD, Villalobos C, Andrade R. TRPC channels mediate a muscarinic receptor-induced after depolarization in cerebral cortex. *J Neurosci*. 2009;29(32):10038–10046.
- Caseras X, Mataix-Cols D, Giampietro V, et al. Probing the working memory system in chronic fatigue syndrome: a functional magnetic resonance imaging study using the n-back task. *Psychosom Med*. 2006;68(6):947–955.
- Caseras X, Mataix-Cols D, Rimes KA, et al. The neural correlates of fatigue: an exploratory imaginal fatigue provocation study in chronic fatigue syndrome. *Psychol Med*. 2008;38(7):941–951.
- Nakatomi Y, Mizuno K, Ishii A, et al. Neuroinflammation in patients with chronic fatigue syndrome/myalgic encephalomyelitis: an 11C-(R)-PK11195 PET Study. *J Nucl Med*. 2014;55(6):945–950.
- Tsvilovskyy VV, Zholos AV, Aberle T, et al. Deletion of TRPC4 and TRPC6 in mice impairs smooth muscle contraction and intestinal motility in vivo. *Gastroenterology*. 2009;137(4):1415–1424.
- Lakhan SE, Kirchgessner A. Gut inflammation in chronic fatigue syndrome. *Nutr Metab*. 2010;7:79.
- Kim H, Kim J, Jeon JP, et al. The roles of G proteins in the activation of TRPC4 and TRPC5 transient receptor potential channels. *Channels*. 2012;6(5):333–343.
- Bautista DM, Pellegrino M, Tsunozaki M. TRPA1: a gatekeeper for inflammation. *Annu Rev Physiol*. 2013;75:181–200.
- Wang S, Dai Y, Fukuoka T, et al. Phospholipase C and protein kinase A mediate bradykinin sensitization of TRPA1: a molecular mechanism of inflammatory pain. *Brain*. 2008;131(pt 5):1241–1251.
- Wilson SR, Gerhold KA, Bifolck-Fisher A, et al. TRPA1 is required for histamine-independent, Mas-related G protein-coupled receptor-mediated itch. *Nat Neurosci*. 2011;14(5):595–602.
- Nilius B, Prenen J, Owsianik G. Irritating channels: the case of TRPA1. *J Physiol*. 2011;589(pt 7):1543–1549.
- Wagner TF, Loch S, Lambert S, et al. Transient receptor potential M3 channels are ionotropic steroid receptors in pancreatic beta cells. *Nat Cell Biol*. 2008;10(12):1421–1430.
- Schütz M, Oertel BG, Heimann D, et al. Consequences of a human TRPA1 genetic variant on the perception of nociceptive and olfactory stimuli. *PLoS One*. 2014;9(4):e95592.
- Shigetomi E, Tong X, Kwan KY, Corey DP, Khakh BS. TRPA1 channels regulate astrocyte resting calcium and inhibitory synapse efficacy through GAT-3. *Nat Neurosci*. 2012;15(1):70–80.
- Nassini R, Materazzi S, Vriens J, et al. The 'headache tree' via umbrellulone and TRPA1 activates the trigeminovascular system. *Brain*. 2012;135(pt 2):376–390.
- Garrison SR, Stucky CL. The dynamic TRPA1 channel: a suitable pharmacological pain target? *Curr Pharm Biotechnol*. 2011;12(10):1689–1697.
- Garrison SR, Stucky CL. Contribution of transient receptor potential ankyrin 1 to chronic pain in aged mice with complete Freund's adjuvant-induced arthritis. *Arthritis Rheumatol*. 2014;66(9):2380–2390.



50. Fischer MJ, Balasuriya D, Jeggle P, et al. Direct evidence for functional TRPV1/TRPA1 heteromers. *Pflugers Arch*. 2014;466(12):2229–2241.
51. Carreño O, Corominas R, Fernández-Morales J, et al. SNP variants within the vanilloid TRPV1 and TRPV3 receptor genes are associated with migraine in the Spanish population. *Am J Med Genet B Neuropsychiatr Genet*. 2012;159B(1):94–103.
52. Marincsak R, Toth BI, Czifra G, Szabo T, Kovacs L, Biro T. The analgesic drug, tramadol, acts as an agonist of the transient receptor potential vanilloid-1. *Anesth Analg*. 2008;106(6):1890–1896.
53. Andersson DA, Gentry C, Alenmyr L, et al. TRPA1 mediates spinal antinociception induced by acetaminophen and the cannabinoid delta(9)-tetrahydrocannabinol. *Nat Commun*. 2011;2:551.
54. De Petrocellis L, Vellani V, Schiano-Moriello A, et al. Plant-derived cannabinoids modulate the activity of transient receptor potential channels of ankyrin type-1 and melastatin type-8. *J Pharmacol Exp Ther*. 2008;325(3):1007–1015.
55. Oberwinkler J, Philipp SE. Trpm3. *Handb Exp Pharmacol*. 2014;222:427–459.
56. Thiel G, Muller I, Rossler OG. Signal transduction via TRPM3 channels in pancreatic beta-cells. *J Mol Endocrinol*. 2013;50(3):R75–R83.
57. Colsoul B, Vennekens R, Nilius B. Transient receptor potential cation channels in pancreatic beta cells. *Rev Physiol Biochem Pharmacol*. 2011;161:87–110.
58. Wagner TF, Drews A, Loch S, et al. TRPM3 channels provide a regulated influx pathway for zinc in pancreatic beta cells. *Pflugers Arch*. 2010;460(4):755–765.
59. Bennett TM, Mackay DS, Siegfried CJ, Shiels A. Mutation of the melastatin-related cation channel, TRPM3, underlies inherited cataract and glaucoma. *PLoS One*. 2014;9(8):e104000.
60. Wyller VB, Godang K, Morkrid L, Saul JP, Thaulow E, Walloe L. Abnormal thermoregulatory responses in adolescents with chronic fatigue syndrome: relation to clinical symptoms. *Pediatrics*. 2007;120(1):e129–e137.
61. Mecus M, Nijs J. Central sensitization: a biopsychosocial explanation for chronic widespread pain in patients with fibromyalgia and chronic fatigue syndrome. *Clin Rheumatol*. 2007;26(4):465–473.
62. Nilius B, Voets T. A TRP channel-steroid marriage. *Nat Cell Biol*. 2008;10(12):1383–1384.
63. Drews A, Mohr F, Rizun O, et al. Structural requirements of steroidal agonists of transient receptor potential melastatin 3 (TRPM3) cation channels. *Br J Pharmacol*. 2014;171(4):1019–1032.
64. Zamudio-Bulcock PA, Everett J, Harteneck C, Valenzuela CF. Activation of steroid-sensitive TRPM3 channels potentiates glutamatergic transmission at cerebellar Purkinje neurons from developing rats. *J Neurochem*. 2011;119(3):474–485.