### CD151—A Striking Marker for Cancer Therapy



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**ABSTRACT:** Cluster of differentiation 151 (CD151) is a member of the mammalian tetraspanin family, which is involved in diverse functions such as maintaining normal cellular integrity, cell-to-cell communication, wound healing, platelet aggregation, trafficking, cell motility and angiogenesis. CD151 also supports de novo carcinogenesis in human skin squamous cell carcinoma (SCC) and tumor metastasis. CD151 interacts with  $\alpha$ 3 $\beta$ 1 and  $\alpha$ 6 $\beta$ 4 integrins through palmitoylation where cysteine plays an important role in the association of CD151 with integrins and non-integrin proteins. Invasion and metastasis of cancer cells were diminished by decreasing CD151 association with integrins. CD151 functions at various stages of cancer, including metastatic cascade and primary tumor growth, thus reinforcing the importance of CD151 as a target in oncology. The present review highlights the role of CD151 in tumor metastasis and its importance in cancer therapy.

KEYWORDS: CD151, integrins, metastasis, tetraspanin

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#### Insight on CD151 (Cluster of Differentiation 151)

Tetraspanins are four-transmembrane-spanning proteins with short cytoplasmic N- and C-termini and one small and one large extracellular domain, namely (EC1 and EC2), with a unique cysteine motif in EC2 domain.<sup>1</sup> They are highly expressed on cell surface and/or intracellular vesicle. Palmitoylation of intracellular and juxtamembrane cysteine of tetraspanins along with specific integrins contributes to tetraspanin complex formation. This complex formation protects tetraspanins from lysosomal degradation and promotes increased cell–cell interaction.<sup>2</sup>

CD151 (glycoprotein-27 (GP-27)/Red blood cell antigen MER 2 (MER 2)/platelet-endothelial tetraspan antigen-3 (PETA-3)/Raph blood group antigen (RAPH)/Membrane Glycoprotein SFA-1 (SFA-1)./tetraspanin-24 (TSPAN-24)) is a plasma membrane protein that belongs to tetraspanin superfamily. CD151 forms tetraspanin web with integrins such as  $\alpha$ 3 $\beta$ 1,  $\alpha$ 6 $\beta$ 1,  $\alpha$ 7 $\beta$ 1, and  $\alpha$ 6 $\beta$ 4;<sup>2</sup> membrane receptors; intracellular signaling molecules such as phosphoinositide 4-kinase (PI4K) and protein kinase C (PKC);<sup>3,4</sup> immunoglobulin super family proteins and other tetraspanins such as CD9, CD81 and CD63. These tetraspanin-enriched microdomains (TEMs) serve as molecular facilitator.<sup>5</sup> CD151 contains a potential tyrosine-based sorting motif in the C-terminal domain. The YXX¢ (tyrosine linked with hydrophobic amino acid) motif (where <sup>\$\phi\$</sup> is a hydrophobic residue) is required for endocytosis and sorting of proteins from the trans-Golgi network (TGN)

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to lysosomes for degradation. A mutation in the CD151 YXX¢ motif diminishes CD151 internalization and affects integrindependent cell migration.<sup>6,7</sup> The major functions of CD151 are maintenance of epithelial cell integrity, wound healing, platelet aggregation, regulation of membrane fusion, trafficking, cell motility, angiogenesis and tumor metastasis.<sup>8</sup> It is normally expressed in endothelial cells and platelets and frequently overexpressed in cancer cells where it is functionally associated with cancer progression and metastasis.<sup>9</sup>

#### **Genetics of CD151**

CD151 (Raph blood group) gene is represented as *CD151*, which is located on the short (p) arm of chromosome 11 at position 15.5. More precisely, CD151 gene is located from base pair 832, 951 to base pair 838, 834 on chromosome 11.<sup>10</sup> It is an autosomal gene with sexually dimorphic expression, involved in signal transduction, cell proliferation and death. It is also involved in differential neural development, cognitive function and neurological diseases.<sup>11</sup>

## Association of CD151 with Integrins and Other Proteins

Metastasis is facilitated by cell–cell interactions between tumor cells and endothelium in which cell-adhesion molecules, such as integrins and selectins, play an important role. Hemler<sup>12</sup> has demonstrated that CD151 interacts directly with  $\alpha 3\beta 1$  and  $\alpha 6\beta 4$  integrins through their palmitoylated

cysteine residues. CD151 acts as master regulators of  $\alpha 6\beta 4$ ,  $\alpha 6\beta 1$  and  $\alpha 3\beta 1$  integrin-assembly into TEMs.<sup>13</sup> The major laminin-332 cellular receptors with  $\alpha 3\beta 1$  and  $\alpha 6\beta 4$  integrinsdependent adhesion, migration and signaling are impaired or altered by CD151 excision, indicating the vital role of CD151 in association with integrins.<sup>14,15</sup> Mapping of the integrin-CD151 association was performed using chimeric  $\alpha 6/\alpha 3$  integrins and CD151/NAG2 TM4SF proteins, which showed an interaction of amino acids from 570 to 705 at the extracellular sites of  $\alpha_2$  integrin and amino acids from 186 to 217 on a large extracellular loop of CD151.16 Devbhandari et al17 reviewed the role of CD151/integrin  $\beta$ 1 complex in cancer metastasis. Yauch et al<sup>3</sup> reported that specific site on the large extracellular loop of CD151 associates with  $\alpha$ 3 $\beta$ 1 integrin with unusually high stoichiometry, proximity and stability. In the same year, Sterk et al<sup>18</sup> reported that CD151 uses the same site for interaction with  $\alpha 6\beta 1$ ,  $\alpha 6\beta 4$  and other integrins.

#### Role of CD151 in De Novo Carcinogenesis

CD151, in association with laminin and laminin-binding integrins, involves in the regulation of carcinogenesis. Li et al<sup>19</sup> evaluated the role of CD151 in de novo carcinogenesis, multiplicity and progression of skin squamous cell carcinoma (SCC) in comparison with normal cells. CD151 supports survival and proliferation of keratinocytes by activation of transcription factor STAT3, a regulator of cell proliferation and apoptosis. CD151 regulates  $\alpha 6\beta 4$  distribution by enhancing protein kinase C $\alpha$ (PKC $\alpha$ )— $\alpha 6\beta 4$  integrin association and PKC-dependent  $\beta 4$  S1424 phosphorylation. CD151 in complex with PKC $\alpha$ enhances invasive behavior by phoshorylating  $\beta 4$  integrin, thus affects subcellular localization and epithelial disruption. In addition, CD151 knockout mice showed a decrease in tumor latency, tumor incidence, multiplicity and size. Hence, CD151 targeting may be therapeutically beneficial in cancer therapy.



#### CD151 and Cancer Metastasis

Deregulation of various tetraspanins is reported in human malignancy.<sup>20</sup> Overexpression of CD151 was also reported to associate with poor prognosis of lung,<sup>21</sup> colon,<sup>22</sup> esophageal,<sup>23</sup> pancreatic<sup>24</sup> and endometrial cancers.<sup>25</sup> In addition, several evidences have supported the contribution of CD151 in cancer metastasis.<sup>26</sup> Loss of CD151 decreased the integrin-mediated cell migration, spreading, and invasion through FAK and Rac1mediated signaling.<sup>13</sup> Earlier reports have shown that downregulation of CD151 by short-hairpin RNA decreased the tumorigenicity and communication between tumor and endothelial cells. This emphasizes CD151 as a potential prognostic marker.<sup>27</sup> CD151 also modulates the activity of cytokine-like transforming growth factor- $\beta$  (TGF- $\beta$ ). CD151 in association with dimerized TGF-B receptor promotes invasion and metastasis through the activation of Smad2/3, c-Akt, Erk1/2, JNK, JUN and matrix metalloproteinase-9 (MMP-9) signaling pathways (Fig. 1). Sadej et al reported that CD151 is a positive regulator of TGF-β-induced signaling in cancer metastasis.<sup>28</sup>

Studies on CD151-null mice showed impaired pathological angiogenesis without vascular defect during normal development.<sup>29</sup> Adhesion-dependent activation of endothelial cells caused diminished expressions of PKC/c-Akt, e-NO, Rac and Cdc42, which are important signaling pathways of angiogenesis and cytoskeleton reorganization without altering the expression of Raf, ERK, p38, MAP kinase, FAK and Src. CD151 regulates cytoskeletal reorganization, invasion and cell-adhesion functions of endothelial cells during pathological angiogenesis by modulating laminin-binding integrinsmediated activation of PI3K/Akt, JNK and PKC pathways<sup>29</sup> (Fig. 2). Evaluation of prognostic significance of CD151 expression with estrogen receptor (ER), progesterone receptor (PR), human epidermal growth factor receptor-2 "(HER-2) and E-cadherin indicated a strong concordance of CD151



**Figure 1.** Tumor growth factor beta 1 (TGF- $\beta$ 1) and CD151-mediated signaling. TGF- $\beta$ 1 binds to TGF- $\beta$ 1 receptors and leads to dimerization of receptors. These receptors associate with CD151 and activate JNK, JUN, MMP-9, Smad 2/3, c-Akt, and Erk1/2 signaling pathways involved in invasion and metastasis. Yellow circle—TGF- $\beta$ ; double-headed arrow—TGF- $\beta$  receptor.





**Figure 2.** Role of CD151 in angiogenesis, invasion, and cytoskeleton reorganization. An association of integrin  $\alpha$ 6 $\beta$ 1 (orange) with CD151 activates PI3K, Akt, NOS, JNK, JUN, and MMP-9 signaling, leading to angiogenesis and invasion. CD151 interacts with  $\alpha$ 3 $\beta$ 1 (yellow) and causes activation of invasive signaling, PKC, and cdc42, leading to cytoskeleton reorganization.

with E-cadherin in type 2 endometrial cancer. Kohno et al demonstrated that CD151 also enhanced cell motility, invasion and metastasis through focal adhesion kinase (FAK).<sup>30</sup>

Haeuw et al<sup>31</sup> reported that monoclonal antibodies (mAbs) targeted to CD151 inhibited cell motility. Similarly, Fei et al<sup>32</sup> reported that CD151-AAA mutant inhibited cell-proliferation, migration and chemotaxis in HepG2 cells. Genetic ablation of CD151 inhibited metastasis in a transgenic mouse model without showing any noticeable effect on expression of markers associated with proliferation, apoptosis or angiogenesis in primary tumors.<sup>33</sup>

Shanmukhappa et al have demonstrated for the first time that CD151siRNA and anti-CD151 antibodies significantly reduced the porcine reproductive and respiratory syndrome virus (PRRSV) infection.<sup>34</sup> One recent study showed that simultaneous inhibition of CD9/CD81 and CD151 has a profound inhibitory effect on cancer metastasis through  $\alpha$ 3 $\beta$ 1-PKC $\alpha$ mediated signaling in MDA-MB 231 breast cancer cells.<sup>35</sup>

Several studies have demonstrated that CD151 is involved in cell-adhesion and formation of hemidesmosomes; thus, inhibition of CD151 expression by hypoxia decreased the cell–cell interaction and cell–matrix adhesion.<sup>36</sup> A study by RR Malla et al have demonstrated that silencing of cathepsin B and uPAR using siRNA decreased the interactions of CD151 with laminin-binding integrin  $\alpha$ 3 $\beta$ 1 and inhibits cell-adhesion and invasion in glioma.<sup>37</sup>

#### **CD151** Pathogenesis and Clinical Studies

CD151 is implicated in pathological processes associated with cancer progression, neoangiogenesis and epithelialmesenchymal transition (EMT).<sup>38–41</sup> CD151 overexpressing cancer cells acquired high migratory abilities, which are crucial for tumor cell invasion.<sup>42</sup> Overexpression of CD151 was also reported to associate with intercellular adhesiveness and wound healing.<sup>43</sup> CD151 plays a crucial role in the interaction of podocyte-GBM binding adhesion receptor and integrin  $\alpha 3\beta 1$ . Deletion of CD151 leads to reduced adhesiveness at cell–matrix interface, leading to glomerular nephropathy.<sup>44</sup>

CD151-null mice showed diminished tumor cell residence in the lungs after injection with Lewis lung carcinoma, which may be because of inhibition of angiogenesis.<sup>45</sup> In vitro studies showed that tumor-endothelial adhesion, tumortransendothelial migration and tumor –induced permeability were defective in CD151-null endothelial cells.<sup>46</sup> Blocking of CD151 using specific mAbs inhibited invasion without affecting primary tumor growth and tumor cell arrest or growth at the secondary site.<sup>26</sup> Knockdown of CD151 inhibited downstream signaling through Akt, ErK1/2 and FAK and markedly sensitizes ErbB2+ cancers.<sup>47</sup>

Kwon et al reported that overexpression of CD151 may be a potential molecular therapeutic target in advanced stages of breast cancer.<sup>48</sup> Recent clinical studies have demonstrated that a positive correlation exists between CD151 expression and progression of cancer cells.<sup>33</sup> An interesting fact has been reported recently that integrin-free CD151 can promote tumor cell migration without binding to integrin. This study suggests that CD151 can control migration, independent of integrin association.<sup>49</sup>

#### **Expression Levels of CD151**

Comparative analysis of tetraspanin expression in various types of cancers revealed that CD151 is highly expressed in solid tumor compared with non-solid tumor (Table 1). CD151 is usually localized to the basal and lateral junctions of tumor



 Table 1. Expression of tetraspanins in different cancers.

| TYPE OF CANCER   | CD9 | CD63 | CD81 | CD82 | CD151 | REFERENCE                        |
|------------------|-----|------|------|------|-------|----------------------------------|
| Breast cancer    | ++  | +    | +++  | +    | +++   | Penas et al, 200054              |
| Lung cancer      | +++ | ++   | ++   | +++  | +++   | Funakoshi et al, 200355          |
| Colon cancer     | +++ | +    | +++  | ++   | +++   | Le Naour et al, 200656           |
| Burkitt lymphoma | +   | +++  | +    | +    | +     | Ferrer et al, 1998 <sup>57</sup> |

Notes: Expression levels of tetraspanin proteins (CD9, CD63, CD81, CD82, and CD151) involved in cancer progression in different cancer cells. +, low; ++, medium; +++, high.

cells.<sup>50</sup> Solid tumors exhibit heterogeneity of neoplastic and normal cells at histological, genetic and gene expression levels. Considerable heterogeneity of CD151 expression was reported in various tumor tissues. The intensity of staining was noticeably weaker in well-differentiated cells of oral SCC. The gradient of CD151 expression was particularly prominent in the invasive front of tumors.<sup>51</sup> Overexpression of CD151 can be correlated with large tumor size, depth of invasion and advanced stage of tumor.<sup>52</sup> A number of investigations have suggested that high CD151 gene expression in cancer is associated with poor prognosis.<sup>53</sup> Tokuhara et al have reported that survival rate of patients with CD151-positive tumors was much lower than that of CD151-negative patients.<sup>21</sup> For optimal separation between low and high risk for overall survival, prognostic factors are usually considered as an optimized cutoff point.<sup>53</sup> The cutoff point of CD151 in ER negative breast cancer patients was reported as 14%<sup>48</sup> and 31%,<sup>13</sup> in lung cancer patients as 50%,<sup>21</sup> and in gastric cancer patients as >50%,<sup>52</sup> using immunohistochemical and RT-PCR analysis. These studies show the cutoff values to identify CD151 positivity, which may vary based on cancer type. However, large-scale prospective and retrospective studies on various cancers established CD151 as a prognostic marker for cancer therapy with minimal side effects on normal cells.<sup>13</sup>

#### **Future Prospects**

This review explores the importance of CD151 in the maintenance of cellular integrity and cell communication. It also imparts the role of CD151 in cancer cell migration, invasion, and metastasis. Thus, CD151 is a crucial target for cancer therapy. Disruption of tetraspanin web by targeting CD151 may affect the signaling pathways involved in cell survival and cancer progression. CD151 can be downregulated using mAbs, shRNA or gene knockout, which may find application in cancer therapy. Understanding the mechanism of palmitoylation of CD151 may give a wide scope for cancer research. Studying the role of CD151 in molecular mechanisms associated with self-renewal, differentiation, DNA damage response, epigenetic mechanisms and anchoragedependent and -independent tumor cell survival using gene silencing methods may provide an ample scope for future cancer research.

#### Abbreviations

CD151, cluster of differentiation 151; GP-27, glycoprotein-27; PETA-3, platelet-endothelial tetraspan antigen-3; TSPAN-24, tetraspanin-24; TEM, tetraspanin-enriched microdomains; ECM, extracellular membrane; MMP, matrix metalloproteinase.

#### **Author Contributions**

Conceived and designed the experiments: RM. Analyzed the data: SK. Wrote the first draft of the manuscript: GV. Contributed to the writing of the manuscript: AB. Agree with the manuscript results and conclusions: SK. Jointly developed the structure and arguments for the paper: RM. Made critical revisions and approved final version: RM and VRD. All authors reviewed and approved of the manuscript.

#### REFERENCES

- Orlowski E, Chand R, Yip J, et al. A platelet tetraspanin superfamily member, CD151, is required for regulation of thrombus growth and stability *in vivo*. *J Thromb Haemost*. 2009;7:2074–2084.
- 2. Sterk LM, Geuijen CA, Van Den Berg JG, Claessen N, Weening JJ, Sonnenberg A. Association of the tetraspanin CD151 with the laminin-binding integrins  $\alpha 3\beta 1$ ,  $\alpha 6\beta 1$ ,  $\alpha 6\beta 4$  and  $\alpha 7\beta 1$  in cells in culture and *in vivo. J Cell Sci.* 2002;115: 1161–1173.
- 3. Yauch RL, Berditchevski F, Harler MB, Reichner J, Hemler ME. Highly stoichiometric, stable and specific association of integrin  $\alpha 3\beta 1$  with CD151 provides a major link to phosphatidylinositol 4-kinase and may regulate cell migration. *Mol Biol Cell*. 1998;9:2751–2765.
- 4. Zhang XA, Bontrager AL, Hemler ME. TM4SF proteins associate with activated PKC and link PKC to specific  $\beta$ 1 integrins. J Biol Chem. 2001;276: 25005–25013.
- Zoller M. Tetraspanins: push and pull in suppressing and promoting metastasis. Nat Rev Cancer. 2009;9:40–55.
- Wang H-X, Li Q, Sharma C, Knoblich K, Hemler ME. Tetraspanin protein contributions to cancer. *Biochem Soc Trans*. 2011;39:547–552.
- Liu L, He B, Liu WM, Zhou D, Cox JV, Zhang XA. Tetraspanin CD151 promotes cell migration by regulating integrin trafficking. *J Biol Chem.* 2007;282: 31631–31642.
- Baleato RM, Guthrie PL, Gubler MC, Ashman LK, Roselli S. Deletion of CD151 results in a strain-dependent glomerular disease due to severe alterations of the glomerular basement membrane. *Am J Pathol.* 2008;173:927–937.
- Yoo SH, Lee K, Chae JY, Moon KC. CD151 expression can predict cancer progression in clear cell renal cell carcinoma. *Histopathology*. 2011;58(2):191–197.
- Whittock NV, McLean WH. Genomic organization, amplification, fine mapping, and intragenic polymorphisms of the human hemidesmosomal tetraspanin CD151 gene. *Biochem Biophys Res Commun.* 2001;281(2):425–430.
- Armoskus C, Moreira D, Bollinger K, Jimenez O, Taniguchi S, Tsai HW. Identification of sexually dimorphic genes in the neonatal mouse cortex and hippocamus. *Brain Res.* 2014;1562:23–38.
- 12. Hemler ME. Integrin-associated proteins. Curr Opin Cell Biol. 1998;10:578-585.
- Yang XH, Richardson AL, Torres-Arzayus MI, et al. CD151 accelerates breast cancer by regulating alpha 6 integrin functions, signalling, and molecular organization. *Cancer Res.* 2008;68(9):3204–3213.



- Geary SM, Cowin AJ, Copeland B, Baleato RM, Miyazaki K, Ashman LK. The role of the tetraspanin CD151 in primary keratinocyte and fibroblast functions: implications for wound healing. *Exp Cell Res.* 2008;314(11–12):2165–2175.
- Hasegawa M, Furuya M, Kasuya Y, et al. CD151 dynamics in carcinoma-stroma interaction: integrin expression, adhesion strength and proteolytic activity. *Lab Invest.* 2007;87(9):882-892.
- Yauch RL, Kazarov AR, Desai B, Lee RT, Hemler ME. Direct extracellular contact between integrin alpha (3) beta(1) and TM4SF protein CD151. *J Biol Chem.* 2000;275(13):9230–9238.
- Devbhandari RP, Shi GM, Ke AW, et al. Profiling of the tetraspanin CD151 web and conspiracy of CD151/integrin β1 complex in the progression of hepatocellular carcinoma. *PLoS One*. 2011;6(9):e24901.
- Sterk LM, Geuijen CA, Oomen LC, Calafat J, Janssen H, Sonnenberg A. The tetraspan molecule CD151, a novel constituent of hemidesmosomes, associates with the integrin alpha6 beta4 and may regulate the spatial organization of hemidesmosomes. *J Cell Biol.* 2000;149:969–982.
- Li Q, Yang XH, Xu F, et al. Tetraspanin CD151 plays a key role in skin squamous cell carcinoma. *Oncogene*. 2013;32(14):1772–1783.
- Romanska HM, Berditchevski F. Tetraspanins in human epithelial malignancies. J Pathol. 2011;223(1):4–14.
- Tokuhara T, Hasegawa H, Hattori N, et al. Clinical significance of CD151 gene expression in non-small cell lung cancer. *Clin Cancer Res.* 2001;7(12):4109–4114.
- Hashida H, Takabayashi A, Tokuhara T, et al. Clinical significance of transmembrane 4 superfamily in colon cancer. Br J Cancer. 2003;89(1):158–167.
- Zhu GH, Huang C, Qiu ZJ, et al. Expression and prognostic significance of CD151, c-Met, and integrin alpha3/alpha6 in pancreatic ductal adenocarcinoma. *Dig Dis Sci.* 2011;56(4):1090–1098.
- Suzuki S, Miyazaki T, Tanaka N, et al. Prognostic significance of CD151 expression in esophageal squamous cell carcinoma with aggressive cell proliferation and invasiveness. *Ann Surg Oncol.* 2010;18(3):888–893.
- Voss MA, Gordon N, Maloney S, et al. Tetraspanin CD151 is a novel prognostic marker in poor outcome endometrial cancer. BrJ Cancer. 2011;104(10):1611–1618.
- Zijlstra A, Lewis J, Degryse B, Stuhlmann H, Quigley JP. The inhibition of tumor cell intravasation and subsequent metastasis via regulation of *in vivo* tumor cell motility by the tetraspanin CD151. *Cancer Cell*. 2008;13(3):221–234.
- Sadej R, Romanska H, Baldwin G, et al. CD151 regulates tumorigenesis by modulating the communication between tumor cells and endothelium. *Mol Cancer Res.* 2009;7(6):787–798.
- Sadej R, Romanska H, Kavanagh D, et al. Tetraspanin CD151 regulates transforming growth factor beta signalling: implication in tumor metastasis. *Cancer Res.* 2010;70(14):6059–6070.
- 29. Takeda Y, Kazarov AR, Butterfield CE, et al. Deletion of tetraspanin CD151 results in decreased pathologic angiogenesis *in vivo* and *in vitro*. *Blood*. 2007;109(4): 1524–1532.
- Kohno M, Hasegawa H, Miyake M, Yamamoto T, Fujita S. CD151 enhances cell motility and metastasis of cancer cells in the presence of focal adhesion kinase. *Int J Cancer.* 2002;97:336–343.
- Haeuw JF, Goetsch L, Bailly C, Corvaia N. Tetraspanin CD151 as a target for antibody-based cancer immunotherapy. *Biochem Soc Trans.* 2011;39(2):553–558.
- Fei Y, Wang J, Liu W, et al. CD151 promotes cancer cell metastasis via integrins α3β1 and α6β1 in vitro. Mol Med Rep. 2012;6(6):1226–1230.
- Copeland BT, Bowman MJ, Ashman LK. Genetic ablation of the tetraspanin CD151 reduces spontaneous metastatic spread of prostate cancer in the TRAMP model. *Mol Cancer Res.* 2013;11(1):95–105.
- Shanmukhappa K, Kim JK, Kapil S. Role of CD151, a tetraspanin, in porcine reproductive and respiratory syndrome virus infection. *Virol J.* 2007;4(62):1–12.
- Gustafson-Wagner E, Stipp CS. The CD9/CD81 tetraspanin complex and tetraspanin CD151 regulate α3β1 integrin-dependent tumor cell behaviors by overlapping but distinct mechanisms. *PLoS One.* 2013;8(4):e61834.
- Chien CW, Lin SC, Lai YY, et al. Regulation of CD151 by hypoxia controls cell adhesion and metastasis in colorectal cancer. *Clin Cancer Res.* 2008;14: 8043–8051.

- Rao Malla R, Gopinath S, Alapati K, Gorantla B, Gondi CS, Rao JS. Knockdown of cathepsin B and uPAR inhibits CD151 and α3β1 integrin-mediated cell adhesion and invasion in Glioma. *Mol Carcinog.* 2012;52(10):777–790.
- Hong IK, Jin YJ, Byun HJ, Jeoung DI, Kim YM. Homophilic interactions of tetraspanin CD151 up-regulate motility and matrix metalloproteinase-9 expression of human melanoma cells through adhesion-dependent c-Jun activation signalling pathways. J Biol Chem. 2006;281:24279–24292.
- Shi GM, Ke AW, Zhou J, Wang XY, Xu Y. CD151 modulates expression of matrix metalloproteinase 9 and promotes neoangiogenesis and progression of hepatocellular carcinoma. *Hepatology*. 2010;52:183–196.
- Ke AW, Shi GM, Zhou J, Huang XY, Shi YH. CD151 amplifies signalling by integrin alpha6beta1 to PI3K and induces the epithelial-mesenchymal transition in HCC cells. *Gastroenterology*. 2011;140:1629–1641.
- Shi GM, Xu Y, Fan J, Zhou J, Yang XR. Identification of side population cells in human hepatocellular carcinoma cell lines with stepwise metastatic potentials. *J Cancer Res Clin Oncol.* 2008;134:1155–1163.
- Ke AW, Shi GM, Zhou J, Wu FZ, Ding ZB. Role of overexpression of CD151 and/or c-Met in predicting prognosis of hepatocellular carcinoma. *Hepatology*. 2009;49:491–503.
- Shigeta M, Sanzen N, Ozawa M, Gu J, Hasegawa H. CD151 regulates epithelial cell-cell adhesion through PKC- and Cdc42-dependent actin cytoskeletal reorganization. *J Cell Biol.* 2003;163:165–176.
- Sachs N, Claessen N, Aten J, et al. A blood pressure influences end-stage renal disease of CD151 knockout mice. J Clin Invest. 2012;(1):3.
- Nakashima Y, Yano M, Kobayashi Y, et al. Endostatin gene therapy on murine lung metastases model utilizing cationic vector-mediated intravenous gene delivery. *Gene Ther.* 2003;10(2):123–130.
- Takeda Y, Li Q, Kazarov AR, et al. Diminished metastasis in tetraspanin CD151—knockout mice. Am Soc Hematol. 2011;118(2):464–472.
- Yang XH, Flores LM, Li Q, et al. Disruption of laminin-integrin-CD151-focal adhesion kinase axis sensitizes breast cancer cells to ErbB2 antagonists. *Cancer Res.* 2010;70:2256–2263.
- Kwon MJ, Park S, Choi JY, et al. Clinical significance of CD151 overexpression in subtypes of invasive breast cancer. Br J Cancer. 2012;106(5):923–930.
- Palmer TD, Martínez CH, Vasquez C, et al. Integrin-free tetraspanin CD151 can inhibit tumor cell motility upon clustering and is a clinical indicator of prostate cancer progression. *Cancer Res.* 2014;74(1):173–187.
- Berditchevski F, Odintsova E. Characterization of integrin-tetraspanin adhesion complexes: role of tetraspanins in integrin signalling. *J Cell Biol.* 1999;146(2): 477–492.
- Romanska HM, Potemski P, Collins SI, Williams H, Parmar S, Berditchevski F. Loss of CD151/TSPAN24 from the complex with integrin α3β1 in invasive front of the tumour is a negative predictor of disease-free survival in oral squamous cell carcinoma. *Oral Oncol.* 2013;49(3):224–229.
- Yang YM, Zhang ZW, Liu QM, Sun YF, Yu JR, Xu WX. Overexpression of CD151 predicts prognosis in patients with resected gastric cancer. *PLoS One*. 2013;8(3):e58990.
- Tandon AK, Clark GM, Chamness GC, Chirgwin JM, McGuire WL. Cathepsin D and prognosis in breast cancer. N Engl J Med. 1990;322(5):297–302.
- Penas PF, Garcia-Diez A, Sanchez-Madrid F, Yánez-Mo M. Tetraspanins are localized at motility-related structures and involved in normal human keratinocyte wound healing migration. *J Invest Dermatol.* 2000;114(6):1126–1135.
- Funakoshi T, Tachibana I, Hoshida Y, et al. Expression of tetraspanins in human lung cancer cells: frequent downregulation of CD9 and its contribution to cell motility in small cell lung cancer. *Oncogene*. 2003;22(5):674–687.
- Le Naour F, André M, Greco C, et al. Profiling of the tetraspanin web of human colon cancer cells. *Mol Cell Proteomics*. 2006;5:845–857.
- Ferrer M, Yunta M, Lazo PA. Pattern of expression of tetraspanin antigen genes in Burkitt lymphoma cell lines. *Clin Exp Immunol.* 1998;113(3):346–352.