Future of Bone Repair

Rachana Somaiya and Ginpreet Kaur

SPP School of Pharmacy and Technology Management, SVKM's NMIMS, Mumbai, Maharashtra, India.

Libertas Academica

ABSTRACT: Bone can suffer from various conditions such as fractures and diseases such as osteoporosis, osteogenesis imperfect and tumors. Osteoporosis globally causes >8.9 million fractures each year. Current epidemiological data relevant to this and other diseases urge us to focus critically on promising and efficacious treatments for bone injury. Because of the limitations of conventional treatments for bone fracture, such as limited quantity for autograft, there is a demand to investigate better alternatives for bone healing. The main aim of this review is to highlight repair of bone injury, particularly focusing on several new research methods studied in preclinical trials and in vitro. New research methods such as low-level laser therapy, mesenchymal stem cell-based therapy, nanomaterials, biodegradable hydrogels, extracellular matrix-mimetic materials, and controlled delivery of growth factors from polymer scaffolds look promising for bone healing, and further clinical studies are suggested that use them in routine bone repair treatment in the near future.

KEYWORDS: fracture, bone healing, osteoblast, nanomaterials, growth factors, scaffolds

CITATION: Somaiya and Kaur. Future of Bone Repair. Bone and Tissue Regeneration Insights 2015:6 1–7 doi:10.4137/BTRI.S12333.

RECEIVED: July 30, 2014. RESUBMITTED: November 3, 2014. ACCEPTED FOR PUBLICATION: January 9, 2015.

ACADEMIC EDITOR: Kerstin Rolfe, Editor in Chief

TYPE: Review

FUNDING: Authors disclose no funding sources.

COMPETING INTERESTS: Authors disclose no potential conflicts of interest.

COPYRIGHT: © the authors, publisher and licensee Libertas Academica Limited. This is an open-access article distributed under the terms of the Creative Commons CC-BY-NC 3.0 License.

CORRESPONDENCE: ginpreet.aneja@gmail.com

Paper subject to independent expert blind peer review by minimum of two reviewers. All editorial decisions made by independent academic editor. Upon submission manuscript was subject to anti-plagiarism scanning. Prior to publication all authors have given signed confirmation of agreement to article publication and compliance with all applicable ethical and legal requirements, including the accuracy of author and contributor information, disclosure of competing interests and funding sources, compliance with ethical requirements relating to human and animal study participants, and compliance with any copyright requirements of third parties. This journal is a member of the Committee on Publication Ethics (COPE). Provenance: the authors were invited to submit this paper.

Published by Libertas Academica. Learn more about this journal.

Introduction

Biologically, bone is a living tissue made mostly of collagen and performs various functions within the body.¹ The collagen in the bone is a protein, which provides a soft framework, with minerals like calcium phosphate strengthening and hardening this framework.² Because of the combination of calcium phosphate and collagen, bone is strong and flexible enough to resist stress.³ Bone is also involved in the homeostatic regulation of ions in the circulating fluids of the body.⁴ It provides structural support and helps in maintaining the acid-base balance by absorbing or releasing alkaline salts that buffer the blood against excessive pH changes.⁵ Being a living tissue, bone needs a constant supply of nutrients and oxygen, and, therefore, there is a limit to the size of a defect or fracture it is able to restore to healthy working tissue.⁶ Consequently, bone can suffer from a number of pathological conditions like cancer, and is likely to degenerate as a result of aging and disease, for example, osteoporosis.7

Several bone diseases can occur, such as osteogenesis imperfecta, osteochondroma, osteoporosis, etc., and the most common occurrence is bone fracture. Bone fracture is a condition in which there is a disruption in the continuity of the bone. It can be a result of a large applied force or impact, or a marginal trauma injury as a result of some conditions that weaken the bones, such as bone cancer, osteogenesis imperfecta, or osteoporosis.

Classification of Bone Injury

Bone can suffer from various pathological conditions such as the following:

- Bone can develop infections and cancer;
- Osteoporosis and low bone density can make bones weak and liable to breaking;
- In osteogenesis imperfecta, bones become brittle;
- In Paget's disease, bone become weak;
- Other diseases of bone are caused by genetic factors, poor nutrition, or problems associated with bone growth rate or rebuilding, making the bone more easy to break.

Epidemiology

The prevalence of osteoporosis-related fractures includes aspects linked to the underlying osteoporosis and also those linked to injury, such as falling and age. The incidence of vertebral fracture increases sharply from 50 years of age and thereafter, whereas hip fracture increases from 70 years above.⁸ Most of the fractures are osteoporotic, where the risk of fracture increases with decrease in bone density, with the few exceptions being fractures of fingers, toes, and skull.9 Globally, osteoporosis causes >8.9 million fractures each year, resulting in an osteoporotic fracture every 3 seconds.¹⁰ Also, it is estimated that by the year of 2050 the global incidence of hip fracture will increase by 310% in men and 240% in women.¹¹ The progressive loss of bone and increased risk of falling are two of the factors responsible for the growing risk of fracture with advancing age. Though traumatic features, such as those related to motor vehicle accidents, could be more usual in people with lower bone mass, these are commonly not considered to be osteoporotic.¹² Also, it is observed that, in most parts of the world, women have a greater risk of fractures than men.⁸ One of the extremely important predictors of the

risk of future fractures is a history of prior fractures.¹³ A past record of vertebral fracture increases the risk of a following spine fracture by 5 times and occurrence of non-spine fracture by 2 times.^{14,15}

Pathophysiology

The process of bone fracture healing has three phases that enable the protection and proliferation of the areas surrounding dislocations and fractures. These three phases are 1) the reactive phase, 2) the reparative phase and 3) the remodeling phase.¹⁶ In the reactive phase, which happens after the fracture, there is presence of blood cells inside the tissues neighboring the injury site and this initial change is seen by electron and light microscope. After the fracture, bleeding stops as the blood vessels constrict. After a few hours, a blood clot, known as hematoma, is formed. All the cells within hematoma degenerate and die. Some of the cells that are adjacent to the site of injury but outside the blood clot also degenerate and die. The fibroblasts survive in this area and replicate. These fibroblasts build loose aggregates of cells scattered with small blood vessels, identified as the granulation tissue.¹⁷ Also, there is migration of the mesenchymal cells and ingrowth of vascular tissue. Then the reparative phase starts, where there is formation of cartilage callus and lamellar bone deposition.¹⁸ Many days after the fracture, periosteum cells start replicating and transforming. Some of these cells that are closest to the fracture gap start developing into chondroblasts that later on form the hyaline cartilage. The fibroblasts, which are present within granulation tissue, also develop into chondroblasts, further forming hyaline cartilage. These new tissues continue to grow in size till they start uniting with their counterparts from the other parts of fracture. Such processes culminate in the formation of a new mass of heterogeneous tissue, which is known as the fracture callus. Ultimately, the fracture gap is linked by woven bone and hyaline cartilage, and some of the original strength is restored. Subsequently, there is replacement of woven bone and hyaline cartilage with lamellar bone, and this process of replacement is called bony substitution with reference to woven bone and endochondral ossification with reference to hyaline cartilage. The formation of lamellar bone is seen after the collagen matrix of either tissue becomes mineralized. This mineralized matrix starts penetrating by channels, and each contains numerous osteoblasts and a microvessel. There is formation of new lamellar bone from the osteoblasts, which is in the form of the trabecular bone. Subsequently, the trabecular bone replaces the woven bone and cartilage of the original fracture callus. The compact bone substitutes the trabecular bone in the remodeling phase. This phase takes 3-5 years depending on factors such as the general condition and age.¹⁹ There are three main complications for fracture healing. In delayed union, there is infection or poor blood supply; in non-union, wound contamination or bone loss is found; and in fibrous union, there is improper immobilization.²⁰ Therefore, before initiating any new research idea for the treatment of bone injury, it is important to keep these complications in mind.

Diagnosis

A fracture or other bone injuries can be diagnosed based on the physical examination and the history given. To view the fractured bone, imaging by X-ray is performed. Also, in some situations where X-ray alone is not sufficient, CT scan or MRI is performed.

Conventional Treatments

The treatment for bone fracture is classified as surgical or conservative, where conservative treatment includes pain management, nonsurgical stabilization, and immobilization. For pain management for arm fractures in children, ibuprofen is as effective as a combination of codeine and acetaminophen.²¹ In immobilization, the fractured pieces of bones are aligned to their natural positions, called reduction, and X-ray is used to verify the position, but this method of treatment is very painful without anesthesia. Surgical methods are used only when conservative treatment fails, but they have their own risks and benefits. Due to the nature of recrudescence of bone infection, infection is particularly dangerous in bones. Bone tissue is extracellular matrix, and the few blood vessels required to maintain this low metabolism are able to bring only a limited number of immune cells to an injury site to fight the infection. Therefore, osteotomies and open fractures require prophylactic antibiotics and very careful antiseptic measures.²² Occasionally, bone grafting is also used, which is a surgical method that replaces the missing bone so as to repair fractures that are very complex. This procedure enhances bone's ability to regenerate itself by using methods such as autologous, allograft, or synthetic variants. But this method also has limitations like related chronic donor site pain and the limited quantity available for autograft.²³ These limitations of the conventional treatments urge us to look for more promising, newer ways of healing that will have a shorter treatment course (eg, single-pulsed electromagnetic field) and will accelerate the bone fracture healing process (eg, low-level laser therapy).

Potential Future Treatments

Several risks and disadvantages of conventional treatments have led to the development of new potential research ideas for bone injury repair and have been carried out in preclinical studies or in vitro. These new methods look promising and should be further studied to use them as a regular way of treatment for bone repair.

Extracellular Matrix-mimetic Biomaterials²⁴

Failure of implants such as bone screws, bone grafts, and arthroplasties is caused by the limited osseointegration of currently



SR. NO.	MATERIALS	PROPERTIES AND USES	STUDIED IN	ADVANTAGES	REFERENCES
1.	Extracellular matrix- mimetic biomaterials	Adhesive, short peptide sequences, modulates responses of host cell-implant material to augment implant osseointegration and formation of bone	In vitro	No low solubility/large cost to extract and purify as by use of full-length natural extracellular matrix polymers	24
2.	Controlled delivery of growth factors (BMP-2) from polymer scaffolds	Strong osteo-inductive activity, long half-life	Mouse calvarial defect model	Overcomes problems like high cost and protein stability, which are associated with growth factor-loaded scaffolds. Longer half-life	51
3.	Cell therapy (Cultured MSCs)	Can differentiate into osteo- genic cells	In vitro MSC culturing	Easily administered alone via percutaneous injection/ also can be implanted during an open surgery with bioma- terials. Large quantity can be obtained unlike the natural one that is available from iliac-crest	62
4.	Low-level Laser Therapy	Stimulation of cells and molecules of body, improves the expression of osteogenic factors	Rat calvarial cells, <i>in vitro</i> , rat tibia	Accelerates the bone frac- ture healing process, causes callus increase in bone mineral density and volume	66–73
5.	Incorporation of Platelet-rich plasma into PLGA/CPC composite scaffold	CPC is extremely osteocon- ductive, PLGA on the scaffold enhances the mechanical prop- erties of bioceramic scaffolds	Radial and femoral defects in rabbit model, <i>in vitro</i>	Enhances <i>in vitro</i> cell response like cell attach- ment, cell proliferation, and cell differentiation, improves bone formation and angiogenesis	74–80
5.	Biodegradable hydro- gels (glycol-chitosan)	Enzymatic degradability, aque- ous solubility, antibacterial activity biocompatibility	Mouse calvarial defect model	Locally releases BMP-2 in a bioactive form, by incorporating statin in the injectable gel, the bioactivity of hydrogel increases	81
6.	Administration of Sclerostin Antibody	Sclerostin neutralizing antibody exerts anabolic effect during bone fracture healing	Critical-sized femoral defect rat model, hindlimb- immobilization rat model, rat model of postmeno- pausal osteoporosis	Increases bone formation	82–98
7.	Nanomaterials	Nanostructured scaffolds regulates cell proliferation, cell differentiation, and migration, Nanomaterials increases sur- face area and wettability	Female adult sheep, noncritical-size calvarial defects in rabbits, rabbit subcutaneous and bilat- eral femoral defect model, calvarial bone defect rat model	Increased protein adsorption in comparison to conven- tional biomaterials	99–104
8.	Single-pulsed electromagnetic field	Osteogenic effects, improves bone repair and cell growth	<i>In vitro</i> , necrotic bone mice model	Shorter treatment course, short daily application, quick- ens osteogenic differentia- tion of hBMSCs	105–109

 Table 1. Summary of different new research methods of treatment for bone injury.

available orthopedic biomaterials, which present a great socioeconomic cost. The use of full-length natural extracellular matrix (ECM) polymers has several disadvantages, such as low solubility, high cost to extract and purify it in large quantities, suffering from batch-to-batch variation, and potentially suffering from immunogenicity. Additionally, it is a great challenge to alter, characterize, and maintain the presentation of natural ECM polymers. This emphasizes the need for adhesive biomaterials that mimic the extracellular matrix and which will modulate responses of host cell–implant material to augment implant osseointegration and formation of bone. Osteoprogenitors and osteoblasts play an important role in complex processes like host reactions to implants and bone remodeling. The protein signals identified by the osteoprogenitor and osteoblast receptors found on the implant surface significantly influence the host responses to implants. Only a few short peptide sequences out of the thousands of amino acids found in natural ECM polymers help in integrin recognition and binding sequences that trigger other responses like adhesion, signaling, and distribution. Because of this, short peptide sequences such as GFOGER,^{25,26} RGD,²⁷

PHSRN,28 REDV,29 and LDV30 with ECM-derived protein fragment such as FNIII7-10 are utilized to biofunctionalize bone tissue engineering scaffolds and titanium surfaces. Also, the structure and conformation of the ligand is a vital factor in their capacity to bind to the integrin receptors as well as to trigger the signaling pathways. Covalent immobilization and simple adsorption onto titanium surfaces are two of the common methods of protein fragment functionalization for titanium implants. RGD is an adhesive protein sequence that is found on many ECM molecules and can bind to several integrins, though for some integrins binding to RGD is greatly controlled by additional sequences like the PHSRN synergy site for $\alpha 5\beta 1.^{31,32}$ Many biomaterial approaches have used RGD as an adhesive ligand because it serves as a potent binding site. But according to different studies performed, RGD does not stimulate bone formation and bone repair in vivo.³³ Also, fibronectin-mimetic peptide fragments such as FNIII7-10 improve both osteoblast differentiation and adhesion strength.³⁴ Other ECM-derived proteins that have been found to improve osteoblast differentiation and adhesion strength in vitro include KRSR, which is a heparin binding site (HBP) located on several ECM proteins;35-40 FHRRIKA, which results from the HBP of bone sialoprotein (BSP);^{35,41-44} an osteopontin-derived peptide;⁴⁵ HBP12;⁴⁶ and the human vitronectin peptide.⁴⁷⁻⁵⁰ Though these ECM-derived protein fragments have shown potential as bone materials in vitro, many more studies need to be carried out to validate their osteogenic capacity in vivo also.

Controlled Delivery of Growth Factors from Polymer Scaffolds⁵¹

Natural bone repair process can be stimulated by using growth factors in bone tissue engineering.⁵² The use of a localized and sustained delivery approach can overcome problems like high cost and protein stability, which are associated with growth factor-loaded scaffolds.⁵³ Because of the controlled delivery profile of growth factors in scaffolds, cells can migrate to the area of the defect, proliferate, and differentiate, boosting tissue repair.⁵⁴ Bone morphogenetic proteins (BMPs), which are involved in maintaining differentiation processes of a variety of cells during fracture repair and skeletal development, are protein members of transforming growth factor-β superfamily.^{55,56} BMP-2 proteins have strong osteo-inductive activity, but they exhibit a short half-life in vivo of 7 minutes.⁵⁷⁻⁶¹ Therefore, sustained and controlled delivery of BMP-2 to use as scaffold has been suggested. It was observed that, on average, 70% of BMP-2 into the scaffold was released in a mouse calvarial defect model by the end of 3 weeks. BMP-2 was shown to be active, and there was substantial increase (55%) in the new bone volume. Conversely, only 31% increase in new bone volume was found in scaffolds without BMP-2 in comparison to empty defect controls, suggesting the potential of novel scaffolds for sustained and controlled BMP-2 delivery for bone-regeneration purposes.



Cell Therapy⁶²

Autologous bone grafting is preferred when the natural bone repair mechanism fails to work. Bone matrix and osteogenic cells in the graft provide the osteo-conductive and osteoinductive activity required for proper bone repair. Bone marrow mesenchymal stem cells (MSCs) are able to differentiate into osteogenic cells. Treatment by MSC-based cell therapy has shown potential to enhance bone repair. The quantity of MSCs that is available from iliac-crest aspirates is very small to be useful clinically. Therefore, either culture or concentration must be used to increase the MSC population.⁶³ These MSCs can be easily administered alone via percutaneous injection, or can be implanted during an open surgery with biomaterials.⁶⁴ Patients with avascular necrosis of femoral head or delayed repair of long bone fractures have shown encouraging preliminary results.⁶⁵ In vitro MSC culturing on specific biomaterials such as β-calcium triphosphate granules or biphasic hydroxyapatite is used to obtain colonization of the biomaterials and cell differentiation. After that, the biomaterial-cell construct is implanted into the zone that is to be treated. As there are challenges to promoting implant vascularization and increasing cell survival, much work still remains to be done before knowing that this method is appropriate for the regular filling of bone tissue defects.

Low-level Laser Therapy

Low-level laser therapy (LLLT) is a technique that supplies biostimulative light energy to cells of the body. The light that is absorbed leads to the stimulation of cells and molecules of the body.⁶⁶ LLLT has shown potential for its positive effects on fracture repair and bone metabolism.^{67–70} This therapy improves the expression of osteogenic factors in the bone repair process.⁷¹ Moreover, it also accelerates the bone fracture healing process and causes callus increase in bone mineral density and volume.^{72,73} Though LLLT seems to have great advantages, the biomechanical properties of bones do not show any improvement.⁷¹ Therefore, further research on the LLLT is suggested to prove its efficiency.

Incorporation of Platelet-rich Plasma into PLGA/CPC Composite Scaffold⁷⁴

Calcium phosphate cement (CPC) is extremely osteoconductive and biocompatible, as has been demonstrated by the rapid deposition of new bone on the CPC surface.^{75,76} Coating a polymer such as poly(lactic-*co*-glycolic acid) (PLGA) on the scaffold surface has proven to be a successful approach to enhance the mechanical properties of bioceramic scaffolds.^{77–80} The incorporation of platelet-rich plasma (PRP) into a PLGA/ CPC scaffold with unidirectional pore structure has shown positive effects in improving bone repair of radial and femoral defects in a rabbit model. It was observed that the introduction of PRP into PLGA/CPC scaffold enhanced in vitro cell responses such as cell attachment, cell proliferation, and cell differentiation. It also boosted bone formation and angiogenesis.

4



Therefore, this scaffold with a unidirectional pore structure seems to be a potential candidate for bone repair.

Biodegradable Hydrogels⁸¹

Hydrogels, which are biodegradable and injectable, have proven to be effective candidates as cell delivery vehicles to sustain tissue regeneration. Glycol-chitosan has several intrinsic properties such as enzymatic degradability, aqueous solubility, antibacterial activity, and biocompatibility. Because of such properties, glycol-chitosan is one of the most preferred natural scaffolds for bone tissue engineering. This gel has been observed to have the ability to locally release BMP-2 in a bioactive form to stimulate bone formation at the implantation site. Moreover, it has also been demonstrated that by incorporating statin in the injectable gel, the bioactivity of hydrogel increases. Therefore, the incorporation of statin in the injectable glycol-chitosan seems to be a potential way in the process of bone repair.

Administration of Sclerostin Antibody⁸²

The glycoprotein sclerostin is expressed by osteocytes and acts as a negative regulator of bone formation and osteoblast development.^{83,84} Sclerosteosis and Van Buchem disease are caused by mutations in the gene coding for sclerostin and are described by bone thickening and high bone mass due to amplified bone formation.⁸⁵⁻⁸⁸ Although the mechanism for inhibition of bone formation by sclerostin is still under investigation, it has been suggested that sclerostin inhibits the canonical Wnt signaling pathway and/or BMP pathway by modulating their receptors.⁸⁹⁻⁹⁴ The preclinical studies performed in models of osteoporosis, as well as a clinical trial, have shown that systemic administration of the sclerostin neutralizing antibody increases bone formation and prevents bone loss.^{95–98} It was also observed that systemic administration of the sclerostin neutralizing antibody leads to increased bone formation and enhances bone repair in a critical-sized femoral defect in a rat model.

Nanomaterials⁹⁹

Nanostructured scaffolds regulate cell proliferation, cell differentiation, and migration, resulting in the formation of functional tissues. They also provide cells with structural support. Nanomaterials possess unique properties such as increased surface area and wettability, which result in increased protein adsorption in comparison to conventional biomaterials. Nanocomposites such as collagen/hydroxyapatite (HA) has three-gradient multilayer scaffolds, which are made of assembled collagen fibers with/without HA. This nanocomposite finds application in the repair of osteochondrial defects, as has been demonstrated in female adult sheep.100 Also, the poly(lactic-co-glycolic acid)/tricalcium phosphate (PLGA/TCP) composite possesses unique properties like flexibility and mouldability, and has been shown to heal circular noncritical-size calvarial defects in rabbits.¹⁰¹ The poly(propylene fumarate)/propylene fumarate

diacrylate/carbon nanotube (PPF/PF-DA/CNT) composite has been demonstrated to heal rabbit subcutaneous and bilateral femoral defects.¹⁰² Nanomaterials such as poly-L-lactic acid (PLLA) as a nanofibrous scaffold have found application in healing a critical-sized calvarial bone defect in rats.¹⁰³ Nanofibrous PLLA membrane with a collagenous-guided bone renewal membrane, ie, a bilayer membrane, was able to heal a defect in the anteromedial cortex of the proximal tibia in rabbits.¹⁰⁴ Ultimately, novel strategies that combine nanoscale properties and various compositions could be established in the near future.

Single-pulsed Electromagnetic Field

Pulsed electromagnetic field (PEMF) has been confirmed to have osteogenic effects for treatment of bone fractures.¹⁰⁵⁻¹⁰⁸ But the main disadvantage of PEMF treatment is time utilization, which is a minimum of 10 h/day for the treatment duration, as suggested by the U.S. FDA (Federal Drug Administration). So there was a search for an efficient model for PEMF treatment. In a recent study, there was modification as a single-pulsed electromagnetic field (SPEMF), which required only a 3-minute daily treatment.¹⁰⁹ In an in vitro study, osteogenic differentiation and cell proliferation were observed in human bone marrow mesenchymal stem cells (hBMSCs), whereas in vitro revascularization and new bone formation were evaluated. There was no significant cytotoxic effect of SPEMF on hBMSCs in the in vitro study. Also, there was increase in osteogenic differentiation of hBMSCs after 3-7 days of treatment. Mineralization also increased after 10, 15, 20, and 25 days of SPEMF treatment. The study demonstrated that a 7-day short course has similar results on osteogenesis and proliferation as the 25-day SPEMF treatment.¹⁰⁹ This suggested that this novel SPEMF treatment quickens osteogenic differentiation of hBMSCs and improves bone repair and cell growth in the necrotic bone in mice. Therefore, there is a potential advantage of SPEMF due to shorter treatment course and short daily application. Also, it is a superior treatment to inductive coupling, where the disadvantage is the need for cooperation between the patient and the treating physician, as patient noncompliance may occur because of the heavy weight of the different units used in it. Additionally, SPEMF is also a better treatment than capacitive coupling, where the disadvantage that the units, although lightweight and small, can cause irritation to the skin from the electrodes.

Conclusion

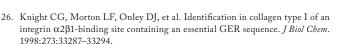
Bone repair is a complex process, and the current conventional methods have their own disadvantages. Due to the increasing prevalence of bone injuries, there is a need to find a better alternative for bone repair. The current new approaches are studied in animal models or in vitro studies, and have shown to be efficient by demonstrating several advantages over the conventional treatments. The advantages of potential future treatments, such as shorter treatment course, low cost, and increase in the rate of osteogenesis, will accelerate the bonehealing process and eventually decrease the prevalence of bone injuries. Therefore, future work needs to be done in direction of bone repair. And these new methods are to be studied in clinical trials as well to make them routine in the treatment of bone injuries.

Author Contributions

Conceived and designed the experiments: RS and GK. Analyzed the data: RS and GK. Wrote the first draft of the manuscript: RS and GK. Contributed to the writing of the manuscript: RS and GK. Agree with manuscript results and conclusions: RS and GK. Jointly developed the structure and arguments for the paper: RS and GK. Made critical revisions and approved final version: RS and GK. Both authors reviewed and approved of the final manuscript.

REFERENCES

- RHO JY. Mechanical properties and the hierarchical structure of bone. *Med Eng Phys.* 1998;20:92–102.
- Fei Y, Zhang M, Li M, et al. Element analysis in femur of diabetic osteoporosis model by SRXRF microprobe. *Micron*. 2007;38:637–642.
- Loreille OM, Diegoli TM, Irwin JA, Coble MD, Parsons TJ. High efficiency DNA extraction from bone by total demineralization. *Forensic Sci Int Genet*. 2007; 1:191–195.
- McLean FC. The ultrastructure and function of bone. *Science*. 1958;127:451–456.
 Green J, Kleeman CR. Role of bone in regulation of systemic acid-base balance. *Kidnev Int*. 1991:39:9–26.
- Ninikoski J. Hunt TK. Oxygen and healing wounds: tissue-bone repair enhancement. Handbook on Hyperbaric Medicine. Milan: Springer; 1996:485–507.
- 7. Wiita B. Osteoporosis: causes and consequences. Curr Sci. 1995;68:446-450.
- Lane NE. Epidemiology, etiology, and diagnosis of osteoporosis. Am J Obstet Gynecol. 2006;194:S3-S11.
- Seeley DG, Browner WS, Nevitt MC, Genant HK, Scott JC, Cummings SR. Which fractures are associated with low appendicular bone mass in elderly women? *Ann Intern Med.* 1991;115:837–842.
- Johnell O, Kanis JA. An estimate of the worldwide prevalence and disability associated with osteoporotic fractures. Osteoporosis Int. 2006;17:1726–1733.
- Gullberg B, Johnell O, Kanis JA. World-wide projections for hip fracture. Osteoporosis Int. 1997;7:407–413.
- Chau DL, Edelman SV, Chandran M. Osteoporosis and diabetes. Curr Diab Rep. 2003;3:37-42.
- Cummings SR, Nevitt MC, Browner WS, et al. Risk factors for hip fracture in white women. Study of Osteoporotic Fractures Research Group. N Engl J Med. 1995;332:767–773.
- Ross PD, Davis JW, Epstein RS, Wasnich RD. Pre-existing fractures and bone mass predict vertebral fracture incidence in women. *Ann Intern Med.* 1991; 114:919–923.
- Black DM, Arden NK, Palermo L, Pearson J, Cummings SR. Prevalent vertebral deformities predict hip fractures and new vertebral deformities but not wrist fractures. *J Bone Miner Res.* 1999;14:821–828.
- McKibbin B. The biology of fracture healing in long bones. J Bone Joint Surg Br. 1978;60:150.
- 17. Kalfas IH. Principles of bone healing. Neurosurg Focus. 2001;10:1-4.
- Ham AW, Harris WR. Repair and transplantation of bone. *Biochem Physiol Bone*. 2012;3:337.
- 19. Remedios A. Bone and bone healing. Vet Clin North Am Small Anim Pract. 1999;29:1029-1044.
- 20. Panagiotis M. Classification of non-union. Injury. 2005;36:S30-S37.
- Drendel AL, Gorelick MH, Weisman SJ, Lyon R, Brousseau DC, Kim MK. A randomized clinical trial of ibuprofen versus acetaminophen with codeine for acute pediatric arm fracture pain. *Ann Emerg Med.* 2009;54:553–560.
- Francel TJ, Birely BC, Ringelman PR, Manson PN. The fate of plates and screws after facial fracture reconstruction. *Plast Reconstr Surg.* 1992;90:568–573.
- Parikh SN. Bone graft substitutes: past, present, future. *JPostgrad Med*. 2002;48:142.
 Shekaran A, García AJ. Extracellular matrix-mimetic adhesive biomaterials for bone repair. *J Biomed Mater Res A*. 2011;6:261–272.
- Emsley J, Knight CG, Farndale RW, Barnes MJ. Structure of the integrin α2β1binding collagen peptide. J Mol Biol. 2004;335:1019–1028.



- Leahy DJ, Aukhil I, Erickson HP. 2.0 Å crystal structure of a four-domain segment of human fibronectin encompassing the RGD loop and synergy region. *Cell*. 1996;84:155–164.
- Aota SI, Nomizu M, Yamada KM. The short amino acid sequence Pro-His-Ser-Arg-Asn in human fibronectin enhances cell-adhesive function. J Biol Chem. 1994;269:24756–24761.
- Humphries MJ, Akiyama SK, Komoriya A, Olden K, Yamada KM. Identification of an alternatively spliced site in human plasma fibronectin that mediates cell type-specific adhesion. J Cell Biol. 1986;103:2637–2647.
- 30. Komoriya A, Green LJ, Mervic M, Yamada SS, Yamada KM, Humphries MJ. The minimal essential sequence for a major cell type-specific adhesion site (CS1) within the alternatively spliced type III connecting segment domain of fibronectin is leucine-aspartic acid-valine. *J Biol Chem.* 1991;266:15075–15079.
- Pytela R, Pierschbacher MD, Argraves S, Suzuki S, Ruoslahti E. Arginineglycine-aspartic acid adhesion receptors. *Methods Enzymol.* 1987;144:475–489.
- Redick SD, Settles DL, Briscoe G, Erickson HP. Defining fibronectin's cell adhesion synergy site by site-directed mutagenesis. J Cell Biol. 2000;149:521–527.
- Miljkovic ND, Cooper GM, Hott SL, et al. Calcium aluminate, RGD-modified calcium aluminate, and β-tricalcium phosphate implants in a calvarial defect. *J Craniofac Surg.* 2009;20:1538–1543.
- Petrie TA, Raynor JE, Reyes CD, Burns KL, Collard DM, García AJ. The effect of integrin-specific bioactive coatings on tissue healing and implant osseointegration. *Biomaterials*. 2008;29:2849–2857.
- 35. Schuler M, Hamilton DW, Kunzler TP, et al. Comparison of the response of cultured osteoblasts and osteoblasts outgrown from rat calvarial bone chips to nonfouling KRSR and FHRRIKA-peptide modified rough titanium surfaces. *J Biomed Mater Res B Appl Biomater*. 2009;91:517–527.
- Dee KC, Andersen TT, Bizios R. Design and function of novel osteoblast-adhesive peptides for chemical modification of biomaterials. *J Biomed Mater Res.* 1998;40: 371–377.
- Dettin M, Conconi MT, Gambaretto R, et al. Novel osteoblast-adhesive peptides for dental/orthopedic biomaterials. *J Biomed Mater Res.* 2002;60:466–471.
- Hasenbein M, Andersen TT, Bizios R. Micropatterned surfaces modified with select peptides promote exclusive interactions with osteoblasts. *Biomaterials*. 2002; 23:3937–3942.
- Nelson M, Balasundaram G, Webster TJ. Increased osteoblast adhesion on nanoparticulate crystalline hydroxyapatite functionalized with KRSR. Int J Nanomedicine. 2006;1:339.
- Balasundaram G, Webster TJ. Increased osteoblast adhesion on nanograined Ti modified with KRSR. J Biomed Mater Res A. 2007;80:602–611.
- Healy KE, Rezania A, Stile RA. Designing biomaterials to direct biological responses. Ann NY Acad Sci. 1999;875:24–35.
- Rezania A, Healy KE. Integrin subunits responsible for adhesion of human osteoblast-like cells to biomimetic peptide surfaces. JOrthop Res. 1999;17:615–623.
- Rezania A, Healy KE. Biomimetic peptide surfaces that regulate adhesion, spreading, cytoskeletal organization, and mineralization of the matrix deposited by osteoblast-like cells. *Biotechnol Prog.* 1999;15:19–32.
- Stile RA, Healy KE. Thermo-responsive peptide-modified hydrogels for tissue regeneration. *Biomacromolecules*. 2001;2:185–194.
- Shin H, Zygourakis K, Farach-Carson MC, Yaszemski MJ, Mikos AG. Attachment, proliferation, and migration of marrow stromal osteoblasts cultured on biomimetic hydrogels modified with an osteopontin-derived peptide. *Biomaterials*. 2004;25:895–906.
- Kim HE, Kim HW, Jang JH. Identification and characterization of a novel heparin-binding peptide for promoting osteoblast adhesion and proliferation by screening an *Escherichia coli* cell surface display peptide library. *J Pept Sci.* 2009; 15:43–47.
- 47. Dettin M, Bagno A, Morpurgo M, et al. Evaluation of silicon dioxide-based coating enriched with bioactive peptides mapped on human vitronectin and fibronectin: in vitro and in vivo assays. *Tissue Eng.* 2006;12:3509–3523.
- Bagno A, Piovan A, Dettin M, et al. Human osteoblast-like cell adhesion on titanium substrates covalently functionalized with synthetic peptides. *Bone*. 2007; 40:693–699.
- Cacchioli A, Ravanetti F, Bagno A, Dettin M, Gabbi C. Human vitronectin derived peptide covalently grafted onto titanium surface improves osteogenic activity: a pilot in vivo study on rabbits. *Tissue Eng Part A*. 2009;15:2917–2926.
- Dettin M, Bagno A, Gambaretto R, et al. Covalent surface modification of titanium oxide with different adhesive peptides: surface characterization and osteoblast-like cell adhesion. *J Biomed Mater Res A*. 2009;90:35–45.
- Rahman CV, Ben-David D, Dhillon A, et al. Controlled release of BMP-2 from a sintered polymer scaffold enhances bone repair in a mouse calvarial defect model. J Tissue Eng Regen Med. 2014;8:59–66.
- Ryoo HM, Lee MH, Kim YJ. Critical molecular switches involved in BMP-2-induced osteogenic differentiation of mesenchymal cells. *Gene.* 2006;366: 51–57.

6





- Putney SD, Burke PA. Improving protein therapeutics with sustained-release formulations. *Nat Biotechnol.* 1998;16:153–157.
- Issa JP, Bentley MV, Iyomasa MM, Sebald W, De Albuquerque RF. Sustained release carriers used to delivery bone morphogenetic proteins in the bone healing process. *Anat Histol Embryol.* 2008;37:181–187.
- Duguy N, Petite H, Arnaud E. Biomaterials and osseous regeneration. Ann Chir Plast Esthet. 2000;45:364–376.
- Groeneveld EH, Burger EH. Bone morphogenetic proteins in human bone regeneration. *Eur J Endocrinol.* 2000;142:9–21.
- Fujimura K, Bessho K, Kusumoto K, Ogawa Y, Iizuka T. Experimental studies on bone inducing activity of composites of atelopeptide type I collagen as a carrier for ectopic osteoinduction by rhBMP-2. *Biochem Biophys Res Commun.* 1995;208: 316–322.
- Boyne P, Marx RE, Nevins M, Lazaro E, Lilly Le AM, Nummikoski P. A feasibility study evaluating rhbmp-2/absorbable collagen sponge for maxillary sinus floor augmentation. *Int J Period Restor Dent*. 1997;17:11–25.
- Kusumoto K, Bessho K, Fujimura K, Akioka J, Ogawa Y, Iizuka T. Prefabricated muscle flap including bone induced by recombinant human bone morphogenetic protein-2: an experimental study of ectopic osteoinduction in a rat latissimus dorsi muscle flap. *Br J Plast Surg.* 1998;51:275–280.
- Okubo Y, Bessho K, Fujimura K, Iizuka T, Miyatake SI. Osteoinduction by bone morphogenetic protein-2 via adenoviral vector under transient immunosuppression. *Biochem Biophys Res Commun.* 2000;267:382–387.
- Poynton AR, Lane JM. Safety profile for the clinical use of bone morphogenetic proteins in the spine. *Spine*. 2002;27:S40–S48.
- Rosset P, Deschaseaux F, Layrolle P. Cell therapy for bone repair. Orthop Traumatol Surg Res. 2014;100:S107–S112.
- 63. Park IH, Micic ID, Jeon IH. A study of 23 unicameral bone cysts of the calcaneus: open chip allogeneic bone graft versus percutaneous injection of bone powder with autogenous bone marrow. *Foot Ankle Int.* 2008;29:164–170.
- Knight MN, Hankenson KD. Mesenchymal stem cells in bone regeneration. Adv Wound Care. 2013;2:306–316.
- Hernigou P, Poignard A, Zilber S, Rouard H. Cell therapy of hip osteonecrosis with autologous bone marrow grafting. *Indian J Orthop.* 2009;43:40–45.
- Ebrahimi T, Moslemi N, Rokn AR, Heidari M, Nokhbatolfoghahaie H, Fekrazad R. The influence of low-intensity laser therapy on bone healing. *J Dent.* 2012;9:238–248.
- Oliveira P, Sperandio E, Fernandes KR, Pastor FA, Nonaka KO, Renno A. Comparison of the effects of low-level laser therapy and low-intensity pulsed ultrasound on the process of bone repair in the rat tibia. *Rev Bras Fisioter*. 2011;15: 200–205.
- Renno AC, McDonnell PA, Parizotto NA, Laakso EL. The effects of laser irradiation on osteoblast and osteosarcoma cell proliferation and differentiation *in vitro*. *Photomed Laser Surg*. 2007;25:275–280.
- Luger EJ, Rochkind S, Wollman Y, Kogan G, Dekel S. Effect of low-power laser irradiation on the mechanical properties of bone fracture healing in rats. *Lasers* Surg Med. 1998;22:97–102.
- Ozawa Y, Shimizu N, Kariya G, Abiko Y. Low-energy laser irradiation stimulates bone nodule formation at early stages of cell culture in rat calvarial cells. *Bone*. 1998;22:347–354.
- Tim CR, Pinto KN, Rossi BR, et al. Low-level laser therapy enhances the expression of osteogenic factors during bone repair in rats. *Lasers Med Sci.* 2014;29: 147–156.
- Liu X, Lyon R, Meier HT, Thometz J, Haworth ST. Effect of lowerlevel laser therapy on rabbit tibial fracture. *Photomed Laser Surg.* 2007;25: 487–494.
- Lirani-Galvão AP, Jorgetti V, Da Silva OL. Comparative study of how lowlevel laser therapy and low-intensity pulsed ultrasound affect bone repair in rats. *Photomed Laser Surg.* 2006;24:735–740.
- 74. He F, Chen Y, Li J, et al. Improving bone repair of femoral and radial defects in rabbit by incorporating PRP into PLGA/CPC composite scaffold with unidirectional pore structure. J Biomed Mater Res A. 2014:00A:000–000.
- Ooms EM, Wolke JG, Van Der Waerden JP, Jansen JA. Trabecular bone response to injectable calcium phosphate (Ca-P) cement. *J Biomed Mater Res.* 2002;61:9–18.
- Comuzzi L, Ooms E, Jansen JA. Injectable calcium phosphate cement as a filler for bone defects around oral implants: an experimental study in goats. *Clin Oral Implants Res.* 2002;13:304–311.
- Chevalier J, Gremillard L. Ceramics for medical applications: a picture for the next 20 years. J Eur Ceram Soc. 2009;29:1245–1255.
- Zhang R, Ma PX. Poly (alpha-hydroxyl acids)/hydroxyapatite porous composites for bone-tissue engineering. I. Preparation and morphology. *J Biomed Mater Res.* 1999;44:446–455.
- Miao X, Tan DM, Li J, Xiao Y, Crawford R. Mechanical and biological properties of hydroxyapatite/tricalcium phosphate scaffolds coated with poly (lactic-co-glycolic acid). *Acta Biomater*. 2008;4:638–645.
- Zhao J, Guo LY, Yang XB, Weng J. Preparation of bioactive porous HA/PCL composite scaffolds. *Appl Surf Sci.* 2008;255:2942–2946.

- Brittain SB. Development and Characterization of a Bioactive Injectable Chitosan Hydrogel for Bone Repair. [Master's Theses]. Mansfield: University of Connecticut; 2013. [Paper 418].
- Virk MS, Alaee F, Tang H, Ominsky MS, Ke HZ, Lieberman JR. Systemic administration of sclerostin antibody enhances bone repair in a critical-sized femoral defect in a rat model. *J Bone Joint Surg.* 2013;95:694–701.
- Poole KE, van Bezooijen RL, Loveridge N, et al. Sclerostin is a delayed secreted product of osteocytes that inhibits bone formation. *FASEBJ*. 2005;19:1842–1844.
- Bezooijen RL, Dijke PT, Papapoulos SE, Löwik CW. SOST/sclerostin, an osteocyte-derived negative regulator of bone formation. *Cytokine Growth Factor Rev.* 2005;16:319–327.
- Van Buchem FS, Hadders HN, Ubbens R. An uncommon familial systemic disease of the skeleton: hyperostosis corticalis generalisata familiaris. *Acta Radiol.* 1995;44: 109–120.
- Balemans W, Patel N, Ebeling M, et al. Identification of a 52 kb deletion downstream of the SOST gene in patients with van Buchem disease. *J Med Genet*. 2002; 39:91–97.
- Balemans W, Ebeling M, Patel N, et al. Increased bone density in sclerosteosis is due to the deficiency of a novel secreted protein (SOST). *Hum Mol Genet*. 2001;10: 537–543.
- Beighton P, Barnard A, Hamersma H, Wouden A. The syndromic status of sclerosteosis and van Buchem disease. *Clin Genet.* 1984;25:175–181.
- Kamiya N, Ye L, Kobayashi T, et al. BMP signaling negatively regulates bone mass through sclerostin by inhibiting the canonical Wnt pathway. *Development*. 2008;135:3801–3811.
- ten Dijke P, Krause C, De Gorter DJ, Löwik CW, Van Bezooijen RL. Osteocytederived sclerostin inhibits bone formation: its role in bone morphogenetic protein and Wnt signaling. *J Bone Joint Surg.* 2008;90:31–35.
- Li X, Zhang Y, Kang H, et al. Sclerostin binds to LRP5/6 and antagonizes canonical Wnt signaling. J Biol Chem. 2005;280:19883–19887.
- van Bezooijen RL, Svensson JP, Eefting D, et al. Wnt but not BMP signaling is involved in the inhibitory action of sclerostin on BMP-stimulated bone formation. J Bone Miner Res. 2007;22:19–28.
- Löwik CW, Bezooijen RL. Wnt signaling is involved in the inhibitory action of sclerostin on BMP-stimulated bone formation. J Musculoskelet Neuronal Interact. 2006;6:357.
- Krause C, Korchynskyi O, de Rooij K, et al. Distinct modes of inhibition by sclerostin on bone morphogenetic protein and Wnt signaling pathways. J Biol Chem. 2010;285:41614–41626.
- Tian X, Jee WS, Li X, Paszty C, Ke HZ. Sclerostin antibody increases bone mass by stimulating bone formation and inhibiting bone resorption in a hindlimbimmobilization rat model. *Bone*. 2011;48:197–201.
- Li X, Warmington KS, Niu QT, et al. Inhibition of sclerostin by monoclonal antibody increases bone formation, bone mass, and bone strength in aged male rats. *J Bone Miner Res.* 2010;25:2647–2656.
- Li X, Ominsky MS, Warmington KS, et al. Sclerostin antibody treatment increases bone formation, bone mass, and bone strength in a rat model of postmenopausal osteoporosis. *J Bone Miner Res.* 2009;24:578–588.
- Padhi D, Jang G, Stouch B, Fang L, Posvar E. Single-dose, placebo-controlled, randomized study of AMG 785, a sclerostin monoclonal antibody. *J Bone Miner Res.* 2011;26:19–26.
- McMahon RE, Wang L, Skoracki R, Mathur AB. Development of nanomaterials for bone repair and regeneration. *J Biomed Mater Res Part B Appl Biomater*. 2013;101:387–397.
- McManus MC, Boland ED, Koo HP, et al. Mechanical properties of electrospun fibrinogen structures. *Acta Biomater*. 2006;2:19–28.
- Murugan R, Ramakrishna S. Development of nanocomposites for bone grafting. Compos Sci Technol. 2005;65:2385–2406.
- Woo KM, Chen VJ, Ma PX. Nano-fibrous scaffolding architecture selectively enhances protein adsorption contributing to cell attachment. *J Biomed Mater Res* A. 2003;67:531–537.
- Jang JH, Castano O, Kim HW. Electrospun materials as potential platforms for bone tissue engineering. *Adv Drug Deliv Rev.* 2009;61:1065–1083.
- Woo KM, Yu B, Jung HM, Lee YK. Comparative evaluation of different crystalstructured calcium sulfates as bone-filling materials. J Biomed Mater Res B Appl Biomater. 2009;91:545–554.
- Heckman JD, Ingram AJ, Loyd RD, Luck JV Jr, Mayer PW. Nonunion treatment with pulsed electromagnetic fields. *Clin Orthop Relat Res.* 1980;161:58–66.
- Friedenstein AJ, Piatetzky-Shapiro II, Petrakova KV. Osteogenesis in transplants of bone marrow cells. J Embryol Exp Morphol. 1966;16:381–390.
- Gossling HR, Bernstein RA, Abbott J. Treatment of ununited tibial fractures: a comparison of surgery and pulsed electromagnetic fields (PEMF). *Orthopedics*. 1992;15:711–719.
- McLeod KJ, Rubin CT. The effect of low-frequency electrical fields on osteogenesis. J Bone Joint Surg Am. 1992;74:920–929.
- Fu YC, Lin CC, Chang JK, et al. A novel single pulsed electromagnetic field stimulates osteogenesis of bone marrow mesenchymal stem cells and bone repair. *PLoS One.* 2014;9:e91581.

7