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Correlation Between Hepatocyte Growth Factor (HGF) and Gamma-Aminobutyric Acid (GABA) Plasma Levels in Autistic Children

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Abstract: There is much support for the role of Gamma-Aminobutyric acid (GABA) in the etiology of autism. Recent research has shown that hepatocyte growth factor (HGF) modulates GABAergic inhibition and seizure susceptibility. This study was designed to determine and correlate plasma levels of HGF, GABA, as well as symptom severity, in autistic children and neurotypical controls. Plasma from 48 autistic children and 29 neurotypical controls was assessed for HGF and GABA concentration using ELISAs. Symptom severity was assessed in these autistic individuals and compared to HGF and GABA concentrations. We previously reported that autistic children had significantly decreased levels of HGF. In this study, the same autistic children had significantly increased plasma levels of GABA (P = 0.002) and decreased HGF levels correlated with these increased GABA levels (r = 0.3; P = 0.05). High GABA levels correlated with increasing hyperactivity (r = 0.6; P = 0.0007) and impulsivity severity (r = 0.5; P = 0.007), tip toeing severity (r = 0.35; P = 0.03), light sensitivity (r = 0.4; P = 0.02), and tactile sensitivity (r = 0.4; P = 0.01). HGF levels did not correlate significantly with any symptom severity. These results suggest an association between HGF and GABA levels and suggest that plasma GABA levels are related to symptom severity in autistic children.

Keywords: GABA, HGF, autism, symptom severity

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Introduction

Autism spectrum disorders (ASDs) comprise a complex and heterogeneous group of pathological conditions characterized by impaired social interactions, deficits in verbal and nonverbal communication, and a limited interest in the surrounding environment associated with stereotyped and repetitive behaviors.¹

Evidence suggests that an impairment of GABAergic transmission contributes to the development of ASDs.^{2–5} While in the mature brain gammaaminobutyric acid (GABA) acts as an inhibitory transmitter, during the embryonic and the perinatal period, it depolarizes targeted cells and triggers calcium influx, regulating different developmental processes from cell proliferation migration, differentiation, synapse maturation, and cell death.⁶

Hepatocyte growth factor (HGF), an 82 kDa, 674 amino acid residue heterodimeric glycoprotein, was originally isolated from rat platelets.^{7,8} This growth factor has also been called scatter factor, hepatopoietin A, and mammary growth factor.⁹ It is one of a small family of factors lacking significant homology with other known growth factors, while including an HGF-like factor known as macrophage stimulating protein (MSP).^{10–13} HGF has mitogenic, morphogenic, and motogenic effects on hepatocytes, as well as endothelial, mesenchymal, and hematopoietic cell types,^{12,14,15} and demonstrates noticeable species cross-reactivity.¹⁶

HGF regulates cell growth, cell motility, and morphogenesis by activating a tyrosine kinase-signaling cascade after binding to the proto-oncogenic c-Met receptor (translated by the MET gene). HGF is secreted by mesenchymal cells and, although it was first considered to exert biological effects only on specific target cells, it has since been demonstrated to mediate inflammatory responses to tissue injury, and regulate cell growth, cell motility, and morphogenesis in a wide variety of cells. Its ability to stimulate branching morphogenesis, cell migration, survival, and proliferation gives it a central role in angiogenesis, tissue regeneration, as well as tumorogenesis.^{7–13}

Signaling by HGF has also been found to have anti-inflammatory, antifibrotic, and pro-regenerative activity on various types of tissue. But it seems to be particularly active in the nervous system, where it has been found to have neurotrophic and angiogenetic activity on central nervous system (CNS) neurons, promote both the survival of neurons and the regeneration of injured nerves, and function as a targetderived axonal chemoattractant, guiding axons to their target. As a result, it plays significant roles in the development of the CNS.¹⁷

GABA is the most abundant inhibitory neurotransmitter in the mammalian brain, where it is widely distributed.¹⁸ HGF has been shown to modulate GABAergic activity¹⁹ and enhance NMDA currents in the hippocampus.²⁰

Specific evidence to support HGF modulation of GABA activity is data associated with a mouse model of autism, the uPAR^{-/-} mouse, which displays a spatially selective defect in interneuron migration, such that the frontoparietal cortices of these mice show 50% less calbindin-positive interneurons (with a near absence of PV cells) whereas more caudal cortices are spared.^{21,22} These mice display autistic-like behaviors with increased anxiety and altered socialization, as well as interictal epileptiform EEG activity and an increased susceptibility to seizures.^{21,22} uPAR encodes a urokinase plasminogen activator (uPA) which is required for the proper processing of the HGF. In turn, HGF, through its receptor MET, has been shown to be a critical motogen for interneuron migration and is able to rescue the interneuron migration defect and seizure susceptibility of uPAR^{-/-} mice.^{19,23} Interestingly, polymorphisms in the MET promoter have recently been described to confer an increased susceptibility to autism and this gene is included in one of the genomic sequences linked to autism susceptibility (7q31).^{24,25}

Autism is a complex disorder and alterations in other GABAergic circuits, including the striatocortical circuits, likely contribute to this behavioral phenotype. Indeed, an interneuron-selective ablation of MET results in decreased cortical PV cells, but massively increased dorsal striatal PV interneurons, leading to a disruption in striatal-mediated procedural and reversal learning.²⁶ Nonetheless, cortical and hippocampal GABAergic deficits certainly play a role in some of the cognitive-behavioral manifestations of autism, as well as in the associated susceptibility to seizures.

The aim of this study was to determine and correlate plasma levels of HGF, GABA, as well as symptom severity, in autistic children and neurotypical controls.





Correlation between HGF and GABA in autistic children

Materials and Methods

Enzyme linked immunosorbest assay (ELISA) was used to measure serum HGF and GABA (ELISA kits, R&D Systems, Minneapolis, Minn. and USCN Life Sciences, Wuhan, China).

All reagents and specimens were equilibrated to room temperature before the assay was performed. A 1:51 dilution of the patient samples was prepared by mixing 10 μ L of the patient's sera with 0.5 mL of Serum Diluent. One hundred microliters of calibrators (20-200 Eu/mL antibodies), positive and negative control serums, serum diluent alone, and diluted patient samples were added to the appropriate microwells of a microculture plate (each well contained affinity purified polyclonal IgG to HGF or GABA). Wells were incubated for 60 min (±5 min) at room temperature, then washed 4 times with wash buffer. One hundred microliters of pre-diluter antihuman IgG conjugated with HRP was added to all microwells, incubated for 30 min (±5 min) at room temperature, then washed 4 times with wash buffer. One hundred microliters of enzyme substrate was added to each microwell. After approximately 30 min at room temperature, the reaction was stopped by adding 50 µL of 1M sulfuric acid. The wells were then read at 405 nm with an ELISA reader (BioRad Laboratories, Inc., Hercules, CA, USA).

Plasma

All plasma, experimental and control, were treated in an identical fashion. Whole blood was drawn into tubes containing EDTA, then spun at 1500 rpm for 10 min. Plasma was removed and immediately frozen until thawed for use in ELISAs.

Subjects

Experimental and control

Plasma from consecutive individuals diagnosed with autism (n = 48; 42 male; mean age 8 years) and neurotypical controls (n = 29; 20 male; mean age 9.5 years) were all obtained from patients presenting at the Health Research Institute/Pfeiffer Treatment Center over a two-year period beginning in October 2008. The autistic individuals meet the DSM-IV criteria and many were diagnosed using The Autism Diagnostic Interview-Revised (ADI-R) before presenting for treatment at the Pfeiffer Treatment Center, Warrenville, II. Patient consent was obtained from all patients involved in this study and this study was approved by the Institutional Review Board of the Health Research Institute/Pfeiffer Treatment Center.

Severity of disease

An autism symptom severity questionnaire was used to evaluate symptoms. The questionnaire (Pfeiffer Questionnaire) asked parents or caregivers to assess the severity of the following symptoms: Awareness, Expressive Language, Receptive Language (Conversational), Pragmatic Language, Focus, Attention, Hyperactivity, Impulsivity, Perseveration, Fine Motor Skills, Gross Motor Skills, Hypotonia (low muscle tone), Tip Toeing, Rocking/Pacing, Stimming, Obsessions/Fixations, Eye Contact, Sound Sensitivity, Light Sensitivity, and Tactile Sensitivity. The symptoms were rated by parents/guardians on a scale of 0-5 (5 being the highest severity) for each of these behaviors.

Statistics

Inferential statistics were derived from unpaired *t*-test and odds ratios with 95% confidence intervals (http://studentsttest.com). Pearson moment correlation test was used to establish degree of correlation between groups (http://www.wessa.net/rwasp_correlation.wasp#output).

Results

We previously reported that autistic children had significantly decreased levels of HGF.²⁷ In this study, Plasma from 48 autistic children and 29 neurotypical controls was assessed for GABA concentration using ELISAs. A random group of 29 of the above (N = 48) autistic children were also tested for HGF plasma concentration. Symptom severity was assessed in a random group of these autistic individuals and compared to HGF and GABA concentrations.

Autistic children in this study had significantly increased plasma levels of GABA (P = 0.002) (Fig. 1). Decreased HGF levels correlated with these increased GABA levels (r = 0.3; P = 0.05) (Fig. 4). High GABA levels correlated with increasing hyperactivity (r = 0.6; P = 0.0007) (Fig. 2), impulsivity severity (r = 0.5; P = 0.007) (Fig. 3), tip toeing severity (r = 0.35; P = 0.03), light sensitivity (r = 0.4; P = 0.02), and tactile sensitivity (r = 0.4; P = 0.01). Russo



Figure 1. Box and Whisker Plot: Plasma GABA concentration of autistic children significantly higher in autistic children compared to neurotypical, age-and gender similar controls.



Figure 2. GABA plasma concentration correlates (r = 0.6; P = 0.0007) with severity of hyperactivity in autistic children. (O) represents GABA and severity level of hyperactivity (0–5; 5 is most severe).



Figure 3. GABA plasma concentration correlates (r = 0.6; P = 0.007) with severity of impulsivity in autistic children. (O) represents GABA and severity level of impulsivity (0–5; 5 is most severe).



Figure 4. Decreased HGF levels correlate with increased GABA levels in autistic children (r = 0.3; P = 0.05).

HGF levels did not correlate significantly with any symptom severity (Table 1).

Discussion

Histological, biochemical, and molecular approaches have demonstrated altered levels and distribution of GABA and GABA receptors in peripheral blood and plasma, as well as in the brain, including decreased GABA-A receptors and benzodiazepine binding sites in the hippocampal formation.^{28–30}

Our data, demonstrating significantly high levels of GABA in the plasma of autistic children compared to controls, supports the work of others who have found elevated GABA levels in autistic children.²⁸

To our knowledge, this is the first study to show a correlation between these high GABA levels and low levels of HGF, and, although the number of patients with measured symptom severity is low, our data shows a causal relationship between GABA and hyperactivity and impulsivity, prevalent in autistic behavior.

We did not find any correlation between HGF levels and symptom severity. This may suggest an indirect role for HGF in the etiology of the disease. It is conceivable that altered uPA and its receptor, uPAR, displayed in animal behavioral models of autism, are associated with altered behavior unrelated to altered HGF levels.



	HGF	GABA
Symptom severity (0–5; 5 most	severe beha	avior)
r	-0.03	0.01
P	0.41	0.46
Expressive language	0.22	0.10
P	-0.22 0.1	0.19
Receptive language		
r P	-0.07 0.33	-0.14 0.24
Conversational language	0.00	0.21
r	-0.11	-0.018
P Focus/attention	0.20	0.40
r	0.14	0.328
<i>P</i> Hyperactivity	0.2	0.043
r	-0.1	0.57
P	0.27	0.0007
r	-0.01	0.46
P	0.47	0.007
Perseveration	0.15	0.13
P	-0.15 0.19	0.13
Fine motor skills		
r P	-0.05 0.37	-0.06 0.36
Gross motor skills	0.07	0.00
r	0.13	-0.3
P Hypotonia	0.22	0.06
r	-0.11	-0.11
P Tip tooing	0.25	0.28
r	-0.05	0.348
P	0.38	0.03
r	-0.02	-0.16
P	0.44	0.2
Hand flapping/finger stimming	0.00	0 407
r P	-0.06 0.34	0.187
Obsession/fixation	0.0.	•••••
r	-0.06	0.03
P Eve contact	0.35	0.43
ř.	0.06	-0.07
P Sound sensitivity	0.35	0.34
ſ	0.06	0.22
Р	0.34	0.12
	((Continued)

 Table 1. Relationship between symptom severity and GABA and HGF.

Table 1. (Continued)

	HGF	GABA
Light sensitivity		
r	0.09	0.39
Р	0.29	0.02
Tactile sensitivity		
r	0.002	0.415
Р	0.49	0.01

Notes: GABA concentration correlates with severity of focus/attention, hyperactivity, impulsivity, tip toeing, light sensitivity and tactile sensitivity (shaded data).

GABA is released by interneurons which contain the GABA synthesizing enzymes glutamic acid decarboxylase (GAD) 65 and GAD67, which convert the excitatory transmitter, glutamine, to GABA. GAD65 and GAD67 proteins are reduced in autistic parietal and cerebellar cortices³¹ and anti-GAD antibodies have been characterized in a pathogenetic model of autism involving Purkinje cell loss through.³² It is possible that specific altered behavior associated with hyperactivity is more closely associated with altered GAD, rather than the uPA/HGF system in autistic individuals.

GABA, the main inhibitory neurotransmitter in adulthood, is released by interneurons which contain the GABA synthesizing enzymes GAD65 and GAD67. GABAergic interneurons, which constitute a heterogeneous group of cells, differently classified in virtue of their anatomical, physiological, and molecular features,² represent only 10%-15% of the total neuronal population. Nevertheless, they provide the functional balance, complexity, and computational architecture of neuronal circuits.³ They play a key role in regulating neuronal excitability via feedback and feed-forward inhibition. Axons of different inhibitory cells target different postsynaptic subcellular compartments, allowing them to selectively control the output of pyramidal cells,⁴ thus providing the temporal structure that orchestrates the activity of neuronal ensembles leading to coherent network oscillations.5

Considering the many facets of GABA activities, particularly during development, it is not surprising that disturbance of GABAergic signaling can result in aberrant information processing, as found in neurodevelopmental disorders such as ASDs.

Evidence suggests that GABAergic mechanisms play a role in the etiology of ASD. Future studies on



the clinical relationship between GABA and behavior may provide the necessary data to formulate a coherent theory of GABA dysfunction. More trials of medication with known or suspected effects on GABA function are warranted.

Conclusions

We report a significant inverse relationship between plasma HGF and GABA levels in autistic children. This supports findings using a mouse model lacking the uPAR, demonstrating that decreased numbers of neocortical GABAergic interneurons would result in increased plasma GABA, spontaneous seizures, and reduced HGF.¹⁹

Although we found a significant relationship between HGF and GABA, we also found that the severity level of hyperactivity related symptoms correlated significantly with high levels of GABA, but none of the symptoms correlated with HGF levels. These results suggest that, despite the apparent regulatory effect of HGF on GABA levels, GABA levels, but not HGF levels, are associated with autistic behavior.

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Author Contributions

AR carried out the immunoassays, participated in the design of the study and performed the statistical analysis. AR conceived of the study, and participated in its design and coordination. AR drafted and approved the final manuscript.

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Competing Interests

Author(s) disclose no potential conflicts of interest.

Disclosures and Ethics

As a requirement of publication the author has provided signed confirmation of compliance with ethical and legal obligations including but not limited to compliance with ICMJE authorship and competing interests guidelines, that the article is neither under consideration for publication nor published elsewhere, of their compliance with legal and ethical guidelines concerning human and animal research participants (if applicable), and that permission has been obtained for reproduction of any copyrighted material. This article was subject to blind, independent, expert peer review. The reviewers reported no competing interests.

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