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Evolution of Type II Antifreeze Protein Genes in Teleost Fish: A Complex Scenario Involving Lateral Gene Transfers and Episodic Directional Selection

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Abstract: I examined hypotheses about lateral transfer of type II antifreeze protein (AFP) genes among "distantly" related teleost fish. The effects of episodic directional selection on amino acid evolution were also investigated. The strict consensus results showed that the type II AFP and type II antifreeze-like protein genes were transferred from *Osmerus mordax* to *Clupea harengus*, from the ancestral lineage of the *Brachyopsis rostratus—Hemitripterus americanus* clade to the ancestor of the *Hypomesus nipponensis—Osmerus mordax* group and from the ancestral lineage of *Brachyopsis rostratus—Hemitripterus americanus*—Hemitripterus americanus—Siniperca chuatsi—Perca flavescens to *Perca flavescens*. At the present time, the available evidence is more consistent with the LGT hypothesis than with other alternative explanations. The overall results indicate that evolutionary history of the type II AFP gene is complex, and that episodic directional selection was instrumental in the evolution of this freeze-preventing protein from a C-type lectin precursor.

Keywords: teleostei, antifreeze protein, lateral gene transfer, episodic directional selection

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Introduction

Many prokaryotes and eukaryotes that are found in cold environments survive freezing because their cells produce antifreeze proteins (AFPs).¹⁻⁹ These molecules function by either preventing endogenous fluids from freezing, or by enabling survival despite the freezing of these fluids. Based on structural differences, five different types of AFPs have been described for teleost fish.^{5,10,11} Type II AFP,^{5,11} shows a high degree of structural similarities among "distantly" related taxa, such as between Clupea harengus (herring) and the Osmerus mordax (rainbow smelt)-Hypomesus nipponensis (Japanese smelt) clade, Hemitripterus americanus (sea raven)-Brachyopsis rostratus (longsnout poacher) lineage, Siniperca chuatsi. C. harengus is a member of Ostarioclupeomorpha (order: Clupeiformes) whereas O. mordax (order: Osmeriformes), H. nipponensis (order: Osmeriformes), H. americanus (order: Scorpaeniformes), B. rostratus (order: Scorpaeniformes) and Siniperca chuatsi (order: Perciformes, see below) are classified in the Euteleosteomorpha (also known as Euteleostei) (see Fig. 1).12 According to Wiley and Johnson,¹² the infraclass Teleostei is divided into 4 major monophyletic taxa or cohorts, including Elopomorpha, Osteoglossomorpha, Ostarioclupeomorpha and Euteleosteomorpha (=Euteleostei).

Herring, rainbow and Japanese smelts produce type II AFPs that require calcium ions for their antifreeze activity (ie, Ca^{2+} -dependent) whereas type II AFPs produced by the sea raven and longsnout poacher do not (ie, Ca^{2+} -independent).^{5,11} Due to this,



it has been speculated that the calcium-dependent and independent type II AFPs have different mechanisms for preventing ice formation.⁵ It is also noteworthy that this group of freeze-preventing proteins, which are thought to have evolved from the sugar-binding domain of C-type (Ca²⁺-dependent) lectins, have 10 cysteine residues, giving rise to 5 disulfide bridges in identical positions.^{5,11,13,14} Five disulfide bridges are uniqueto the type II AFP, whereas C-type lectins typically form 2 to 3 disulfide bridges (see Fig. 1 by Graham and colleagues¹¹).

Evolutionary analyses have been conducted to further understand the remarkable structural similarity of the type II AFP genes in "distantly" related teleost fish species. One investigation, that was based on several independent pieces of evidence, proposed lateral gene transfer (LGT) as a possible explanation.11 However, the common origin and independent evolution (ie, convergent evolution) hypotheses can potentially also account for the taxonomic distribution/conservation of type II AFP genes. The common origin hypothesis holds that the type II AFP gene evolved once in the most recent common ancestor of all the type II AFP producing species, after which it remained highly static and was lost independently at least 3 different times.⁵ The convergent evolution hypothesis explains the taxonomic distribution of the similar type II AFP genes as being the result of at least three independent evolutionary events. In the current study, the notion of LGT was further tested using methods implemented in the program Prunier,¹⁵ and further discussed in light of the convergent evolution



Figure 1. A rooted species phylogeny showing 3 lateral gene transfers (LGTs).

Notes: Arrows indicate the direction of the gene transfers. The dotted lines highlight the uncertain phylogenetic positions of *D.rerio* and *C. myriaster* (see text for additional details). The cross hatch specifies the internal branch on which the cysteine in amino acid site 110 (see table 2) most likely evolved; that is, when assuming that the LGT events happened as shown here (see text for details).



and common ancestor hypotheses. In addition, effects of episodic directional selection on codon evolution, especially those sites coding for the 10 cysteines involved in disulfide bridge formation, were assessed using a mixed effects model of evolution (MEME)¹⁶ available on the datamonkey website.¹⁷ The results of the various analyses indicated that at least three lateral transfer events involving genes coding for the type II AFP/antifreeze-like protein (AFLP) have occurred, and that episodic directional selection has affected the evolution of the two cysteine-containing sites that are unique to the type II AFP.

Material and Methods

Sequences

The nucleotide sequences of all the taxa shown in Figure 2 in the paper by Liu and colleagues⁵ were downloaded from the GenBank, except for *Pimephales promelas*, for which no codon data was available (Table 1). To increase taxon sampling, nucleotide sequences of *Siniperca chuatsi* and *Perca flavescens*, which were determined through multiple NCBI blasts

to be similar to the type II AFPs of *C. harengus*, *O. mordax*, *H. nipponensis*, *H. americanus* and *B. rostratus*, were also included in the current study (Table 1). All these taxa are characterized by having 10 cysteine sites, except *P. flavescens*, which has only 9 cysteine sites, suggesting that it is uncertain whether this fish species produces a type II AFP. Due to this circumstance, this particular protein will here be referred to as a type II AFLP.

Data analyses

The software package DAMBE (Version 5.2.57)¹⁸ was implemented to manage the data and to match the codons against the aligned amino acid sequences. The alignments of the amino acid (Table 2), which are available upon request, were obtained using the MAFFT (Version 6)¹⁹ web server (available at http://mafft.cbrc.jp/alignment/server/). The alignment mode G-INS-i,²⁰ with an offset value of 0.2 and the MAFFT homolog function "turned on" (all the other parameter settings were default values), was used for the amino acids.



Figure 2. An unrooted maximum likelihood tree of the type II AFP/AFLPs and lectins.

Notes: The incongruent nodes (ie, bipartitions) between the gene and species trees are indicated by arrows. The values next to the nodes are LR-ELW edge supports. The scale bar shows the number of substitutions per nucleotide.



Table 1. Taxa and GenBank accession numbers/sourcereferences of the type II AFPs/AFLPs and lectins used inthe study.

Species	GenBank accession number/source references
Clupea harengus	L14722
Hypomesus nipponensis ³⁵	
Osmerus mordax	M96154
Brachyopsis rostratus	AB283044
Hemitripterus americanus	J05100
Siniperca chuatsi	EU719616
Perca flavescens	FJ826540
Oncorhynchys mykissA	AF363271
Oncorhynchys mykissB	AM041212
Salmo salarA	AY191312
Salmo salarB	CB504037
Spirinchus lanceolatus	AB114832
Rutilus rutilus	EG543056
Cyprinus carpio	AB034807
Carassius auratus	AY157616
Danio rerioA	NM_001245031
Danio rerioB	XM_003199234
Anguilla japonicaA	AB060539
Anguilla japonicaB	AB060538
Anguilla japonicaC	AB050703
Echidna delicatulaA	AB183016
Echidna delicatulaB	AB099491
Gymnothorax	AB099490
Tlavimarginatus Conger myriaster	AB239175

A species phylogeny was created, using the ITOL web server, based on the NCBI taxonomic classification (available at http://itol.embl.de/other trees.shtml) and the information reported by Liu and colleagues.⁵ In the resulting tree, the phylogenetic relationships of Rutilus rutilus, Danio rerio and Congermyriaster(outgroup lineage)were unresolved (Fig. 1). Since a resolved species topology is required for the LGT analyses, additional systematic information of the aforementioned species was used in an effort to infer their most likely phylogenetic positions in the teleost tree. Comprehensive phylogenetic studies of Cypriniformes, based on the mitochondrial genome data or/and multiple nuclear genes, indicated that *D. rerio* is more closely related to a large cyprinid clade, made up of taxa such as Leuciscinae than to the monophyletic group consisting of representatives of Cyprininae.^{21,22} However, the opposite relationship is also possible, that is that D. rerio is more closely related to the monophyletic group consisting of representatives of Cyprininae than to the large cyprinid clade, made up of taxa such as Leuciscinae.²¹ R. rutilus is considered to belong to the latter subfamily, whereas Cyprinus carpio and Carassius auratus are part of the former subfamily.^{21,23} No trees reported by Mayden and colleagues²¹ or Saitoh and colleagues²² suggested that R. rutilus (ie, the Leuciscinae) shares a more recent common ancestor with the clade composed of Cyprinus carpio and Carassius auratus than it does with the D. rerio lineage. It was also deduced from a study, using mitochondrial gene order information, that the phylogenetic position of C. myriaster is unresolved with to regard to Anguillidae (ie, Anguilla japonica) and to the Muraenidae (Echidna delicatula, Gymnothorax flavimarginatus) clades.²⁴ Consequently, based on the currently available phylogenetic studies of Rutilus rutilus, Danio rerio and Conger myriaster, it is not possible to come up with a single resolved species tree, but rather four different resolved species trees (Fig. 1). All 4 phylogenies were the used in the LGT analyses. The software package FigTree (available at http://tree. bio.ed.ac.uk/software/figtree/) was used to generate the trees shown in Figures 1 and 2.

Prunier (version 2.0),¹⁵ a program that works in conjunction with Treefinder,²⁵ was used to detect potential LGT events. Both the "slow" and the "fast" methods were implemented in this investigation. The "slow" method uses Kishino-Hasegawa²⁶ Shimodaira-Hasegawa²⁷ expected likelihood weights²⁸ and the approximately unbiased²⁹ tests to identify statistically significant topological differences between a gene and a species tree. LGT events are inferred in the part of the tree where statistically significant topological conflicts between the species and gene trees exist (see details by Abby and colleagues¹⁵). The gene tree was inferred, in both the "fast" and "slow" Prunier analyses, based on the nucleotide alignment and the model GTR + G8 + I. The "fast" method works by finding a maximum statistical agreement forest (MSAF) between the gene and species trees (see details by Abby and colleagues¹⁵). MSAF is defined as the minimum number of branches that are required to be eliminated in order to obtain statistical agreement between the two topologies.¹⁵ LGT events were inferred in situations in which branches with statistically significant support (ie, support of 95% or higher) must be cut in order to achieve MSAF.







Note: The numbers specify the amino acid sites (see text for additional information) and (*) shows sites with no variability across taxa.

The "fast" Prunier method calculates Expected-Likelihood Weights applied to Local Rearrangements (LR-ELW) edge support values²⁸ (see Fig. 2) for the gene tree. The aligned codon sequences and the four different species phylogenies (see previous paragraph) were provided as input for all Prunier analyses. For the "slow" method the default parameter values were used. In the "fast" analyses, the following parameters were assigned values that were different from the default settings: boot.thresh.conflict = 95 (ie, cut-off support value for topological conflict); fwd.depth = 1 (ie, maximal depth at which Prunier looks forward to find a significant LGT event when the current LGT is not significant).

A MEME¹⁶ was implemented for the purpose of detecting possible episodic directional selection on codon sites that has occurred in a small number of branches in the type II AFP gene tree. MEME models variable dN/dS across lineages at a given codon site, such that a certain fraction of branches are allowed

to evolve neutrally or under negative selection, whereas the remaining proportion is permitted to evolve under episodic directional selection. To test for evidence of this type of selection, a likelihood ratio test is performed between the aforementioned model and the nested null model that forces parameter values for episodic directional selection to vary between 0 and 1. Prior to running the MEME analysis the codon alignments of type II AFP were screened for recombination events using a program called GARD³⁰ with the following settings: Site-tosite variation = general discrete and rate classes = 4. Recombination has the potential to confound inferences of codon selection and, consequently, needs to be accounted for before running the analyses. No significant break points in the current alignment were detected by the GARD algorithm. The model selection tool, available in Datamonkey,¹⁷ chose the following optimal model for the MEME analysis: 001102 with AIC of 15118.9.

Results

Species and gene trees

The rooted species phylogeny, generated based on the NCBI taxonomic classification and published systematic studies of teleost fish, is shown in Figure 1. As noted in the material and methods section, the phylogenetic positions of *R. rutilus, D. rerio* and *C. myriaster* (outgroup lineage) are, at the present time, uncertain. The dotted lines in Figure 1 show the most probable relationships of the aforementioned species. Due to these uncertainties, 4 possible resolved species phylogenies were generated for the Prunier analysis.

Figure 2 shows the unrooted gene tree that was derived from the aligned nucleotide data in the 4 Prunier runs. A number of major conflicts, indicated by the arrows in Figure 2, are apparent between the gene and species trees. In the former, all of the type II AFP and AFLP producing taxa form a monophyletic group, which is not the case in the latter, in which C. harengus constitutes a deep branch within the Ostarioclupeomorpha clade and the H. nipponensis-O. mordax cluster share a most recent common ancestor with the S. salar-O. mykiss-S. lanceolatus lineage. Moreover, P. flavescens and S. chuatsi show a sister relationship in the species tree but not in the gene tree, in which they form a paraphyletic group. The outgroup, or the Elopomorpha clade, is similar in both topologies, except that the phylogenetic position of C. myriaster is unresolved in the species tree. The S. salar—O. mykiss—S. lanceolatus lineage share a common ancestor with Ostarioclupeomorpha in the genetreebutnotinthespeciesphylogeny. The branching order of the taxa within Ostarioclupeomorpha are resolved in the gene tree but not in the species tree.

Lateral gene transfer

The results of the Prunier runs differed somewhat depending on the species phylogeny and method used. The solid arrows, which indicate the direction of LGTs (Fig. 1), constitute the strict consensus results obtained from all 8 analyses; that is, when implementing 4 different species trees for each method (ie, either the "fast" or "slow" method). These events included the transfer of type II AFP gene from *O. mordax* to *C. harengus*, from the ancestral lineage of the *B. rostratus*—*H. americanus* clade to the ancestor of



the *H. nipponensis—O. mordax* group and from the ancestral lineage of B. rostratus-H. americanus-S.chuatsi-P. flavescens to P. flavescens (Fig. 1). Because of the low support value (ie, 47.6%) for the node uniting O. mordax and C. harengus (Fig. 2), the relationship among O. mordax, C. harengus, and *H. nipponensis* can be interpreted as being unresolved. Due to this circumstance, one can consider either O. mordax or H. nipponensis as possible donors (the specific donor being unresolved) of the type II AFP gene to C. harengus. Four out of the 8 Prunier runs suggest that R. rutilus has received the C-type lectin gene from the ancestral lineage of the Cypriniformes clade within Ostarioclupeomorpha. This scenario was obtained from both the "fast" and "slow" analyses; that is, when the input species tree in which R. rutilus is the sister lineage of D. rerio (Fig. 1) was used. There are two versions of this phylogeny because C. myriaster most likely constitutes a sister lineage of either the Anguillidae or the Muraenidae (see Fig. 1).²⁴ When one of these two possible species phylogenies was used in conjunction with the "slow" method, the E. delicatula—G. flavimarginatus clade received the C-lectin gene from the A. japonica-C. myriaster cluster.

Episodic directional selection

The MEME¹⁶ analysis showed evidence for 16 codon/amino acid sites in the alignment having been influenced by episodic directional selection (significance level = 0.05) (see Tables 2 and 3). Two of them are cysteine-containing sites (ie, sites 71 and 110; see Table 2) that are involved in forming the unique 5th disulfide bridge of the type II AFP.^{5,11}

Discussion

Lateral gene transfer and the evolution of the type II AFP

The evolutionary history of the type II AFP gene appears to be rather complex, involving at least 3 events of LGT.¹¹ When it comes to improving our understanding of the evolution of this freezepreventing protein, a key question involves the point in its phylogeny at which the 5th disulfide bridge evolved. The number of disulfide bridges in the C-type lectins and/or type II AFLPs vary quite a bit but are always less than 5.¹¹ Thus, The 5th disulfide bond, in addition to the other 4 disulfide bridges, makes the



α	β-	Pr[β = β⁻]	β+	$\Pr[\beta = \beta^+]$	<i>P</i> -value	q-value
0.71267	0	0.87743	33.051	0.12257	0.0357189	0.446487
0.594053	0.219727	0.83171	8.61427	0.16829	0.0427582	0.493364
0.592732	0.0233223	0.499489	7.0793	0.500511	0.0111048	0.23796
0.626032	0.153589	0.886189	208.891	0.113811	0.0471447	0.441982
0.578003	0	0.743857	3.43805	0.256143	0.0465647	0.465647
0.134445	0	0.846411	32.9029	0.153589	9.0959e-05	0.0136439
0.706383	0.043358	0.572077	7.49808	0.427923	0.0137215	0.257279
1.14485	0	0.823905	8.58932	0.176095	0.044903	0.481103
5e-09	5e-09	0.694404	8.03456	0.305596	0.000501674	0.0376255
5e-09	2. 5e-17	0.589632	3.53825	0.410368	0.0140873	0.234789
0.608495	0	0.854247	10.7894	0.145753	0.00485977	0.242988
0.320033	0.320033	0.938072	40.1285	0.061928	0.0289885	0.434828
0.330462	0.282797	0.966787	190.886	0.033213	0.0102668	0.25667
5e-09	2. 5e-17	0.725722	7.7696	0.274278	0.00678347	0.25438
0.934956	0.871756	0.913601	222.694	0.086399	0.0303443	0.413787
0.637836	0.00500758	0.600822	6.96584	0.399178	0.0100967	0.3029
	α 0.71267 0.594053 0.592732 0.626032 0.578003 0.134445 0.706383 1.14485 5e-09 5e-09 0.608495 0.320033 0.330462 5e-09 0.934956 0.637836	α $β^-$ 0.7126700.5940530.2197270.5927320.02332230.6260320.1535890.57800300.13444500.7063830.0433581.1448505e-095e-095e-092.5e-170.60849500.3200330.3200330.3304620.2827975e-092.5e-170.9349560.8717560.6378360.00500758	α $\beta^ \Pr[\beta = \beta^-]$ 0.7126700.877430.5940530.2197270.831710.5927320.02332230.4994890.6260320.1535890.8861890.57800300.7438570.13444500.8464110.7063830.0433580.5720771.1448500.8239055e-095e-090.6944045e-092.5e-170.5896320.60849500.8542470.3200330.3200330.9380720.3304620.2827970.9667875e-092.5e-170.7257220.9349560.8717560.9136010.6378360.005007580.600822	α $\beta^ \Pr[\beta = \beta^-]$ β^+ 0.7126700.8774333.0510.5940530.2197270.831718.614270.5927320.02332230.4994897.07930.6260320.1535890.886189208.8910.57800300.7438573.438050.13444500.84641132.90290.7063830.0433580.5720777.498081.1448500.8239058.589325e-095e-090.6944048.034565e-092.5e-170.5896323.538250.60849500.85424710.78940.3200330.3200330.93807240.12850.3304620.2827970.966787190.8865e-092.5e-170.7257227.76960.9349560.8717560.913601222.6940.6378360.005007580.6008226.96584	α $\beta^ \Pr[\beta = \beta^-]$ β^+ $\Pr[\beta = \beta^+]$ 0.7126700.8774333.0510.122570.5940530.2197270.831718.614270.168290.5927320.02332230.4994897.07930.5005110.6260320.1535890.886189208.8910.1138110.57800300.7438573.438050.2561430.13444500.84641132.90290.1535890.7063830.0433580.5720777.498080.4279231.1448500.8239058.589320.1760955e-095e-090.6944048.034560.3055965e-092.5e-170.5896323.538250.4103680.60849500.85424710.78940.1457530.3200330.3200330.93807240.12850.0619280.3304620.2827970.966787190.8860.0322135e-092. 5e-170.7257227.76960.2742780.9349560.8717560.913601222.6940.0863990.6378360.005007580.6008226.965840.399178	α $\beta^ \Pr[\beta = \beta^-]$ β^+ $\Pr[\beta = \beta^+]$ $P\text{-value}$ 0.7126700.8774333.0510.122570.03571890.5940530.2197270.831718.614270.168290.04275820.5927320.02332230.4994897.07930.5005110.01110480.6260320.1535890.886189208.8910.1138110.04714470.57800300.7438573.438050.2561430.04656470.13444500.84641132.90290.1535899.0959e-050.7063830.0433580.5720777.498080.4279230.01372151.1448500.8239058.589320.1760950.0449035e-095e-090.6944048.034560.3055960.0005016745e-092.5e-170.5896323.538250.4103680.01408730.60849500.85424710.78940.1457530.004859770.3200330.3200330.93807240.12850.0619280.02898850.3304620.2827970.966787190.8860.0332130.01026685e-092.5e-170.7257227.76960.2742780.006783470.9349560.8717560.913601222.6940.0863990.03034430.6378360.005007580.6008226.965840.3991780.0100967

Table 3, Codon (amino acid) sites found to b	pe under episodic	diversifying selection
	unning agia			

Notes: Codon = codon (or amino acid) number in the alignment; *alpha* = the maximum likelihood estimate (MLE) of the synonymoys substitution rate *alpha*; β^- = the maximum likelihood estimate (MLE) of the non-synonymous rate for the branch class with β *smaller or equal to alpha*; $\Pr[\beta = \beta^-]$ = the MLE of the proportion of sites evolving at β^- ; β^+ = the MLE of the unconstrained β non-synonymous rate; $\Pr[\beta = \beta^+]$ = the MLE of the proportion of sites evolving at β^+ ; *P-value* = the probability value for the likelihood ratio test statistic for β^+ = *alpha* (null) versus β^+ *unrestricted* (alternative); *q*-value = false discovery rate under the strict neutral null.

type II AFPs unique relative to the closely related C-type lectins and/or type II AFLPs.^{5,11} The evolution of the 5th disulfide bridge requires the appearance of a cysteine in amino acid position 110 (Table 2 and Fig. 1), provided that it already existed in site 71. In fact, C. harengus, O. mordax, H. nipponensis, H. americanus, B. rostratus and S. chuatsi have this amino acid present in both of these sites, whereas P. flavescens does not (ie, only in site 71 but not in 110) (Table 2), suggesting that P. flavescens does not produce a protein with the 5th disulfide bridge. When optimizing the interspecific amino acid variation in site 110 (Table 2) on the species tree in Figure 1, it becomes evident-under the assumption of 3 LGT events-that the cysteine evolved in this position once in the ancestral lineage of the B. rostratus-H. americanus—S. chuatsi—P. flavescens clade. Consequently, the occurrence of the 10 cysteines and the 5th disulfide bridge in the type II AFPs of C. harengus, O. mordax and H. nipponensis, and the absence of it in P. flavescens, can be explained by 3 separate LGT events. However, it should be noted that this scenario assumes that P. flavescens received a type II AFLP or C-type lectin gene, through LGT, from the ancestral lineage of the B. rostratus—H. americanus— S. chuatsi-P. flavescens clade before the type II AFP gene evolved (Fig. 1). This observation further

suggests that the evolution of the 9th and 10th cysteine were temporally displaced, such that the former (ie, site 71) appeared prior to latter (ie, in site 110).

Based on the currently available data, it is not possible to determine-under the assumption of 3 LGTs-whether the calcium-dependent type II AFP gene evolved prior to the calcium-independent one or vice versa. Thus, two equally mostparsimonious scenarios, requiring a minimum of two changes when optimized on the species tree (Fig. 1), are possible. In the first situation the calciumdependent type II AFP gene evolved first and was subsequently transferred from the ancestral lineage of the B. rostratus-H. americanus-S. chuatsi-P. flavescens cluster to that of the H. nipponensis— O. mordax group (Fig. 1). This scenario assumes that calcium-independence evolved after the aforementioned transfer event (Fig. 1). In the second situation the calcium-independent type II AFP gene evolved first and was subsequently transferred from the ancestor of the B. rostratus-H. americanus-S. chuatsi-P. flavescens clade to that of the *H. nipponensis—O. mordax* group. This interpretation assumes that the calcium-dependent type II AFP gene evolved in the ancestral lineage of H. nipponensis-O. mordax; that is, after it had been received from presumed donor.

In addition to the LGT scenario, two other hypotheses can be invoked to explain the distribution/ conservation of type II AFP genes in the taxa included in the current study. Those are the common origin and the independent evolution (ie, convergent evolution) hypotheses. The common origin hypothesis, which is advocated by Liu and colleagues,⁵ holds that the type II AFP gene evolved once in the most recent common ancestor of all the type II AFP producing species after which it was lost independently at least 3 different times.⁵ To explain the remarkable taxonomic conservation/distribution and the evolution of the calcium-independent type II AFP gene, the common origin hypothesis assumes two independent losses of calcium-binding sites⁵ and that the gene has remained highly static since it evolved about 280 million years ago (ie, the estimated divergence time between Ostarioclupeomorpha and Euteleostei³¹). This is especially true with regard to the C. harengus and O. mordax divergence. Given the large amount of time that has passed since its origin in the common ancestor (approximately 280 million years ago), the introns of the type II AFP genes in C. harengus and O. mordax/ H. nipponensis are expected to have differentiated to a degree similar to that seen in, for instance, the intron of the conserved spliceosomal protein gene (Prp8p). Graham and colleagues¹¹ have shown that the former sequences are much more conserved than the latter. This observation, in addition to the assumption that the type II AFP gene has remained largely unchanged for over 200 million years, makes the common origin hypothesis a less likely explanation at this time for the distribution/ conservation of the type II AFP genes among the taxa included here.

The convergent evolution hypothesis explains the taxonomic distribution/conservation of the type II AFP genes analyzed here as being the result of a number of independent origination events. Independent evolution of a gene in "distantly" related lineages can sometimes be difficult to discern from LGT because both processes can give rise to sequences with similar nucleotide compositions. However, transferred nucleotide sequences (ie, as opposed to convergent evolution) are more likely to show distinctive evolutionary patterns relative to the other genes (regions) in the genome of the putative recipient species, while at the same time exhibiting substitution



Episodic directional selection

The MEME analysis showed that the cysteine containing sites 71 and 110 (Table 2), which are responsible for the formation of the 5th disulfide bridge, have been affected by episodic directional selection (Table 3). This result suggests that this mechanism was instrumental in driving the evolution of the type II AFP gene from a C-type lectin precursor. Such an evolutionary event is highly likely to have been adaptive in nature since directional positive selection is sometimes thought to be associated with adaptive (functional) changes in protein-coding genes;^{32,33} that is, by favoring fitnessenhancing mutations. An earlier study suggested that episodic positive selection also was important in driving the differentiation of duplicated antifreeze protein genes in two Fragilariopsis species after the ancestral lineage acquired the sequence from the basidiomycetes.³³ It is also noteworthy that none of the ice-binding sites (here Thr104 and Thr106)





and Ca⁺-coordinating residues (here Asp102 and Glu109) identified in the *C. harengus* type II AFP by Graham and colleagues⁵ showed any evidence of having been influenced by episodic directional selection (Table 2).

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Author Contributions

Conceived and designed the experiments: US. Analyzed the data: US. Wrote the first draft of the manuscript: US. Contributed to the writing of the manuscript: US. Agree with manuscript results and conclusions: US. Jointly developed the structure and arguments for the paper: US. Made critical revisions and approved final version: US.

Disclosures and Ethics

As a requirement of publication author(s) have provided to the publisher signed confirmation of compliance with legal and ethical obligations including but not limited to the following: authorship and contributorship, conflicts of interest, privacy and confidentiality and (where applicable) protection of human and animal research subjects. The authors have read and confirmed their agreement with the ICMJE authorship and conflict of interest criteria. The authors have also confirmed that this article is unique and not under consideration or published in any other publication, and that they have permission from rights holders to reproduce any copyrighted material. Any disclosures are made in this section. The external blind peer reviewers report no conflicts of interest.

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