

Is the Adipose Tissue the Key Road to Inflammation?

Stéphanie Lucas, Claudie Verwaerde and Isabelle Wolowczuk

Laboratoire de NeuroImmunoEndocrinologie Institut Pasteur de Lille, BP447 and IFR 142 1, rue A. Calmette Lille, F-59019, France.

Abstract: It is now broadly accepted that white adipose tissue disorders, such as obesity, are associated with a chronic low-grade inflammation predisposing to the development of insulin-resistance, type 2 diabetes and cardiovascular complications. In obesity, accumulation of visceral adipose tissue, rather than subcutaneous adipose tissue, is regarded as the most critical factor contributing to the pathogenesis of these metabolic diseases. Recently has emerged the notion that inflammatory response accompanying obesity corresponds to a cytokine-mediated activation of innate immunity.

The purpose of this review is to provide an update on this emerging concept and to show the reader how innate immune metabolic pathways engaged within white adipose tissue could interfere with innate inflammatory immune defense. First, adipose tissue is reported as an important *in vivo* source of inflammatory cytokines and adipocytes express some receptors of the innate immune system (namely the Toll-like receptors). Second, both innate and adaptive immune cells (respectively, macrophages, dendritic-like cells and T-lymphocytes) appear more and more essential to the initiation and the development of adipose tissue inflammation. More specifically, adipose tissue macrophages have recently emerged as key players in the inflammatory process of obese adipose tissue. Their number and their phenotypic switch from a non inflammatory (i.e. M2) to an inflammatory (i.e. M1) state are likely crucial in the onset of obese adipose tissue inflammation and in the development of insulin-resistance. Finally, the hormonal regulation of adipose tissue inflammation is exemplified by recent data regarding the role of glucocorticoids, both at the level of adipose cells and macrophages.

Altogether, adipose tissue might therefore be regarded as a true immune organ, at the crossroad between metabolism and immune system.

1. The Adipose Tissue: At the Crossroad between Innate and Adaptive Immune Systems

The apparent simplicity of white adipose tissue (WAT), histologically and metabolically, is the main reason why it has relatively been ignored for a long time. Indeed, the primary function of WAT is to store energy in the form of triglycerides during periods of energy excess and to release energy during fasting or starvation as free fatty acids and glycerol. This simplicity is, however, illusory. At the cellular level, there is some heterogeneity in white adipose tissue, with approximately one third of lipid-filled cells (i.e. mature adipocytes) whereas the remaining 2/3 is mostly composed of stromal vascular cells (i.e. fibroblasts and adipocyte precursors) associated with endothelial cells, nerves, and immune cells such as macrophages (*cf II. 1*) and, as recently shown, dendritic-like cells (*cf II. 2*) and lymphocytes (*cf II. 3*). In the 1990's, the demonstration that, in addition to releasing fatty acids during fasting, adipocytes also secreted the pro-inflammatory cytokine tumor necrosis factor α (TNF α)¹ and the hormone leptin,² the latter playing a key role in the regulation of energy metabolism, increased the complexity of this tissue.

Thereby, adipose tissue has lately switched from being a passive and silent energy “reservoir” to representing a complex, highly active and essential metabolic and endocrine organ, secreting an assortment of hormones, cytokines, chemokines and growth factors which regulate whole-body metabolism and immune function.

Below, we will sum up some critical studies demonstrating that adipose tissue might be regarded as a new member of the immune system (*cf I*) and that the inflammatory process, which is an essential early event in the development of obesity, starts and develops within the adipose tissue thanks to the cells of both the innate and adaptive immune systems (*cf II*).

1.1. Adipose tissue as a source of inflammatory cytokines

As stated above, the identification of leptin in 1994² led to the recognition that WAT is an important endocrine secretory organ. Indeed, white adipocytes secrete a multiplicity of factors termed “adipokines”,

Correspondence: Isabelle Wolowczuk, Tel: 00 33 (0)3 20 87 11 59; Email: Isabelle.wolowczuk@ibl.fr



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highly diverse in terms of both structure and function (reviewed in³). These factors encompass cytokines (e.g. TNF α , interleukin-6 (IL-6)), chemokines (e.g. monocyte chemoattractant protein-1 (MCP-1)), proteins of the alternative complement system (e.g. adipsin), and a series of proteins involved in processes ranging from regulation of blood pressure (e.g. angiotensinogen), to glucose homeostasis (e.g. leptin, adiponectin) and angiogenesis (e.g. vascular endothelial growth factor).

Importantly, several adipokines are linked to inflammation and immune response.³ The inflammation-related adipokines include cytokines (e.g. TNF α , IL-1 β , -6, -8, -10, -18, transforming growth factor β (TGF β) and macrophage migration inhibitory factor), chemokines (MCP-1), and acute phase proteins. In addition, the two major adipocyte hormones, leptin and adiponectin, were shown to exert, respectively, pro- and anti-inflammatory actions.⁴⁻⁶

TNF α and IL-6 are the best-studied adipocyte-derived pro-inflammatory factors, both are increased in adipose tissue with obesity.^{7,8} TNF α was the first inflammatory cytokine shown to be produced by adipocytes,¹ even though adipose tissue macrophages have been later identified as being the main cell source in this tissue⁹ (*cf* II. I). TNF α level is increased in adipose tissue and plasma of obese patients and has been related to obesity-associated complications.^{7,8} The involvement of TNF α in insulin-resistance probably results from its multiple direct effects on adipocytes, ranging from alteration of adipocyte differentiation, metabolism, insulin sensitivity and endocrine function (reviewed in^{10,11}). Indeed, TNF α inhibits the transcription of many mature adipocyte-specific genes, such as those involved in glucose uptake (e.g. glucose transporter-4), insulin responsiveness (e.g. insulin receptor and insulin receptor substrate-1),¹²⁻¹⁴ and lipogenesis (e.g. lipoprotein lipase).^{15,14} NF- κ B activation is necessary for TNF α -induced repression of many adipocyte-specific genes.¹⁴ Importantly, the expression level of the nuclear factor peroxisome proliferator-activated receptor γ (PPAR γ), which is necessary to maintain mature adipocyte phenotype, is down-regulated by TNF α exposure.¹⁴ TNF α also stimulates lipolysis¹⁶ *via* various mechanisms. Overall, TNF α reduces adipocyte capacity for triglyceride storage and promotes adipocyte insulin resistance. Indeed, beside its impact on adipocyte gene transcription, TNF α has also been shown to

negatively interfere with the insulin signaling pathway.¹⁷ The cytokine also down-regulate the mRNA level of adiponectin,^{13,14} an adipocyte-derived hormone which contributes to the maintenance of peripheral glucose and lipid homeostasis. Moreover, TNF α inhibits the conversion of pre-adipocytes to mature adipocytes, allowing further recruitment of uncommitted cells and thus possible expansion of adipose tissue mass.¹⁸ Nevertheless, its influence on immune response mostly results from its enhancing effect on the production of other cytokines, such as IL-6, rather than from a direct effect.

Like TNF α , the levels of the other major inflammatory cytokine IL-6 correlate with body mass index.^{8,19} One third of circulating IL-6 is produced by adipose tissue, with visceral WAT producing more IL-6 than subcutaneous WAT. In fat tissue, only a fraction (estimated to ~10%) of IL-6 is secreted by adipocytes, the other part being produced by other cells, particularly macrophages.²⁰ *In vitro*, IL-6 production by adipocytes is strongly increased by TNF α .²¹ The respective role of TNF α and IL-6 produced by adipocytes and macrophages in WAT and during obesity-related inflammation, is difficult to estimate precisely. Nevertheless, as we will describe in section 2, both cells function in a coordinated manner, and macrophage recruitment in WAT is largely attributable to factors secreted by adipocytes, such as MCP-1.²²

The circulating levels of leptin and adiponectin, two hormones predominantly secreted by adipocytes, are respectively increased²³ and decreased²⁴ in obesity. Interestingly, these factors were shown to exert opposite effects on inflammation and on immune response. Leptin was initially described as an adipostat signal, secreted in proportion to adipose mass and controlling appetite and body weight in both humans and rodents.²⁵ The importance of leptin in immunity (reviewed in²⁶) was first revealed in obese mice with homozygous mutation in leptin (*ob/ob*) or leptin receptor (*db/db*), in which impaired immune responses were evidenced.²⁵⁻²⁸ Our group, as well as others, has recently further clarified these immune dysfunctions by demonstrating that obese condition is associated with impaired functionality of T-lymphocytes, dendritic cells and macrophages,²⁹⁻³¹ cells of respectively, adaptive and innate immune systems. The demonstration of increased leptinemia during infection and inflammation further reinforced the role of leptin in inflammation and immunity.³²

Most if not all cells involved in innate immunity express leptin receptor at their surface and thereby are sensitive to leptin (reviewed in²⁵). Leptin induces activation of phagocytosis by monocytes/macrophages and their production of nitric oxide and pro-inflammatory cytokines; induces the chemotaxis of neutrophils and the release of oxygen radicals; up-regulates natural killer (NK) cell-function including proliferation and cytotoxic activity by improving the expression of IL-2 and perforin; and stimulates the production of growth hormone by blood mononuclear cells, allowing survival and proliferation of immune cells. Leptin also modulates adaptive immune responses by interfering in the proliferation of certain sub-populations of T-cells: leptin increases the proliferation of naive CD4⁺ T-cells, whilst inhibiting the proliferation of memory CD4⁺ T-cells. In addition, leptin promotes T helper (Th)1- and suppresses Th2- type response (reviewed in²⁵).

In opposite to leptin, adiponectin exhibits potent immunosuppressive and anti-inflammatory properties.⁶ Adiponectin impairs the production of TNF α , IL-6, IL-8 and interferon γ (IFN γ) by activated macrophages while inducing the production of anti-inflammatory mediators IL-10 and IL-1 receptor antagonist (IL-1Ra) by monocytes/macrophages and dendritic cells. The inhibition of nuclear factor kB (NF-kB) by adiponectin might explain some of these effects.³³ Interestingly, adiponectin modulates endothelial inflammatory responses by reducing the induction of adhesion molecules (ICAM-1, VCAM-1 and E-selectin), and therefore interferes with the adherence of monocytes to endothelial cells and their subsequent migration to sub-endothelial space.³⁴ This effect, added to its inhibitory capacity on transformation of macrophages to foam cells and on proliferation and migration of smooth muscle cells, has led to consider adiponectin as a potential anti-atherogenic factor. Adiponectin is the most abundant adipokine secreted by adipocytes and is present in high concentrations in the blood of healthy subjects (in the range of $\mu\text{g/ml}$ versus ng/ml for leptin), thus weakening the impact of limited vascular injury. Although produced by adipocytes, its serum level significantly drops in obese patients,²⁴ and this was correlated with the development of cardiovascular disease observed in obesity.

In conclusion, adipose tissue might be regarded as an important *in vivo* source of inflammatory products (e.g. TNF α , IL-6, leptin and adiponectin)

and could therefore actively participate to the initiation and the regulation of immune response and inflammation.

Moreover, adipocytes can be regarded as true innate immune cells: indeed, they expressed some receptors of innate immune system, which could enable them to both sense and respond specifically to any danger signals.

1.2. Receptors of the innate immune system are expressed on adipocytes: An emphasis towards the Toll-like receptors

The innate immune system is the body's first line of defense against microbial, chemical and physical injury, whereby various reactions repair damage, avoid or isolate threats and restore homeostasis. In vertebrates, innate immunity is dependent in large part on myeloid cells that include mononuclear phagocytes, macrophages deriving from blood monocytes, and polymorphonuclear phagocytes. Sentinel trouble-shooting macrophages, as well as other immune cell-types, detect environmental threats through pattern-recognition receptors (PRRs) and release pro-inflammatory cytokines like IL-6 and TNF α .^{35,36}

To date, the best characterized PRRs are Toll-like receptors (TLRs), a family of transmembrane receptors that is remarkably conserved from plants to vertebrates.³⁷ TLRs are broadly expressed in the cells of innate immune system such as macrophages and dendritic cells, but also in epithelial and endothelial cells and in organ parenchyma cells and TLRs have therefore specific roles in local innate immune defense.³⁸ Furthermore, the two major cell-types of adaptive immune response, i.e. T- or B-lymphocytes, express certain TLRs and respond directly to corresponding ligands in concert with triggering, respectively, T-cell and B-cell receptors. Thus, in addition to their well-described role in innate immunity, TLRs are also crucial in shaping adaptive immune response from its initiation to the development of immunological memory.³⁹

Interestingly, in addition to their role in innate and adaptive immunity, TLRs have recently been described to regulate bodily energy metabolism, mostly through acting on adipose tissue. Indeed, it was reported that TLR4 (sensing lipopolysaccharide (LPS) and saturated fatty acids) is expressed in the murine pre-adipose cell line 3T3-L1.⁴⁰ Interestingly, LPS-treated adipose cells secrete increased amounts

of TNF α , and subsequently express higher levels of TLR2 (sensing bacterial lipoproteins). Recently, the presence of functional TLR2 and TLR4 was reported on human adipocytes isolated from subcutaneous fat tissue,⁴¹ and several TLRs (TLR1 to 9) were found on mouse adipocytes.^{42,43} The activation of adipocytes *via* TLRs (mostly TLR4) results in the synthesis of pro-inflammatory factors such as TNF α or IL-6, and of chemokines such as MCP-1 (also known as CCL2), CCL5 or CCL11.^{40,41,44} Conversely, adipocyte-specific knockdown of TLR4 (e.g. shRNAi for TLR4 in 3T3-L1 cells; or adipocytes from TLR4-deficient mouse) prevented LPS-induced cytokine expression. Finally, adipocytes isolated from diet-induced obese mice or genetically obese animals (*ob/ob* or *db/db* mice) exhibited increased TLR expression,^{43,45,46} together with higher inflammatory cytokine production upon stimulation.⁴³ Of note, increased endotoxemia was observed in mice on high fat-feeding. Moreover, metabolic endotoxemia induced by a continuous LPS infusion had comparable effect on mouse body weight and glucose parameters (e.g. glycemia and insulinemia) to that of high-fat diet.⁴⁷ Mice genetically deficient in TLR4 or in CD14 (a co-receptor for TLR4) were reported to be of “ideal body type”: when fed with a chow diet, these mice exhibit increased bone mineral content, density and size, as well as decreased body fat.⁴⁸ Moreover, these mice do not become obese with age, unlike many strains of laboratory wild-type mice. This “perfect” phenotype of low adiposity and strong bones, with normal activity and fertility was baptized as the “Adonis phenotype” and this concept is currently further explored for its potential in the treatment of obesity.

However, this approach has to be considered with caution since contradictory results have been more recently obtained with high-fat-fed TLR4-deficient mice. Indeed, whilst some reports described no effect on body weight,^{49–51} other authors described increased body weight⁴⁵ or, in contrast, protection against diet-induced obesity.⁵² Similarly, adiposity and food intake were either reported to be unchanged, increased or decreased in TLR4-deficient animals.^{45,49–52} These divergent phenotypes could derive from the use of different mouse genetic backgrounds, different TLR4 mutation strategies or different feeding protocols (e.g. diet composition and timing). Despite these discrepancies in body weight and adiposity levels, they all revealed a marked improvement in

insulin sensitivity when TLR4 gene was disrupted. Therefore TLR4, which is expressed in most tissues of the body, including the insulin sensitive ones such as adipose tissue, muscle and liver,⁵² appears to be an essential mediator of bodily insulin-resistance.

TLRs are mostly expressed on innate immune cells such as macrophages and, as reported above, on pre-adipocytes and mature adipocytes. Interestingly, it should be mentioned that pre-adipocytes were shown to be able to convert into macrophage-like cells.⁵³ Indeed, adipocytes and macrophages share macrophage-specific antigens and the differentiation and function of both cell-types is controlled by PPAR γ .⁵⁴ It has therefore been suggested that adipocytes and macrophages might be closely related and possibly interconvertible. Even still debated, this possible conversion between adipose cells and macrophages might nevertheless reinforce the view of adipose tissue as an integral part of innate immune system.

Taken together, the expression of functional TLRs on adipocytes classifies adipose tissue as a new member of innate immune system that is able to respond specifically to microbial or physical insults. This concept opens a new and fascinating perspective on a potential role of adipose tissue in host defense.

The second part of the review will show that adipose tissue is also an important site of inflammation and can recruit immune cells. Indeed, obesity and insulin-resistance have been closely associated to a massive infiltration of pro-inflammatory macrophages that initiates and sustains inflammation in obese adipose tissue.

2. Cells from Both Innate and Adaptive Immunity Initiate White Adipose Tissue Inflammation (Fig. 1)

2.1. Macrophages: infiltration and activated state in obese adipose tissue

Distributed throughout the body, macrophages are a fundamental part of the immediate innate defense mechanisms, which can promote specific adaptive immunity by inducing T-cell recruitment and activation. Due to their collaboration with T- and B-cells, mostly through cell-to-cell contacts, macrophages also play an essential role in triggering,

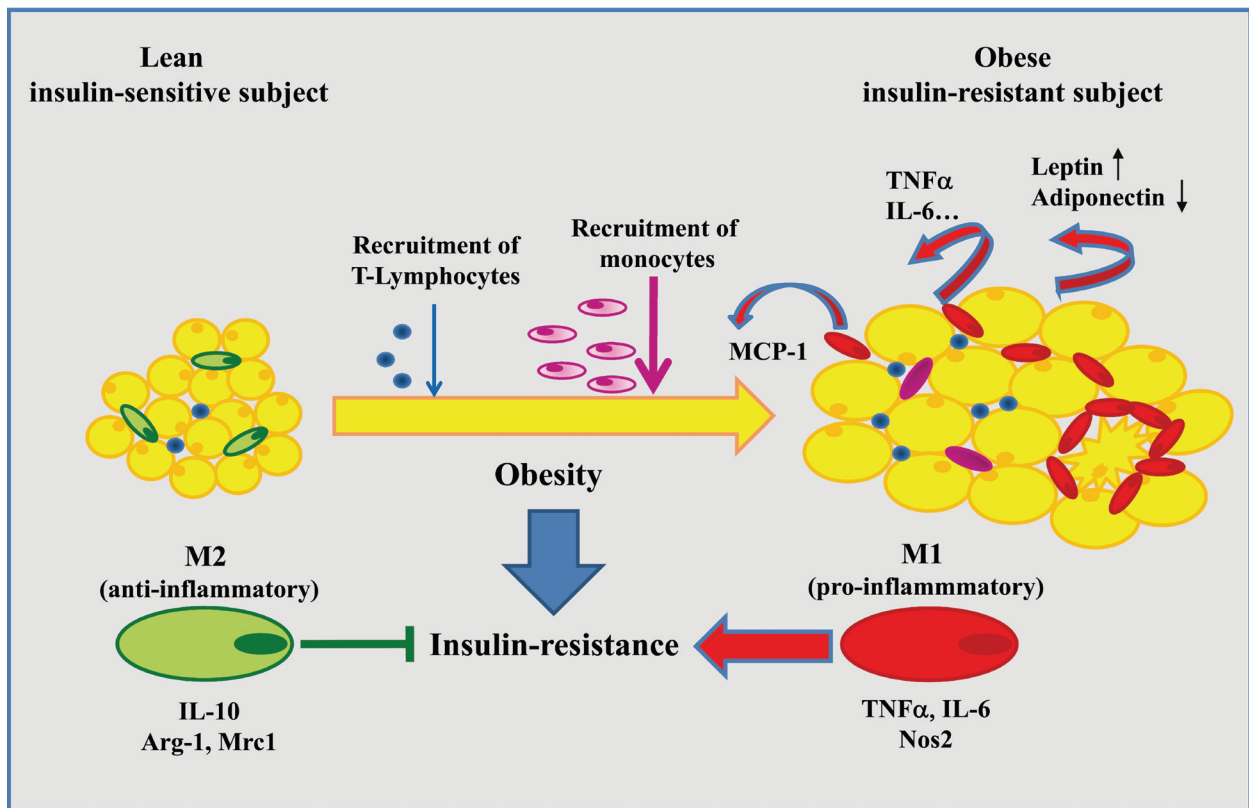


Figure 1. Adipokines and immune cells involved in obesity-induced adipose tissue inflammation and insulin-resistance development. During obesity development, hypertrophic adipocytes release pro-inflammatory factors such as cytokines (TNF α and IL-6), chemokines (MCP-1) and secrete more leptin. These factors, together with the occurrence of adipocyte death, will contribute to enhance T-lymphocytes and monocytes infiltration in the expanding adipose tissue. Macrophages can encircle dead adipocytes forming a « crown-like » structure to scavenge cell debris. Recruited macrophages exacerbate adipose tissue inflammation by releasing pro-inflammatory factors (TNF α , IL-6, MCP-1). These cytokines can worsen adipocyte insulin signaling impairment and could precipitate systemic insulin resistance. As indicated in the lower part of the figure, obesity and insulin-resistance progression are closely linked to a switch of the macrophage activation state: M2-polarized macrophages, which initially reside in adipose tissue, are anti-inflammatory and could partially prevent insulin-sensitivity loss. In the obese insulin-resistant state, M2 macrophages are replaced by M1-polarized macrophages which are pro-inflammatory. Adapted from Zorzanelli Rocha V, Libby P. The multiple facets of the fat tissue. *Thyroid*. 2008;18:175–83.

instructing and terminating the adaptive immune response.

Depending on their tissue localization (lung, brain, liver or adipose tissue) and on the immunological micro-environment, macrophage activation can be either pro- or anti-inflammatory,^{55,56} thereby supporting the activation of the respective T helper (Th) cell-subsets Th1 or Th2, as defined by Mosmann et al.⁵⁷

In adipose tissue, macrophages were shown to play a key role in the development of the inflammatory state associated with obesity. Indeed, macrophages accumulate in the adipose tissues of various obese mouse models (such as the diet-induced obesity or the genetically deficient model in the leptin gene *ob/ob*).^{9,58} The number of macrophages positively correlates with body mass and adipocyte size in both subcutaneous and visceral fat depots, even though macrophage infiltration is more prominent in the

latter. Similar relationships were confirmed in human subcutaneous⁹ and visceral⁵⁹ adipose tissues. Accumulated macrophages are considered to be the critical link between obesity and inflammation since they are the major source of pro-inflammatory cytokine production, notably TNF α and IL-6, in adipose tissues.^{9,58,60} Interestingly, the process appears reversible since macrophage infiltration and pro-inflammatory marker expression in the adipose tissue of obese subjects can be significantly reduced after weight loss.^{9,58,60,61} Similarly to any immune and inflammatory response, macrophage infiltration in expanding adipose tissue results from blood monocytes influx, likely attracted by the chemokine MCP-1.^{9,62} Indeed, it was reported that MCP-1 secretion is markedly enhanced locally and in plasma in obese rodents^{58,22} and human patients. Over-expression, deficiency or mutation-induced dysfunction of MCP-1 in different mouse models

interfere with adipose tissue macrophage (ATM) accumulation along with insulin-resistance development.²² While a pivotal role for MCP-1 and its receptor CCR2 is strongly suggested,⁶³ recent studies have challenged this view by reporting no noticeable impact for the genetic disruption of the chemokine regarding macrophage accumulation and glucose tolerance improvement.^{64,65} Among various potential factors, other chemoattractant cytokines such as osteopontin⁶⁶ and granulocyte-macrophage colony-stimulating factor (GM-CSF)⁶⁷ could also participate in macrophage infiltration during high-fat diet-induced obesity. MCP-1 and osteopontin are rather produced by the various cell types composing the stromal vascular fraction of adipose tissue than by mature adipocytes,^{58,60,68,66} questioning the causal relationship between obesity-associated adipocyte perturbations and macrophage recruitment. Indeed, adipose tissue expansion is not systematically associated with macrophage infiltration and insulin sensitivity impairment. In a mouse model of morbid obesity (consecutive to adiponectin over-secretion in plasma), a massive development of adipose tissue corresponding to adipocyte hyperplasia did not induce macrophage accumulation. The authors suggested that the integrity of adipocyte function may be a more critical determinant of the local inflammation than the increased adipose mass *per se*.⁶⁹ The precise nature of the event priming adipose tissue inflammation definitively remains to be determined even though some hypotheses have already been proposed. For example, it was suggested that leptin may activate endothelial cells to facilitate monocyte diapedesis.⁶² A process of necrosis-like cell death of adipocytes could also induce macrophages recruitment to phagocyte cell debris. Indeed, several immunohistochemical analyses have consistently reported ATM organization into “crown-like” structure around dead adipocytes in adipose tissue of obese rodents^{70,71} and humans.⁶⁰

More recently, some mechanistic insights to explain how macrophages are associated with inflammation in obesity, have emerged from the characterization of ATM activation state. Macrophage population and function have been revealed to be highly heterogeneous and dependent on the surrounding environment, which has led to their characterization and classification following the well-known classification of T-cell activation state into Th1/Th2 sub-types.⁷² Many refer to polarized macrophages as M1 and M2 cells with distinct functions that are elicited in response to the

factors that dominate the inflammatory scene. Typically, macrophages can be distinguished between the M1 phenotype, identified as the pro-inflammatory or “classically”-activated state, secreting various cytokines (e.g. TNF α , IL-6), and the M2 phenotype referred as to the anti-inflammatory or “alternatively”-activated state, which produces IL-10 and TGF β . Cytokines such as IFN γ , secreted by Th1 cells, or IL-4 and IL-13, produced by Th2 cells control the M1/M2 polarity, respectively favouring a classical activation or an alternative activation of macrophages. M1 and M2 activation states are mainly characterized according to distinct gene expression patterns reflecting their inflammatory activities (e.g. inducible nitric oxide synthase Nos2, cyclooxygenase Cox2) or their tissue remodelling and reparation properties (e.g. arginase-1, mannose receptor).

Following a pulse dye labelling of ATM to discriminate newly infiltrated ATM from the resident ATM, it has been shown that recruited ATM during a diet-induced obesity exhibit an inflammatory M1 profile compared to the already settled ATM.⁷³ Moreover, the same group has recently demonstrated that ATM from lean mice retain a gene expression pattern typical of the M2 activated state, while ATM from obese mice are characterized by enhanced expression levels of TNF α and Nos2, both markers of the M1 activated state. This study supports that diet-induced obesity either converts or promotes the replacement of initial M2-polarized ATM by M1-polarized ATM, thereby contributing to the development of insulin-resistance.⁷⁴ Interestingly, fully differentiated 3T3-L1 adipocytes and mouse primary adipocytes were recently shown to release significant amounts of IL-13 and, to a lesser extent, to produce IL-4⁷⁵ suggesting that adipocytes themselves could maintain ATM in the M2 polarity. Furthermore, the M1/M2 balance and intrinsic metabolism of macrophages appear closely intertwined. Indeed, the M2 state programming seems highly dependent on a metabolic switch of macrophages toward fatty acid oxidation: upon IL-4 exposure, bone-marrow-derived macrophages undergo gene expression changes favouring β oxidation and mitochondrial biogenesis; in contrast, oxidative metabolism blockade prevents the IL-4-mediated switch toward an anti-inflammatory phenotype.⁷⁶ A potential relationship between metabolism and macrophage differentiation state has recently been strengthened through the assessment of the role of two isoforms

of PPAR: PPAR γ and δ/β are nuclear transcription factors controlling the expression of genes involved in adipogenesis and fatty acid oxidation/oxidative metabolism.^{77–79} Both PPAR γ and PPAR δ/β are expressed in macrophages where their expression can be respectively induced by IL-4⁷⁶ and IL-13.⁷⁵ Macrophage-specific disruption of the PPAR γ gene⁸⁰ or the PPAR δ/β gene⁷⁵ severely alters the gene expression signature corresponding to the M2 state, promotes M1 polarization and concomitantly worsens insulin sensitivity impairment in mice. The relative contribution of each PPAR isoform as well as the nature of the factors (Th2 cytokines *versus* lipid ligands) triggering PPAR activation in macrophages remain to be further established. However, these studies emphasize that metabolic pathways and their regulating transcriptional factors can modulate the immune functions of macrophages.

In conclusion, ATM are the major source of pro-inflammatory mediators in obese adipose tissue and contribute both to the local and systemic metabolic alterations and to the general inflammatory state. Their number and activation state are likely crucial in the onset of obese tissue inflammation and in the development of insulin-resistance. Nevertheless, it should be stated that most of these recent findings are based on ATM phenotyping in several murine models and their relevance to human obesity and its metabolic complications have not yet been fully addressed. Indeed, ATM from obese patients and mice release the same pro- (TNF α , IL-6, MCP-1) and anti- (IL-10, TGF β) inflammatory factors^{58,74,60,81} but differ with respect to other M1/M2-associated markers such as Nos2 and arginase-1.⁸¹ Key questions are therefore still remaining unanswered such as: What are the mechanisms triggering macrophage intervention and governing the “selection” between M1 or M2 phenotype? Is there coordination between adipocytes and macrophages to regulate their metabolic and immune/inflammatory responses in the context of obesity and its related complications?

2.2. Are dendritic-like cells involved in adipose tissue inflammation?

Recent studies have shown that, besides their classification as M1 or M2-orientated cells (*cf* III), adipose tissue macrophages represent a very heterogeneous population based on the expression of cell-surface markers such as CD11b, CD11c and

F4/80. These findings questioned about the possible participation of dendritic cells (DCs) in the process initiating inflammation in WAT. Indeed, these innate immune cells are the organism's main antigen presenting cells, essential for the induction of any adaptive immune response. A potential role for DCs in adipose tissue inflammation has been first suggested by an increased expression level of the class II major histocompatibility complex (MHCII) genes in newly recruited ATM during high-fat diet feeding.⁷³ Indeed, visceral adipose tissue of different obese mouse models is infiltrated with F4/80⁺CD11c⁺-expressing cells, which are part of the ATM “crown-like” clusters around adipocytes.^{74,82,71} The nature of this sub-population of CD11c⁺-expressing cells remains to be fully clarified. They represent a subset of the F4/80⁺CD11b⁺ population which expresses dendritic cell-specific markers such as CD11c, MHCII and DC-SIGN and have consequently been proposed to be dendritic-like cells.⁸² Surprisingly, the expression of inflammatory markers such as TNF α ⁸² or IL-6 and iNOS,⁷⁴ was shown to be rather restricted to the CD11c⁺ subset than to the CD11b⁺ subset in an obesity context. The participation of dendritic-like cells in adipose tissue inflammation has recently been elegantly addressed using diphtheria toxin-mediated depletion of CD11c⁺ cells in high-fat diet fed mice. This conditional ablation of CD11c⁺ cells drastically prevented ATM accumulation into “crown-like” structures, normalized insulin sensitivity and reduced inflammatory marker expression.⁸³ Even though much has to come regarding the origin, the activating process and the functional role of these dendritic-like cells in adipose tissue, these recent studies clearly stress that these innate immune cells could be crucial in obesity-related inflammation and the concomitant insulin-resistance development.

2.3. Lymphocytes might precede macrophage-infiltration in obese adipose tissue

Whereas T-cells are undoubtedly involved in the regulation of inflammation in atherosclerosis, their role in adipose tissue inflammatory process has just begun to be investigated. Immunohistological and flow cytometry analyses have recently revealed the presence of resident lymphocytes (identified as CD3⁺-T-cells and CD19⁺-B-cells) in mouse visceral and subcutaneous adipose tissues.⁸⁴ Even though first studies failed to detect any change in

T-cell number in adipose tissue of obese mice,^{58,84} several recent studies have pointed out that dietary^{85–87} or genetic¹¹² obesity is also associated with T-cell infiltration including both CD4⁺ and CD8⁺ cells. Immune cell composition assessment at an early stage of high-fat diet-induced obesity suggests that T-cell entry in adipose tissue could precede monocyte attraction and, therefore, might represent one of the processes initiating adipose tissue inflammation.⁸⁶ Indeed, both the secretion of the chemokine CCL5/“Regulated on Activation, Normal T-cell Expressed and Secreted” (RANTES) and the expression level of its receptor CCR5 are enhanced in adipose tissue of obese male mice.⁸⁵ Moreover, RANTES neutralization reduces, *in vitro*, T-cell migration.⁸⁵ Interestingly, RANTES expression was not only restricted to T-cells but was also detected in mature adipocytes, more prominently in presence of TNF α .⁸⁵ If the mechanisms underlying T-cell attraction in adipose tissue are far to be deciphered, even less is known regarding their activation state in the obesity-associated inflammation. However, the implication of the Th1-type cytokine IFN γ in adipose tissue inflammation has recently been explored.⁸⁷ IFN γ mRNA expression was up-regulated in mouse adipose tissue after high-fat diet feeding. 3T3-L1 adipocytes incubated with IFN γ exhibited a marked enhancement of the expression level of various cytokines and chemokines. Moreover, IFN γ deficiency partly prevented the diet-induced increase of both ATM number and pro-inflammatory marker expression (TNF α and MCP-1) in adipose tissue. This phenotype was associated with a slight improvement of glucose intolerance. Overall, this first characterization supports that IFN γ could participate in obesity-associated inflammation.⁸⁷ Eventually, the involvement of other factors known to drive Th1 activation and locally produced in WAT, such as leptin,⁸⁸ would need to be questioned regarding the activation of lymphocytes within the fat tissue.

3. Hormonal Modulation of White Adipose Tissue Inflammation: A Focus on Glucocorticoids

Cortisone is a glucocorticoid (GC) hormone that was first used to treat rheumatoid arthritis in humans in the late 1940s, thereby leading to the discovery of the anti-inflammatory effects of GC. Endogenous GC, however, are rather immunomodulatory

than simply anti-inflammatory. Depending upon concentration and timing, GC either enhance or suppress immune responses, thus shaping both innate and adaptive immunity. Strikingly, prolonged exposure to GC, as seen in Cushing’s syndrome, leads to morphological and metabolic features resembling those of the metabolic syndrome.⁸⁹ Indeed, Cushing patients develop upper body obesity, hypertension and hyperglycemia. Excellent reviews dealing with the effects of GC on metabolism have recently been published.^{90–93} Therefore, considering the scope of our review, we will mainly focus on the modulation of inflammation by GC, likely participating in local inflammation within obese adipose tissue.

During inflammation, GC were reported to promote differentiation and survival of anti-inflammatory macrophages (M2; *cf* II.1) thus promoting the resolution of inflammation.^{94–96} As evoked above, the resulting effects of GC on inflammation depend upon concentration, thus it is essential to note that intracellular GC levels can greatly differ from blood values due to the action of 11 β -hydroxysteroid dehydrogenase (11 β -HSD), an enzyme that converts the intrinsically inert cortisone to the active GC, cortisol. There are two isoenzymes, 11 β -HSD1 and 11 β -HSD2 with opposite activities. The participation of 11 β -HSD1/2 in adipose tissue inflammation has been elegantly investigated using mice over-expressing either isoenzymes under the control of the α P2 promoter. In these animals, expression of the transgene was restricted to brown and white adipose tissues and did not occur in other 11 β -HSD1/2-responsive sites (i.e. brain, liver, skeletal muscles and kidney). 11 β -HSD1 over-expressing mice were hyperphagic and developed abdominal obesity, dyslipidemia, and leptin resistance similar to that observed in human obesity.⁹⁷ Therefore, 11 β -HSD1 transgenic mice exhibited the main features of metabolic syndrome such as glucose intolerance, insulin-resistance (mostly revealed under high-fat feeding), hypertension and a predominantly pro-inflammatory cytokine profile (e.g. increased TNF α and decreased adiponectin). Conversely, mice in which glucocorticoid effects were abolished in adipose tissue (i.e. 11 β -HSD2 transgenic mice), exhibited resistance to both weight gain and fat accumulation, displayed improved responsiveness to glucose and insulin, decreased expression of leptin and increased production of the anti-inflammatory adipose-derived hormone adiponectin.

Importantly, the link between 11 β -HSD1 expression and fat mass has also been reported in human obesity. Both subcutaneous and visceral adipose tissues of obese individuals were shown to express higher levels of 11 β -HSD1 than those of lean subjects.^{98,99} It has been proposed that adipose tissue-specific rise in 11 β -HSD1 in obesity might amplify intracellular glucocorticoid levels, consequently modulating the transcription of several key target genes involved in adipocyte and macrophage functions.

Modulation of WAT inflammation by glucocorticoids is likely not exclusively restricted to an effect on adipocytes. GC could also affect immune cells that are present in adipose tissue, especially macrophages that are essential in WAT inflammatory process (*cf* II, 1). Indeed, 11 β -HSD1 is expressed in murine and human macrophages and its expression level varies according to the macrophage polarization state. In human macrophages, the expression and activity of 11 β -HSD1 were reported to be significantly higher in M1 macrophages than in M2 macrophages.¹⁰⁰ Along those lines, monocyte-derived dendritic cells expressed 11 β -HSD1, this expression being increased upon LPS- or TNF α -induced terminal differentiation.¹⁰¹

Altogether, these recent evidences pointing out the modulatory role of 11 β -HSD1 on inflammation, suggest that the inhibition of 11 β -HSD1, specifically in adipose tissue, could represent a novel therapeutic strategy to treat metabolic diseases such as obesity.

3. Conclusive Remarks

Obesity is a world-wide epidemic currently viewed as a state of chronic, low-grade inflammation characterized by a pro-inflammatory alteration in the serum cytokine profile as well as an infiltration of white adipose tissue by activated macrophages.^{9,59} A better knowledge of how inflammatory pathways are chronically activated is crucial since inflammation undoubtedly contributes to insulin-resistance and type-2 diabetes. Adipose tissue inflammation could exacerbate (notably through TNF α release) adipocyte metabolic and endocrine dysfunctions, both participating to the development of insulin-resistance and type 2 diabetes. Indeed, the impairment of free fatty acid storage in adipose tissue can also be involved in the loss of peripheral insulin sensitivity and insulin secretion: reduced fatty acid uptake and increased lipolysis can lead to inappropriate accumulation of lipids in non-adipose tissues such as liver, skeletal muscles and pancreatic islets,

subsequently leading to defects in insulin action and secretion (reviewed in^{102,103}).

Leptin and adiponectin participate to the control of glucose homeostasis^{104,105} and the secretion level of both adipokines is altered in obesity.^{23,24} Interestingly, the alterations of adipose tissue metabolic and endocrine functions likely participate to the development of insulin-resistance and type 2 diabetes that occurs in lipodystrophic patients who are characterized by a selective loss of adipose tissue (reviewed in^{106,107}). Due to its low incidence and high heterogeneity, the lipodystrophic syndrome has not been as well-explored as obesity and studies regarding the inflammatory status of the remaining fat depots of lipodystrophic patients are currently missing.

It has also to be emphasized that obesity, defined by a high body mass index and an elevated fat mass, is not systematically associated with insulin-resistance, metabolic or cardiovascular complications. As consistently shown, accumulation of visceral fat is more detrimental than accumulation of subcutaneous fat. Moreover, the characterization of different subtypes of obesity has recently led to the identification of a subset of obese, yet highly insulin-sensitive, patients (reviewed in¹⁰⁸). These so-called “metabolically healthy but obese” individuals display less visceral fat mass¹⁰⁹ and lower plasma levels of inflammatory markers (e.g. IL-6¹¹⁰ and C-reactive protein¹¹¹) than unhealthy obese individuals. Further work on the inflammatory state (e.g. macrophage number and activation state, cytokine production) of the adipose tissue of the “metabolically healthy but obese” patients is needed to clearly define whether this sub-population of obese individuals is also protected from adipose tissue inflammation. However, obvious ethical reasons and difficulties to diagnose and “classify” lipodystrophic and obese patients are real limits to such investigations. This partly accounts for the common use of animal models, which have their own limitations due to inherent species- and/or strain-based differences and to difficulties to entirely reproduce the etiology and pathological profiles associated to human adipose tissue disorders.

Beside the hormonal regulation of adipose tissue mass and inflammation (that we exemplified by the exciting results on glucocorticoids), the inciting event that triggers the inflammatory cascade in adipose tissue remains to be elucidated. Yet we reported that a current hypothesis for the accumulation of macrophages within obese white adipose tissue might be the recruitment of T-lymphocytes^{85–87,112}

and possibly of dendritic-like cells,^{74,73,82,81} prior to that of macrophages. Nevertheless macrophages—which accumulate within the adipose tissue of both obese rodents and humans, switch their functional phenotype from M2 to M1,^{74,73} and produce several pro-inflammatory molecules—are believed to significantly modulate this process.

Besides, we also reported that adipocytes express innate receptors such as the Toll-like receptors, mostly TLR4.³⁸ TLR4 is the receptor for bacterial LPS and a key molecular component of innate immune system which function was most expensively studied in macrophages.¹¹³ We described how TLR4 also contributes to insulin-resistance⁵² and how TLR4 stimulation activates pro-inflammatory pathways similar to that encountered in obese tissues.¹¹⁴

Altogether, we attempted in this review to show that adipose tissue is part of innate and adaptive immune systems. Indeed, it produces inflammatory cytokines (TNF α , IL-6), as well as factors regulating monocyte/macrophage function (leptin, adiponectin). Inflamed adipose tissue is massively infiltrated by cells of both innate and adaptive immune system such as macrophages and possibly T-cells and dendritic-like cells. The expression of a broad spectrum of innate immune receptors such as Toll-like receptors on both pre-adipocytes and mature adipocytes, together with a possible conversion of the former into macrophage-like cells,⁵³ collectively establish white adipose tissue as a new member of the immune system and might position this tissue as being “the key road to inflammation”. Adipose tissue is well-spread in the body and could provide immune cells and/or factors which could participate to local inflammation or immune response, as recently proposed for Crohn’s disease.¹¹⁵ Broadly, it opens new and fascinating perspectives on a potential role of adipose tissue in host defense and inflammatory disease.

Acknowledgments

The authors were supported by the Centre National de la Recherche Scientifique (CNRS; to IW), the Institut Pasteur of Lille (to CV) and the Région Nord-Pas de Calais (to SL). We also thank Karl Oulmi (IFR 142) for the Artwork.

Disclosure

The authors report no conflicts of interest.

References

- Hotamisligil GS, Shargill NS, Spiegelman BM. Adipose expression of tumor necrosis factor- α : direct role in obesity-linked insulin resistance. *Science*. 1993;259:87–91.
- Zhang Y, Proenca R, Maffei M, et al. Positional cloning of the mouse obese gene and its human homologue. *Nature*. 1994;372:425–32.
- Tilg H, Moschen AR. Adipocytokines: mediators linking adipose tissue, inflammation and immunity. *Nat Rev Immunol*. 2006;6:772–83.
- Shen J, Sakaida I, Uchida K, et al. Leptin enhances TNF- α production via p38 and JNK MAPK in LPS-stimulated Kupffer cells. *Life Sci*. 2005;77:1502–15.
- Faggioni R, Jones-Carson J, Reed DA, et al. Leptin-deficient (ob/ob) mice are protected from T-cell-mediated hepatotoxicity: role of tumor necrosis factor α and IL-18. *Proc Natl Acad Sci U S A*. 2000;97:2367–72.
- Hu E, Liang P, Spiegelman BM. AdipoQ is a novel adipose-specific gene dysregulated in obesity. *J Biol Chem*. 1996;271:10697–703.
- Hotamisligil GS, Arner P, Caro JF, et al. Increased adipose tissue expression of tumor necrosis factor- α in human obesity and insulin resistance. *J Clin Invest*. 1995;95:2409–15.
- Kern PA, Ranganathan S, Li C, et al. Adipose tissue tumor necrosis factor and interleukin-6 expression in human obesity and insulin resistance. *Am J Physiol Endocrinol Metab*. 2001;280:E745–51.
- Weisberg SP, McCann D, Desai M, et al. Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest*. 2003;112:1796–808.
- Warne JP. Tumour necrosis factor α : a key regulator of adipose tissue mass. *J Endocrinol*. 2003;177:351–5.
- Cawthorn WP, Sethi JK. TNF- α and adipocyte biology. *FEBS Lett*. 2008;582:117–31.
- Stephens JM, Lee J, Pilch PF. Tumor necrosis factor- α -induced insulin resistance in 3T3-L1 adipocytes is accompanied by a loss of insulin receptor substrate-1 and GLUT4 expression without a loss of insulin receptor-mediated signal transduction. *J Biol Chem*. 1997;272:971–6.
- Ruan H, Miles PD, Ladd CM, et al. Profiling gene transcription in vivo reveals adipose tissue as an immediate target of tumor necrosis factor- α : implications for insulin resistance. *Diabetes*. 2002b;51:3176–88.
- Ruan H, Hacohen N, Golub TR, et al. Tumor necrosis factor- α suppresses adipocyte-specific genes and activates expression of preadipocyte genes in 3T3-L1 adipocytes: nuclear factor- κ B activation by TNF- α is obligatory. *Diabetes*. 2002a;51:1319–36.
- Zechner R, Newman TC, Sherry B, et al. Recombinant human cachectin/tumor necrosis factor but not interleukin-1 α downregulates lipoprotein lipase gene expression at the transcriptional level in mouse 3T3-L1 adipocytes. *Mol Cell Biol*. 1988;8:2394–401.
- Green A, Dobias SB, Walters DJ, et al. Tumor necrosis factor increases the rate of lipolysis in primary cultures of adipocytes without altering levels of hormone-sensitive lipase. *Endocrinology*. 1994;134:2581–8.
- Hotamisligil GS, Peraldi P, Budavari A, et al. IRS-1-mediated inhibition of insulin receptor tyrosine kinase activity in TNF- α - and obesity-induced insulin resistance. *Science*. 1996;271:665–8.
- Kras KM, Hausman DB, Martin RJ. Tumor necrosis factor- α stimulates cell proliferation in adipose tissue-derived stromal-vascular cell culture: promotion of adipose tissue expansion by paracrine growth factors. *Obes Res*. 2000;8:186–93.
- Fain JN, Madan AK, Hiler ML, et al. Comparison of the release of adipokines by adipose tissue, adipose tissue matrix, and adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. *Endocrinology*. 2004;145:2273–82.
- Fried SK, Bunkin DA, Greenberg AS. Omental and subcutaneous adipose tissues of obese subjects release interleukin-6: depot difference and regulation by glucocorticoid. *J Clin Endocrinol Metab*. 1998;83:847–50.
- Grunfeld C, Feingold KR. The metabolic effects of tumor necrosis factor and other cytokines. *Biotherapy*. 1991;3:143–58.
- Kanda H, Tateya S, Tamori Y, et al. MCP-1 contributes to macrophage infiltration into adipose tissue, insulin resistance, and hepatic steatosis in obesity. *J Clin Invest*. 2006;116:1494–505.

23. Friedman JM and Halaas JL. Leptin and the regulation of body weight in mammals. *Nature*. 1998;395:763–70.
24. Arita Y, Kihara S, Ouchi N, et al. Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem Biophys Res Commun*. 1999;257:79–83.
25. La Cava A, Matarese G. The weight of leptin in immunity. *Nat Rev Immunol*. 2004;4:371–9.
26. Matarese G, Moschos S, Mantzoros CS. Leptin in immunology. *J Immunol*. 2005;174:3137–42.
27. Otero M, Lago R, Lago F, et al. Leptin, from fat to inflammation: old questions and new insights. *FEBS Lett*. 2005;579:295–301.
28. Faggioni R, Fantuzzi G, Gabay C, et al. Leptin deficiency enhances sensitivity to endotoxin-induced lethality. *Am J Physiol*. 1999;276: R136–42.
29. Papathanassoglou E, El-Haschimi K, Li XC, et al. Leptin receptor expression and signaling in lymphocytes: kinetics during lymphocyte activation, role in lymphocyte survival, and response to high fat diet in mice. *J Immunol*. 2006;176:7745–52.
30. Macia L, Delacre M, Abboud G, et al. Impairment of dendritic cell functionality and steady-state number in obese mice. *J Immunol*. 2006;177:5997–6006.
31. Verwaerde C, Delanoye A, Macia L, et al. Influence of high-fat feeding on both naïve and antigen-experienced T-cell immune response in DO 10.11 mice. *Scand J Immunol*. 2006;64:457–66.
32. Manolakopoulos S, Bethanis S, Liapi C, et al. An assessment of serum leptin levels in patients with chronic viral hepatitis: a prospective study. *BMC Gastroenterol*. 2007;7:17.
33. Ajuwon KM, Spurlock ME. Palmitate activates the NF-kappaB transcription factor and induces IL-6 and TNFalpha expression in 3T3-L1 adipocytes. *J Nutr*. 2005;135:1841–6.
34. Ouchi N, Kihara S, Arita Y, et al. Novel modulator for endothelial adhesion molecules: adipocyte-derived plasma protein adiponectin. *Circulation*. 1999;100:2473–6.
35. Pickup JC. Inflammation and activated innate immunity in the pathogenesis of type 2 diabetes. *Diabetes Care*. 2004;27:813–23.
36. Medzhitov R, Janeway C. Jr. Innate immunity. *N Engl J Med*. 2000;343:338–44.
37. Takeda K, Kaisho T, Akira S. Toll-like receptors. *Annu Rev Immunol*. 2003;21:335–76.
38. Andonegui G, Bonder CS, Green F, et al. Endothelium-derived Toll-like receptor-4 is the key molecule in LPS-induced neutrophil sequestration into lungs. *J Clin Invest*. 2003;111:1011–20.
39. Watts C, Zaru R, Prescott AR, et al. Proximal effects of Toll-like receptor activation in dendritic cells. *Curr Opin Immunol*. 2007;19:73–8.
40. Lin Y, Lee H, Berg AH, et al. The lipopolysaccharide-activated toll-like receptor (TLR)-4 induces synthesis of the closely related receptor TLR-2 in adipocytes. *J Biol Chem*. 2000;275:24255–63.
41. Bes-Houtmann S, Roche R, Hoareau L, et al. Presence of functional TLR2 and TLR4 on human adipocytes. *Histochem Cell Biol*. 2007;127:131–7.
42. Khazen W, M'Bika JP, Collinet M, et al. Differentiation-dependent expression of interferon gamma and toll-like receptor 9 in 3T3-F442A adipocytes. *Biochimie*. 2007;89:669–75.
43. Batra A, Pietsch J, Fedke I, et al. Leptin-dependent toll-like receptor expression and responsiveness in preadipocytes and adipocytes. *Am J Pathol*. 2007;170:1931–41.
44. Poulain-Godefroy O, Froguel P. Preadipocyte response and impairment of differentiation in an inflammatory environment. *Biochem Biophys Res Commun*. 2007;356:662–7.
45. Shi H, Kokoeva MV, Inouye K, et al. TLR4 links innate immunity and fatty acid-induced insulin resistance. *J Clin Invest*. 2006;116: 3015–25.
46. Song MJ, Kim KH, Yoon JM, et al. Activation of Toll-like receptor 4 is associated with insulin resistance in adipocytes. *Biochem Biophys Res Commun*. 2006;346:739–45.
47. Cani PD, Amar J, Iglesias MA, et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes*. 2007;56:1761–72.
48. Johnson GB, Riggs BL, Platt JL. A genetic basis for the “Adonis” phenotype of low adiposity and strong bones. *Faseb J*. 2004;18: 1282–4.
49. Poggi M, Bastelica D, Gual P, et al. C3H/HeJ mice carrying a toll-like receptor 4 mutation are protected against the development of insulin resistance in white adipose tissue in response to a high-fat diet. *Diabetologia*. 2007;50:1267–76.
50. Suganami T, Mieda T, Itoh M, et al. Attenuation of obesity-induced adipose tissue inflammation in C3H/HeJ mice carrying a Toll-like receptor 4 mutation. *Biochem Biophys Res Commun*. 2007;354:45–9.
51. Kim F, Pham M, Luttrell I, et al. Toll-like receptor-4 mediates vascular inflammation and insulin resistance in diet-induced obesity. *Circ Res*. 2007a;100:1589–96.
52. Tsukumo DM, Carvalho-Filho MA, Carnevali JB, et al. Loss-of-function mutation in Toll-like receptor 4 prevents diet-induced obesity and insulin resistance. *Diabetes*. 2007;56:1986–98.
53. Lehrke M, Lazar MA. Inflamed about obesity. *Nat Med*. 2004;10:126–7.
54. Charriere G, Cousin B, Arnaud E, et al. Preadipocyte conversion to macrophage. Evidence of plasticity. *J Biol Chem*. 2003;278:9850–5.
55. Gordon S. Alternative activation of macrophages. *Nat Rev Immunol*. 2003;3:23–35.
56. Mantovani A, Sica A, Sozzani S, et al. The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol*. 2004;25:677–86.
57. Mosmann TR, Cherwinski H, Bond MW, et al. Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. *J Immunol*. 1986;136:2348–57.
58. Xu H, Barnes GT, Yang Q, et al. Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J Clin Invest*. 2003;112:1821–30.
59. Curat CA, Wegner V, Sengenès C, et al. Macrophages in human visceral adipose tissue: increased accumulation in obesity and a source of resistin and visfatin. *Diabetologia*. 2006;49:744–7.
60. Cencello R, Henegar C, Viguier N, et al. Reduction of macrophage infiltration and chemoattractant gene expression changes in white adipose tissue of morbidly obese subjects after surgery-induced weight loss. *Diabetes*. 2005;54:2277–86.
61. Bruun JM, Helge JW, Richelsen B, et al. Diet and exercise reduce low-grade inflammation and macrophage infiltration in adipose tissue but not in skeletal muscle in severely obese subjects. *Am J Physiol Endocrinol Metab*. 2006;290:E961–7.
62. Curat CA, Miranville A, Sengenès C, et al. From blood monocytes to adipose tissue-resident macrophages: induction of diapedesis by human mature adipocytes. *Diabetes*. 2004;53:1285–92.
63. Weisberg SP, Hunter D, Huber R, et al. CCR2 modulates inflammatory and metabolic effects of high-fat feeding. *J Clin Invest*. 2006; 116:115–24.
64. Inouye KE, Shi H, Howard JK, et al. Absence of CC chemokine ligand 2 does not limit obesity-associated infiltration of macrophages into adipose tissue. *Diabetes*. 2007;56:2242–50.
65. Kirk EA, Sagawa ZK, McDonald TO, et al. Macrophage chemoattractant protein-1 deficiency fails to restrain macrophage infiltration into adipose tissue. *Diabetes*. 2008;57:1254–61.
66. Nomiyama T, Perez-Tilve D, Ogawa D, et al. Osteopontin mediates obesity-induced adipose tissue macrophage infiltration and insulin resistance in mice. *J Clin Invest*. 2007;117:2877–88.
67. Kim DH, Sandoval D, Reed J, et al. The role of GM-CSF in adipose tissue inflammation. *Am J Physiol Endocrinol Metab*. 2008;2:2.
68. Zhou HR, Kim EK, Kim H, et al. Obesity-associated mouse adipose stem cell secretion of monocyte chemoattractant protein-1. *Am J Physiol Endocrinol Metab*. 2007;293:E11153–8.
69. Kim JY, van de Wall E, Laplante M, et al. Obesity-associated improvements in metabolic profile through expansion of adipose tissue. *J Clin Invest*. 2007b;117:2621–37.
70. Cinti S, Mitchell G, Barbatelli G, et al. Adipocyte death defines macrophage localization and function in adipose tissue of obese mice and humans. *J Lipid Res*. 2005;46:2347–55.

71. Strissel KJ, Stancheva Z, Miyoshi H, et al. Adipocyte death, adipose tissue remodeling, and obesity complications. *Diabetes*. 2007;56:2910–8.
72. Gordon S. Macrophage heterogeneity and tissue lipids. *J Clin Invest*. 2007;117:89–93.
73. Lumeng CN, Deyoung SM, Bodzin JL, et al. Increased inflammatory properties of adipose tissue macrophages recruited during diet-induced obesity. *Diabetes*. 2007b;56:16–23.
74. Lumeng CN, Bodzin JL, Saltiel AR. Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *J Clin Invest*. 2007a;117:175–84.
75. Kang K, Reilly SM, Karabacak V, et al. Adipocyte-derived Th2 cytokines and myeloid PPARdelta regulate macrophage polarization and insulin sensitivity. *Cell Metab*. 2008;7:485–95.
76. Vats D, Mukundan L, Odegaard JI, et al. Oxidative metabolism and PGC-1beta attenuate macrophage-mediated inflammation. *Cell Metab*. 2006;4:13–24.
77. Brun RP, Kim JB, Hu E, et al. Peroxisome proliferator-activated receptor gamma and the control of adipogenesis. *Curr Opin Lipidol*. 1997;8:212–8.
78. Ferre P. The biology of peroxisome proliferator-activated receptors: relationship with lipid metabolism and insulin sensitivity. *Diabetes*. 2004;53:S43–50.
79. Tontonoz P, Spiegelman BM. Fat and beyond: the diverse biology of PPARgamma. *Annu Rev Biochem*. 2008;77:289–312.
80. Odegaard JI, Ricardo-Gonzalez RR, Goforth MH, et al. Macrophage-specific PPARgamma controls alternative activation and improves insulin resistance. *Nature*. 2007;447:1116–20.
81. Bourlier V, Zakaroff-Girard A, Miranville A, et al. Remodeling phenotype of human subcutaneous adipose tissue macrophages. *Circulation*. 2008;117:806–15. Epub 2008; Jan 28.
82. Nguyen MT, Favelyukis S, Nguyen AK, et al. A subpopulation of macrophages infiltrates hypertrophic adipose tissue and is activated by free fatty acids via Toll-like receptors 2 and 4 and JNK-dependent pathways. *J Biol Chem*. 2007;282:35279–92.
83. Patsouris D, Li PP, Thapar D, et al. Ablation of CD11c-positive cells normalizes insulin sensitivity in obese insulin resistant animals. *Cell Metab*. 2008;8:301–9.
84. Caspar-Bauguil S, Cousin B, Galinier A, et al. Adipose tissues as an ancestral immune organ: site-specific change in obesity. *FEBS Lett*. 2005;579:3487–92.
85. Wu H, Ghosh S, Perrard XD, et al. T-cell accumulation and regulated on activation, normal T-cell expressed and secreted upregulation in adipose tissue in obesity. *Circulation*. 2007;115:1029–38.
86. Kintscher U, Hartge M, Hess K, et al. T-lymphocyte infiltration in visceral adipose tissue: a primary event in adipose tissue inflammation and the development of obesity-mediated insulin resistance. *Arterioscler Thromb Vase Biol*. 2008;28:1304–10.
87. Rocha VZ, Folco EJ, Sukhova G, et al. Interferon- γ , a Th1 Cytokine, Regulates Fat Inflammation. A Role for Adaptive Immunity in Obesity. *Circ Res*. 2008;24:24.
88. Lord GM, Matarese G, Howard JK, et al. Leptin modulates the T-cell immune response and reverses starvation-induced immunosuppression. *Nature*. 1998;394:897–901.
89. Seckl JR, Morton NM, Chapman KE, et al. Glucocorticoids and 11beta-hydroxysteroid dehydrogenase in adipose tissue. *Recent Prog Horm Res*. 2004;59:359–93.
90. Witchel SF, DeFranco DB. Mechanisms of disease: regulation of glucocorticoid and receptor levels-impact on the metabolic syndrome. *Nat Clin Pract Endocrinol Metab*. 2006;2:621–31.
91. Atanasov AG, Odermatt A. Readjusting the glucocorticoid balance: an opportunity for modulators of 11beta-hydroxysteroid dehydrogenase type I activity? *Endocr Metab Immune Disord Drug Targets*. 2007;7:125–40.
92. Mattsson C, Olsson T. Estrogens and glucocorticoid hormones in adipose tissue metabolism. *Curr Med Chem*. 2007;14:2918–24.
93. Tomlinson JW, Stewart PM. Modulation of glucocorticoid action and the treatment of type-2 diabetes. *Best Pract Res Clin Endocrinol Metab*. 2007;21:607–19.
94. Giles KM, Ross K, Rossi AG, et al. Glucocorticoid augmentation of macrophage capacity for phagocytosis of apoptotic cells is associated with reduced p130Cas expression, loss of paxillin/pyk₂ phosphorylation, and high levels of active Rac. *J Immunol*. 2001;167:976–86.
95. Ehrchen J, Steinmuller L, Barczyk K, et al. Glucocorticoids induce differentiation of a specifically activated, anti-inflammatory subtype of human monocytes. *Blood*. 2007;109:1265–74.
96. Varga G, Ehrchen J, Tsianakas A, et al. Glucocorticoids induce an activated, anti-inflammatory monocyte subset in mice that resembles myeloid-derived suppressor cells. *J Leukoc Biol*. 2008;84:644–50.
97. Masuzaki H, Paterson J, Shinyama H, et al. A transgenic model of visceral obesity and the metabolic syndrome. *Science*. 2001;294:2166–70.
98. Desbriere R, Vuaroqueaux V, Achard V, et al. 11beta-hydroxysteroid dehydrogenase type 1 mRNA is increased in both visceral and subcutaneous adipose tissue of obese patients. *Obesity (Silver Spring)*. 2006;14:794–8.
99. Paulsen SK, Pedersen SB, Fisker S, et al. 11Beta-HSD type 1 expression in human adipose tissue: impact of gender, obesity, and fat localization. *Obesity (Silver Spring)*. 2007;15:1954–60.
100. Martinez FO, Gordon S, Locati M, et al. Transcriptional profiling of the human monocyte-to-macrophage differentiation and polarization: new molecules and patterns of gene expression. *J Immunol*. 2006;177:7303–11.
101. Freeman L, Hewison M, Hughes SV, et al. Expression of 11beta-hydroxysteroid dehydrogenase type 1 permits regulation of glucocorticoid bioavailability by human dendritic cells. *Blood*. 2005;106:2042–9.
102. Stumvoll M, Goldstein BJ, van Haeften TW. Type 2 diabetes: principles of pathogenesis and therapy. *Lancet*. 2005;365:1333–46.
103. Kahn SE, Hull RL, Utzschneider KM. Mechanisms linking obesity to insulin resistance and type 2 diabetes. *Nature*. 2006;444:840–6.
104. Schwartz MW, Baskin DG, Bukowski TR, et al. Specificity of leptin action on elevated blood glucose levels and hypothalamic neuropeptide Y gene expression in ob/ob mice. *Diabetes*. 1996;45:531–5.
105. Maeda N, Shimomura I, Kishida K, et al. Diet-induced insulin resistance in mice lacking adiponectin/ACRP30. *Nat Med*. 2002;8:731–7.
106. Garg A, Misra A. Lipodystrophies: rare disorders causing metabolic syndrome. *Endocrinol Metab Clin North Am*. 2004;33:305–31.
107. Agarwal AK, Garg A. Genetic disorders of adipose tissue development, differentiation, and death. *Annu Rev Genomics Hum Genet*. 2006;7:175–99.
108. Karelis AD, St-Pierre DH, Conus F, et al. Metabolic and body composition factors in subgroups of obesity: what do we know? *J Clin Endocrinol Metab*. 2004;89:2569–75.
109. Brochu M, Tchernof A, Dionne IT, et al. What are the physical characteristics associated with a normal metabolic profile despite a high level of obesity in postmenopausal women? *J Clin Endocrinol Metab*. 2001;86:1020–5.
110. Shin MJ, Hyun YJ, Kim OY, et al. Weight loss effect on inflammation and LDL oxidation in metabolically healthy but obese (MHO) individuals: low inflammation and LDL oxidation in MHO women. *Int J Obes (Lond)*. 2006;30:1529–34.
111. Karelis AD, Faraj M, Bastard JP, et al. The metabolically healthy but obese individual presents a favorable inflammation profile. *J Clin Endocrinol Metab*. 2005;90:4145–50.
112. Rausch ME, Weisberg S, Vardhana P, et al. Obesity in C57BL/6J mice is characterized by adipose tissue hypoxia and cytotoxic T-cell infiltration. *Int J Obes (Lond)*. 2008;32:451–63.
113. Fujihara M, Muroi M, Tanamoto K, et al. Molecular mechanisms of macrophage activation and deactivation by lipopolysaccharide: roles of the receptor complex. *Pharmacol Ther*. 2003;100:171–94.
114. Shah PK. Innate immune pathway links obesity to insulin resistance. *Circ Res*. 2007;100:1531–3.
115. Bedford PA, Todorovic V, Westcott ED, et al. Adipose tissue of human omentum is a major source of dendritic cells, which lose MHC Class II and stimulatory function in Crohn's disease. *J Leukoc Biol*. 2006;80:546–54.